



HAL
open science

Dairy starters as "2-in-1" probiotic microorganisms fermenting foods and modulating gut mucosal immunity

Nassima Illikoud, Marine Mantel, Malvyne Rolli-Derkinderen, Valérie Gagnaire, Gwénaél Jan

► To cite this version:

Nassima Illikoud, Marine Mantel, Malvyne Rolli-Derkinderen, Valérie Gagnaire, Gwénaél Jan. Dairy starters as "2-in-1" probiotic microorganisms fermenting foods and modulating gut mucosal immunity. <https://www.beneficialmicrobes2023.org/>. The 10th Beneficial Microbes Conference, Nov 2023, Amsterdam, Netherlands. , 2023. hal-04302652

HAL Id: hal-04302652

<https://hal.inrae.fr/hal-04302652>

Submitted on 23 Nov 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



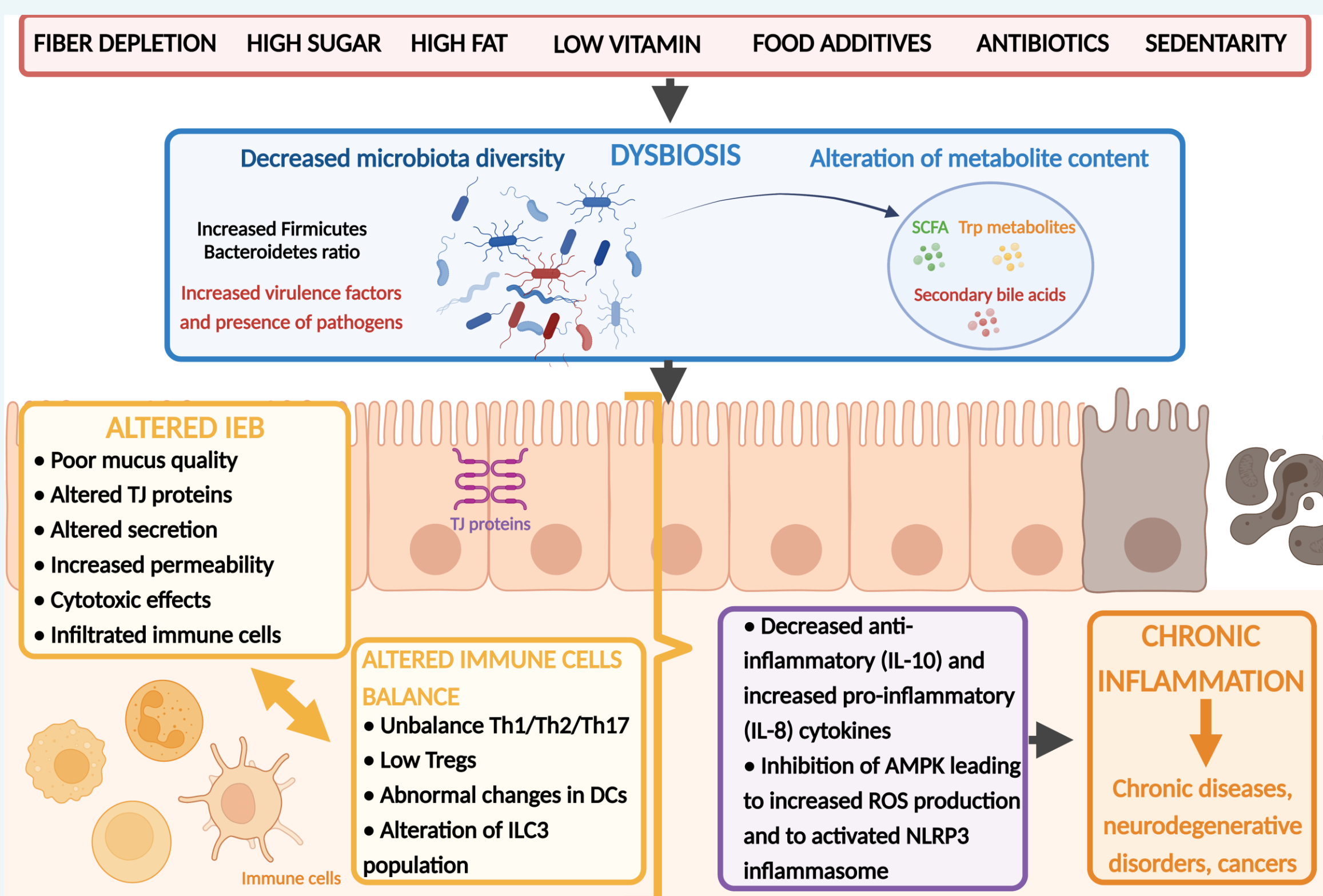
Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License



CONTEXT

✓ The gut microbiota plays a crucial role in the regulation of mucosal immunity and of the function of the intestinal barrier. Dysbiosis is accordingly associated with rupture of mucosal immune homeostasis, leading to inflammatory intestinal diseases. In this context, probiotic bacteria, including a new generation of intestinal probiotics, can maintain intestinal homeostasis and promote health. Surprisingly, little is known about the impact of fermented dairy products in this context, while they represent our main source of live and active bacteria. Indeed, they provide, through our daily diet, a high number of bacteria whose effect on mucosal immunity deserves attention. Among bacteria ingested in fermented dairy products, *Streptococcus thermophilus*, *Lactobacillus delbrueckii*, *Lactobacillus helveticus*, *Lactococcus lactis* and *Propionibacterium freudenreichii* are on top, as they are ingested in high concentrations (close to 10⁹ ufc per gram of product) in fermented milks or cheeses.

THE PROBLEM OF MODERN LIFESTYLE



SCFA: short chain fatty acids; TJ: tight junction; IEC: intestinal epithelial cells; DCs: dendritic cells; ILC3: type 3 innate lymphoid cells; AMPK: Adenosine monophosphate-activated protein kinase; ROS: reactive oxygen species; NLRP3: Nucleotide-binding Oligomerization Domain-like Receptor Protein 3.

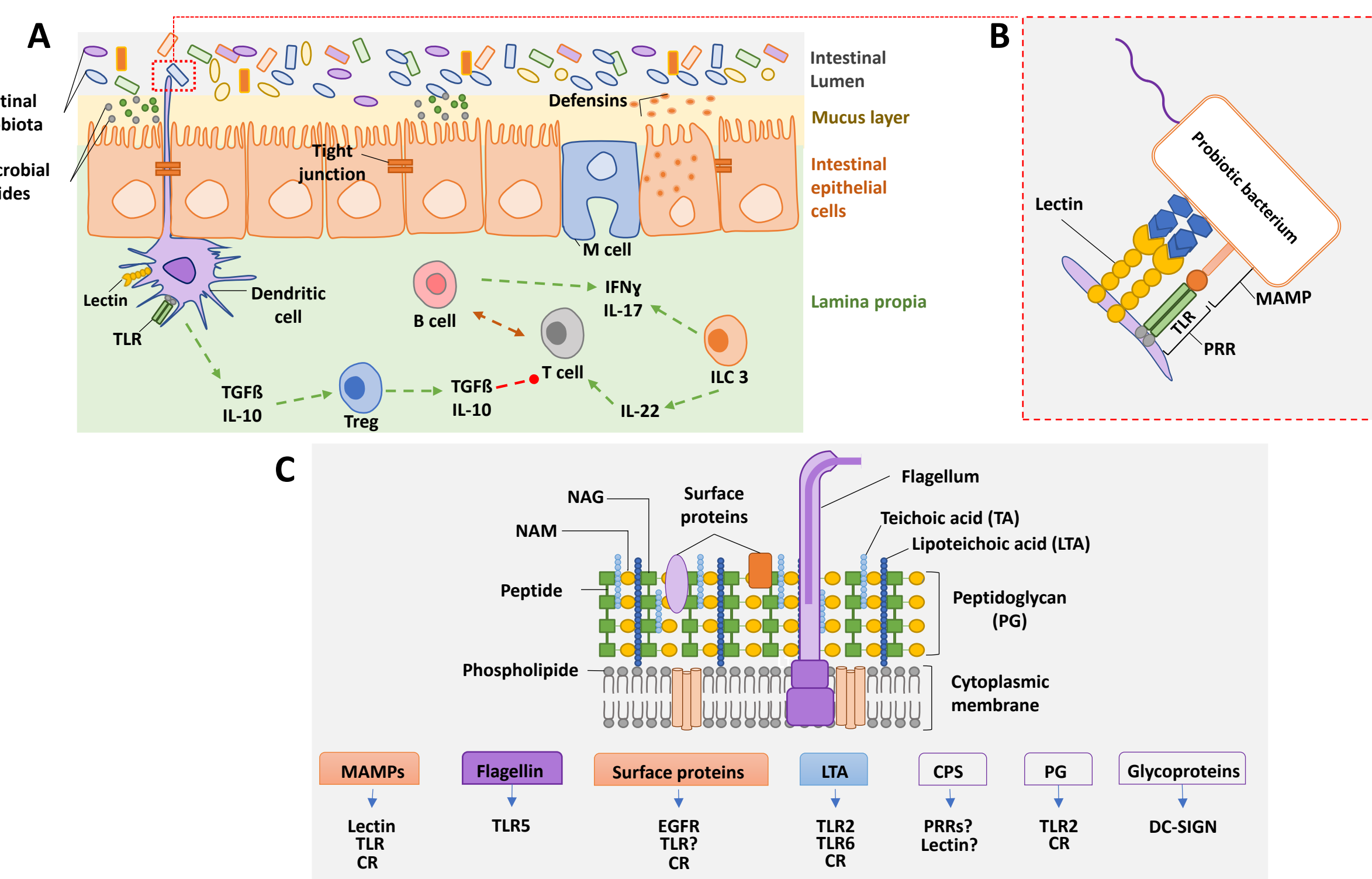
Consequences of changes in modern lifestyle on the gut dysbiosis, intestinal epithelial barrier (IEB) and immune system.

Changes occurring during transition to a modern lifestyle include modification of the eating habits, with reduced intake of fibers, of vitamin, yet enhanced intake of sugar, of food additives, and of fat. These changes also include sedentary and medication, including the use of antibiotics.

They favour dysbiosis, a dysregulation of the gut microbiota, with decreased microbial diversity and alteration of the content in microbial metabolites. This in turn leads to an altered intestinal epithelial barrier, which is involved in an altered balance of immune cells. All these changes finally favour chronic inflammation.

Immunomodulatory bacteria may help

INTERACTIONS BETWEEN INGESTED BACTERIA AND THE HOST



Schematic view of possible interactions between ingested bacteria and the host.

A: Drawing of the key layers of the intestinal epithelial barrier. The intestinal barrier includes: (i) the intestinal lumen with the intestinal microbiota; (ii) the mucus layer that serves as a microbial and biochemical barrier; (iii) the intestinal epithelial cells (IECs) monolayer, maintained by tight junctions, forming a physical barrier; (iv) and the lamina propria which contains different immunological cells that interact together (DCs, T cell, or ILC3). Green arrows represent cytokine secretion or stimulation, brown represents interaction and red represents inhibition.

B: Focus on the interaction between a probiotic bacterium and dendritic cell (DC). Probiotic bacteria interact with DC via microbe-associated molecular patterns (MAMPs) and pattern recognition receptors (PRR).

C: Schematic representation of the main surface components (MAMPs) of Gram-positive probiotic bacteria and the corresponding PRRs in IECs and DCs. Host pattern recognition receptors (PRRs) that recognize probiotic MAMPs include Toll Like Receptors (TLRs), C-type lectin DC specific intercellular adhesion molecule 3-grabbing nonintegrin (DC-SIGN), co-receptors; CPS, cell wall-associated polysaccharide; PG, peptidoglycan; TA, teichoic acid; LTA, lipoteichoic acid; EGFR, epidermal growth factor receptor; NAG, N-acetylglucosamine; NAM, N-acetylmuramic acid.

Examples of immunomodulatory starter bacteria are given below as ID cards

Table 1
Counts of some lactic acid bacteria and ripening flora in fermented dairy products.

Type of fermented milk product at the end of production	Species studied	Count range (log ₁₀ CFU/g product)	Reference
Plain yogurt (n = 5)	<i>S. salivarius</i> subsp. <i>thermophilus</i>	8.20-8.36	(1)
	<i>L. delbrueckii</i>	8.24-8.60	
	<i>C. thermophilus</i>	8.41-8.56	
Liquid yogurt (n = 5)	<i>S. salivarius</i> subsp. <i>thermophilus</i>	8.34-8.59	
	<i>C. thermophilus</i>	8.48-8.53	
Fermented milk (n = 5)	<i>S. salivarius</i> subsp. <i>thermophilus</i>	7.54 ± 0.05	(2)
	<i>Lactobacillus delbrueckii</i>	8.88 ± 0.99	
Commercial Emmental cheese (90% part of France at 30 and 60 d of ripening)	Thermophilic streptococci including <i>C. thermophilus</i>	6.90 ± 0.09	(3)
	Thermophilic lactobacilli including <i>Lactobacillus helveticus</i>	6.4 ± 5.7	
	Propionibacteria including <i>Propionibacterium freudenreichii</i>	6.0 ± 6.9	
Swiss Gruyère-type cheese manufactured with natural whey culture (6-month ripening)	Thermophilic streptococci including <i>C. thermophilus</i>	2.56 ± 0.07	(4)
	Thermophilic lactobacilli including <i>Lactobacillus helveticus</i>	-6.42 ± 0.01	
	Propionibacteria including <i>Propionibacterium freudenreichii</i>	0.02	
Casu cheese (120 d of ripening)	<i>Lactobacillus helveticus</i> ²	6.5 ± 0.5	(5)
	<i>Lactococcus lactis</i>	6.3 ± 0.6	
	<i>Streptococcus thermophilus</i>	6.7 ± 0.9	
Manchego cheese (150 d of ripening)	<i>Propionibacterium freudenreichii</i>	7.4 ± 0.2	(6)
	<i>Lactobacillus helveticus</i>	5.6 ± 0.3	
	Commercial lactococci starter	-	

¹CFU: colony forming units.
² The quantification of *L. helveticus* was performed by quantitative PCR, which specifically targets the *phoA* gene encoding the α-subunit of the phenylalanine-tryptophan synthetase and expressed as log₁₀ copies g⁻¹.

Genus: *Propionibacterium*
Species: *freudenreichii*
Description: rods, Gram+, pleiomorphic, non-pathogenic, food grade, mesophilic, propionic acid bacterium, heterofermentative
Residence: plants, digestive tract, mammary gland and raw milk
Workplace: cheeses
Profession: opening, flavour, texture
Mucosal immunity modulation (in a strain-dependent manner): prevention of colitis and mucositis in mice, may involve production of folate, surface layer proteins, induction of Treg, yet suppression of Th17 responses, modulation of several cytokines, mucus upregulation

Genus: *Streptococcus*
Species: *thermophilus*
Description: cocci, Gram+, in chains, non-pathogenic, food grade, thermophilic, lactic acid bacterium, homofermentative
Residence: mammary gland and raw milk
Workplace: yogurt and cheeses
Profession: acidification, clotting, proteolysis, flavour
Mucosal immunity modulation (in a strain-dependent manner): prevention of colitis and mucositis in mice, may involve production of folate, riboflavin, exopolysaccharides, teichoic acid, suppression of Th17 response, induction of IFN γ , TLR3 stimulation, mucus upregulation

Genus: *Lactobacillus*
Species: *delbrueckii*
Description: rods, Gram+, non-pathogenic, food grade, thermophilic, lactic acid bacterium, homofermentative
Residence: mammary gland and raw milk
Workplace: yogurt and cheeses
Profession: acidification, clotting, proteolysis, flavour
Mucosal immunity modulation (in a strain-dependent manner): prevention of colitis and colitis-associated cancer in mice, may involve surface proteins and exopolysaccharides, expansion of regulatory T cells, modulation of several cytokines

WHAT CLINICAL STUDIES SAY

- ✓ deprivation of fermented foods from the human diet has strong impacts on the human immune system, including a fall in innate immune response (7).
- ✓ In obese women, yogurt consumption decreased serum LPS, lipids, and biomarkers of inflammation and oxidative stress (8).
- ✓ In overweight and obese people, yogurt reduced PBMCs expression of ROR- γ t and serum levels of high-sensitivity C-reactive protein (hs-CRP)(9).
- ✓ In the same category of subjects, yogurt decreased plasma concentration of TNF- α , while increasing soluble TNF- α receptor-1 (s-TNFR-1), indicating an impact on low-grade systemic inflammation (10).
- ✓ consumption of yogurt reduced postprandial inflammation and markers of endotoxin exposure in healthy premenopausal women (11, 12).
- ✓ Yogurt consumption was further associated with lower levels of chronic inflammation, including serum levels of IL-6 and fibrin, in both normal weight and overweight volunteers, in the Framingham Offspring study (13).
- ✓ A randomized control trial evidence that fermented products cheese and sour cream reduced, while non-fermented products increased expression of genes related to lymphocyte activation, cytokine signalling, chemokine signalling, and cell adhesion (14).

CONCLUSION & PERSPECTIVES

- Modern lifestyle endangers gut barrier and favours gut inflammation
- Fermented food products are our main source of live and active bacteria
- Selected strains of fermenting bacteria exert an immunomodulatory effect
- This opens avenues for the development of functional fermented foods

Read More :

(1) Jordano, R. et al. Comparison of Three M17 Media for the Enumeration of *Streptococcus Thermophilus* in Fermented Dairy Products. *J. Food Prot.* 1992, 55, 999-1002. <https://doi.org/10.4315/0362-0288-55.12.999>
(2) Moreno, Y. et al. Viability Assessment of Lactic Acid Bacteria in Commercial Dairy Products Stored at 4 °C Using LIVE/DEAD® BacLight™ Staining and Conventional Plate Counts. *Int. J. Food Sci. Technol.* 2006, 41, 275-280. <https://doi.org/10.1111/j.1365-2621.2005.01060.x>
(3) Thierry, A. et al. Swiss Cheese Ripening: Dynamics of Bacterial Populations and Evolution of the Aqueous Phase Composition for Three Industrial Cheeses. *Le Lait* 1998, 78, 521-542. <https://doi.org/10.1051/lait:1998549>
(4) Moser, A. et al. Population Dynamics of *Lactobacillus helveticus* in Swiss Gruyère-Type Cheese Manufactured with Natural Whey Cultures. *Front. Microbiol.* 2018, 9, <https://doi.org/10.3389/fmicb.2018.00637>
(5) De Freitas, J. et al. In-Depth Dynamic Characterisation of French PDO Cantal Cheese Made from Raw Milk. *Le Lait* 2007, 87, 97-117.
(6) Poveda, J. M. et al. Preliminary Observations on Proteolysis in Manchego Cheese Made with a Defined-Strain Starter Culture and Adjunct Starter (*Lactobacillus Plantarum*) or a Commercial Starter. *Int. Dairy J.* 2003, 13, 169-178. [https://doi.org/10.1016/S0958-6946\(02\)00150-4](https://doi.org/10.1016/S0958-6946(02)00150-4)
(7) Olivares, M. et al. Dietary Deprivation of Fermented Foods Causes a Fall in Innate Immune Response. Lactic Acid Bacteria Can Counteract the Immunological Effect of This Deprivation. *J. Dairy Res.* 2006, 73, 492-498. <https://doi.org/10.1017/S0022299806002068>
(8) Chen, Y. et al. Yogurt Improves Insulin Resistance and Liver Fat in Obese Women with Nonalcoholic Fatty Liver Disease and Metabolic Syndrome: A Randomized Controlled Trial. *Am. J. Clin. Nutr.* 2019, 109, 1611-1619. <https://doi.org/10.1093/ajcn/nwz358>
(9) Zarrabi, M. et al. Effects of Probiotic Yogurt on Fat Distribution and Gene Expression of Proinflammatory Factors in Peripheral Blood Mononuclear Cells in Overweight and Obese People with or without Weight-Loss Diet. *J. Am. Coll. Nutr.* 2014, 33, 417-425. <https://doi.org/10.1080/07315724.2013.874937>
(10) Van Meijl, L. E. C., Mensink, R. P. Effects of Low-Fat Dairy Consumption on Markers of Low-Grade Systemic Inflammation and Endothelial Function in Overweight and Obese Subjects: An Intervention Study. *Br. J. Nutr.* 2010, 104, 1523-1527. <https://doi.org/10.1017/S0007114510002515>
(11) Pei, R. et al. Premenopausal Low-Fat Yogurt Consumption Reduces Postprandial Inflammation and Markers of Endotoxin Exposure in Healthy Premenopausal Women in a Randomized Controlled Trial. *Br. J. Nutr.* 2018, 148, 910-916. <https://doi.org/10.1093/ajcn/nwz046>
(12) Pei, R. et al. Low-Fat Yogurt Consumption Reduces Biomarkers of Chronic Inflammation and Inhibits Markers of Endotoxin Exposure in Healthy Premenopausal Women: A Randomised Controlled Trial. *Br. J. Nutr.* 2017, 118, 1043-1051. <https://doi.org/10.1017/S0007114517000308>
(13) Yuan, M. et al. Yogurt Consumption is Associated with Lower Levels of Chronic Inflammation in the Framingham Offspring Study. *Nutrients* 2021, 13, 506. <https://doi.org/10.3390/nu13020506>
(14) Rundblad, A. et al. Intake of Fermented Dairy Products Induces a Less Pro-Inflammatory Postprandial Peripheral Blood Mononuclear Cell Gene Expression Response than Non-Fermented Dairy Products: A Randomized Controlled Cross-over Trial. *Mol. Nutr. Food Res.* 2020, 64, 2000319. <https://doi.org/10.1002/mnfr.202000319>