



HAL
open science

Assessment of beef sensory attributes and physicochemical characteristics: A comparative study of intermediate versus normal ultimate pH striploin cuts

Iliani Patinho, Cecylyana Leite Cavalcante, Erick Saldaña, Mohammed Gagaoua, Jorge Behrens, Carmen Contreras-Castillo

► To cite this version:

Iliani Patinho, Cecylyana Leite Cavalcante, Erick Saldaña, Mohammed Gagaoua, Jorge Behrens, et al.. Assessment of beef sensory attributes and physicochemical characteristics: A comparative study of intermediate versus normal ultimate pH striploin cuts. Food Research International, 2024, 175, pp.113778. 10.1016/j.foodres.2023.113778 . hal-04309414

HAL Id: hal-04309414

<https://hal.inrae.fr/hal-04309414v1>

Submitted on 4 Dec 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

1 **Assessment of beef sensory attributes and physicochemical characteristics: A**
2 **comparative study of intermediate *versus* normal ultimate pH striploin cuts**

3
4 Iliani Patinho ^{a*}, Cecylyana Leite Cavalcante ^a, Erick Saldaña ^b, Mohammed Gagaoua ^c,
5 Jorge H. Behrens ^d, Carmen J. Contreras-Castillo ^{a*}

6
7 ^a Department of Agri-food Industry, Food and Nutrition, Luiz de Queiroz College of
8 Agriculture, University of Sao Paulo, Piracicaba, SP 13418-900, Brazil

9 ^b Sensory analysis and consumer study group, Escuela Profesional de Ingeniería
10 Agroindustrial, Universidad Nacional de Moquegua, Prolongación Calle Ancash s/n,
11 Moquegua, 18001, Peru.

12 ^c PEGASE, INRAE, Institut Agro, 35590 Saint-Gilles, France

13 ^d Department of Food Science and Nutrition, School of Food Engineering, University of
14 Campinas, SP, Brazil

15
16 *Corresponding authors email: ilianipatinho@usp.br; ccastill@usp.br

35 **Abstract**

36 The quality of beef, defined by key attributes such as texture, flavour, and post-cooking
37 texture, is shaped by various intrinsic and extrinsic factors. This study conducted a detailed
38 examination of Nellore beef, dividing it into two categories based on ultimate pH (pHu)
39 levels: intermediate (pHu \geq 5.8) and normal (pHu $<$ 5.6). A comprehensive approach was
40 taken, involving twenty trained assessors who applied the Optimised Descriptive Profile
41 (ODP) method to evaluate grilled striploin steak samples. In parallel, consumer preferences
42 were measured through a hedonic test and a Check-all-that-apply (CATA) task, involving
43 135 participants. The ODP results revealed that the intermediate pHu samples were
44 significantly juicier ($P < 0.05$) compared to the normal pHu group. The CATA analysis
45 highlighted differences in both intermediate and normal pHu beef, especially in juiciness,
46 a crucial factor for consumer satisfaction. Notably, variations in deoxymyoglobin content
47 linked to ageing were observed, with higher levels on the 3rd day compared to the 28th day,
48 especially in the intermediate pHu samples ($P < 0.05$). Moreover, colour-related aspects
49 such as L^* , b^* , chroma, and oxymyoglobin were significantly influenced ($P < 0.05$) by both
50 the pHu category and ageing duration. Regarding consumer acceptance, the study found
51 no significant difference in perception between the intermediate and normal pHu groups
52 ($P > 0.05$). These findings illuminate the complex interaction between pHu levels, sensory
53 characteristics, and consumer preferences in beef quality, offering valuable insights for
54 both the industry and research community.

55

56

57 **Keywords:** Beef quality, *Longissimus thoracis et lumborum*, Nellore, Consumer
58 acceptability, Sensory evaluation.

59

60

61

62

63

64

65 **1. Introduction**

66 Animal protein, particularly bovine meat, is a fundamental part of the human diet,
67 especially in Western countries where beef is a major source of protein. Consumer
68 preferences for beef vary across different markets and are influenced by diverse factors
69 including (i) cultural and psychological aspects of consumers (Troy & Kerry, 2010), (ii)
70 the sensory properties of the products (Saldaña et al., 2020a), and (iii) marketing
71 considerations such as price, regulation, quality standards, and distribution (Font-i-Furnols
72 & Guerrero, 2014).

73

74 Texture is a key sensory attribute of beef, offering a range of mouthfeel experiences during
75 consumption, such as tenderness, juiciness, and chewiness (Zhu et al., 2021). The aroma
76 and flavour are also critical, arising from chemical compounds like amino acids, peptides,
77 organic acids, and adenine nucleotide metabolism by-products (Spanier et al., 1997). The
78 Maillard reaction and lipid degradation in meat generate various volatile compounds,
79 contributing to the distinctive aromas (Kosowska et al., 2017). Recent research suggests
80 that the chemical properties of muscle fibres (type I and II) vary and significantly influence
81 the formation of volatiles and beef flavour (Li et al., 2023).

82

83 The sensory quality of beef is largely determined by its ultimate pH (pHu), which is linked
84 to the animal's ability to store glycogen in muscles and the differing amounts of
85 mitochondria in muscle fibres (Picard & Gagaoua, 2020; Poletti et al., 2018). Variations in
86 pHu between intermediate and normal beef can result from factors like animal diet,
87 exercise, pre-slaughter stress, and processing conditions, all impacting the final quality,
88 particularly in terms of colour, tenderness, and flavour (Gagaoua et al., 2021a; Loudon et
89 al., 2018; Ponnampalam et al., 2017). This presents a challenge for food scientists and the
90 industry in aligning consumer perception with variations in beef pHu (Warner et al., 2021).

91

92 The meat industry aims to consistently produce high-quality cuts while reducing consumer
93 dissatisfaction (Gagaoua & Picard, 2020). Various sensory evaluation methods have been
94 deployed, including both analytical (trained panels) and holistic (consumer) approaches
95 (Saldaña et al., 2021). According to Aaslyng et al. (2014), a product's sensory profile can
96 be established using the Optimised Descriptive Profile (ODP) by a trained panel, where
97 product attributes are selected and agreed upon by the assessors (Murray et al., 2001).
98 Conversely, affective methods rely on consumer reactions, such as hedonic measures, and

99 can be augmented with techniques like Check-all-that-apply (CATA) questions, a versatile
100 tool for capturing perceptions, feelings, and attitudes (Jaeger et al., 2015).

101 Considering the prominence of beef in Brazilian diets and the preference for this animal
102 protein, this study aimed to develop a comprehensive sensory profile and deepen our
103 understanding of the acceptance and perception of beef cuts within two pHu ranges:
104 intermediate and normal. This innovative study combined the expertise of trained assessors
105 for consistent data, alongside the views of 135 regular beef consumers. Additionally,
106 instrumental measurements taken at 3, 14, and 28 days *post-mortem* were analysed. The
107 hypothesis was that individual reactions to sensory and physicochemical properties of beef
108 could predict consumer perception of beef quality, providing valuable insights to the meat
109 industry about consumer expectations and purchasing decisions.

110

111 **2. Materials and Methods**

112 All procedures and protocols used in this study complied with the Institutional Animal Care
113 and Use Committee Guidelines (protocol 2019/22) and were approved by the committees
114 of the Luiz de Queiroz College of Agriculture at the University of São Paulo. All
115 participants in the sensory tests provided informed consent, in accordance with the form
116 approved by the ethics committee, under CAAE number: 56217222.2.0000.5395.

117

118 *2.1 Animals, treatments, and sampling*

119 Animals were sourced from a commercial meat slaughterhouse under federal inspection,
120 and they were part of a larger experiment involving 104 animals. The normal and
121 intermediate pHu frequencies stood at 76.92% and 16.35%, respectively. From this pool,
122 ten carcasses of Nellore (*Bos Indicus*) bulls, categorised as intermediate (pHu \geq 5.8; n = 5)
123 and normal (pHu $<$ 5.6; n = 5) beef, were selected. The bulls, aged between 18 and 35
124 months, had 2 to 6 permanent incisor teeth and a hot carcass weight of 303 ± 40 kg. Beef
125 samples (~1 g) were excised from the right side of the *Longissimus thoracis* (LT) muscle
126 between the 10th and 11th ribs, aging after 3 days *post-mortem*, for pHu determination. Each
127 animal's LT muscle was sheared perpendicular to the natural muscle fibres, into five steaks
128 (2.0 cm thick), vacuum-packed, and stored at -18 °C for subsequent sensory analysis. For
129 instrumental analysis, the *Longissimus thoracis et lumborum* (striploin cut) muscles were
130 sheared between the 3rd and 6th lumbar vertebrae, portioned into steaks (2.0 cm thick),
131 vacuum-packed, and refrigerated in a chamber (Danic, Model C-EC/U) at 0 ± 2.0 °C without
132 light exposure during the aging period of 3, 14, and 28 days *post-mortem*.

133 2.2 *Sensory analyses*

134 Sensory evaluations were conducted on LT steaks 3 days *post-mortem*, utilising the
135 methodology outlined by Patinho et al. (2019, 2021). For data collection, the Compusense
136 Cloud platform (Compusense Inc., Guelph, Canada) was employed, with the information
137 being recorded on Android tablets (Samsung Galaxy Tab E T560).

138

139 2.3 *Sample preparation*

140 Sensory evaluations of LT steaks, cut into 2.0 cm thick steaks, were carried out 3 days after
141 slaughter. The steaks were grilled on an electric griddle at 200 °C (SSE50, EDANCA, São
142 Paulo, Brazil) until the internal temperature reached 71 °C, monitored using a puncture
143 thermometer. The steaks were flipped every 60 seconds during grilling, a process that
144 lasted about 5 minutes to achieve the desired internal temperature. Post-cooking, each steak
145 was cut into 1 cm³ cubes, as advised by Gomes et al. (2014) and based on preliminary tests.
146 In consumer testing, the steaks were seasoned with salt (1.7% w/w) before grilling (Martins
147 et al., 2021). The samples, approximately 5g each, were then wrapped in aluminium foil,
148 kept warm in an electrically heated display at around 50 °C for 20 minutes, and served to
149 assessors in plastic dishes marked with three-digit codes. Assessors were instructed to
150 cleanse their palates with water between samples.

151

152 2.4 *Optimised Descriptive Profile*

153 The ODP involved candidates including undergraduate and postgraduate students, staff,
154 and faculty from the Department of Agri-food Industry, Food and Nutrition at the
155 University of São Paulo (ESALQ/USP). These individuals were selected for their
156 experience with meat product sensory evaluations, availability, dietary or health
157 constraints, and non-smoking status. The ODP, conducted with 20 trained assessors (age
158 18-65; comprising 57% women, 39% men, and 4% non-binary individuals; all regular beef
159 consumers), spanned six independent one-hour sessions. In the initial session, the ODP
160 methodology was introduced, and a list of attributes from prior studies (Martins et al., 2021;
161 Yang et al., 2021; Gomes et al., 2014) was discussed, leading to the consensus on sixteen
162 descriptors. The second session was dedicated to finalising the attribute order, evaluation
163 techniques, reference samples, and intensity terms defined as "low" or "high". The third
164 session focused on training the assessors in using the Compusense Cloud and the 9-cm
165 unstructured linear scale to measure attribute intensity in each sample. The final three
166 sessions involved the actual evaluations, with each assessor analysing two beef samples

167 per session, one from the intermediate and the other from the normal pHu range, presented
168 monadically following the Latin Square design of Williams (Wakeling & MacFie, 1995).

169

170 2.5 *Consumer testing*

171 For consumer testing, 135 regular beef consumers (age 18-64; 56% women, 43% men, and
172 1% non-binary; 90% with higher education; beef consumption varied from daily to once a
173 week) from the University of Campinas campus rated the samples' overall liking on a 9-
174 point hedonic scale. The scale ranged from (1) "extremely disliked" to (9) "extremely
175 liked". Participants then used a Check-all-that-apply (CATA) task to identify sensory
176 descriptors for each sample. After tasting both samples, they were asked to describe the
177 sensory profile of an ideal beef using the same CATA task, based on the previous study by
178 Martins et al. (2021).

179

180 2.6 *Quality parameters of beef 3 days post-mortem: pHu, composition, drip loss and* 181 *texture analysis*

182

183 2.6.1 *pHu determination*

184 Muscle pHu was determined according to the method by Bendall (1973). Briefly, the LT
185 muscle samples were taken 3 days *post-mortem* and homogenised using an Ultra Turrax
186 (IKA, model T18 basic, Wilmington, NC, USA) with a buffer containing 5mM sodium
187 iodoacetate and 150mM KCl (pH 7.0) at a 1:8 ratio (w/v). The muscle homogenates were
188 centrifuged using an Eppendorf centrifuge (model 5810R, Hamburg, Germany) at
189 13,000×g for 5 minutes at room temperature, equilibrated to 25 °C, and measured using a
190 digital pH meter (Lucadema, model LUCA-210, São José do Rio Preto, Brazil) with
191 automatic temperature compensation and a glass penetration electrode.

192

193 2.6.2 *Proximate composition*

194 The moisture content, crude protein, ash, and total lipids of muscle samples, taken 3 days
195 *post-mortem*, were evaluated (**Table 2**). The moisture content was determined by oven-
196 drying samples at 105 °C until a constant weight was achieved (Association of Official
197 Analytical Chemists [AOAC], 1995). Total nitrogen was measured using the Kjeldahl
198 method (Nx6.25) (ISO 1871:2009) to ascertain the crude protein content, while ash content
199 was determined by calcining organic matter at 550 °C (ISO 936:1998). Total lipids were

200 extracted with hexane using the Soxhlet method (AOAC, 1995). The results were
201 calculated on a wet basis and expressed as a percentage.

202

203 2.6.3 *Drip loss*

204 Drip loss in refrigerated striploin steaks was determined 3 days *post-mortem*.
205 Approximately 50 g of rectangular muscle (30 mm thickness, 60 mm length, 25 mm width)
206 was cut in the direction of the fibre, weighed, placed inside a net, and suspended into air-
207 filled polyethene plastic. After 48 hours at 0±4.0 °C, all samples were weighed again, and
208 the drip loss percentage was calculated according to the method outlined by Honikel
209 (1998), with adaptations from Torres Filho et al. (2017), using the following equation:

210

$$211 \quad \text{Drip loss (\%)} = [(\text{Initial weight} - \text{Final weight})/\text{initial weight}] \times 100 \quad (1)$$

212

213 2.6.4 *Texture profile analysis (TPA)*

214 Before conducting the texture profile analysis (TPA), the *Longissimus thoracis* (LT)
215 muscle samples, taken 3 days *post-mortem* and initially frozen at -18 °C, were thawed at a
216 temperature of 2-4 °C for 24 hours. These were then cooked on an electric griddle until
217 reaching an internal temperature of 71 °C (Martins et al., 2021). The TPA was carried out
218 using a TA-XT texturometer (Stable Micro Systems, Godalming, United Kingdom)
219 equipped with a P-35 probe (long shaft, regular base), adhering to the methods outlined by
220 Jia et al. (2022). For the analysis, the samples were shaped into cylinders using a circular
221 stainless-steel cutter. Each sample yielded five to seven cylinders, each 3.0 cm in diameter.
222 These were then tested at room temperature under specific conditions: a 5.0-second interval
223 between the first and second compressions, a crosshead speed of 1.0 mm/s, a trigger force
224 of 5 g, and a working distance set at 50% strain. The TPA, defined as the peak force needed
225 to compress the sample, mimics the mechanical process of chewing, thus providing data
226 on texture characteristics such as hardness (measured in Newtons), springiness,
227 cohesiveness, chewiness, and gumminess. The average values from five replicates were
228 used for statistical analysis, with the results for hardness specifically expressed in Newtons.

229

230 2.7 *Quality parameters of beef at 3, 14, and 28 days post-mortem: colour, water-* 231 *holding capacity, cooking losses and shear force*

232

233 2.7.1 *Instrumental colour*

234 The instrumental colour measurement of the refrigerated striploin steaks was carried out at
235 3, 14, and 28 days *post-mortem* using a MiniScan XE Plus spectrophotometer® (HunterLab
236 Associates, Virginia, USA). The spectrophotometer was calibrated with a white ceramic
237 plate adjusted to $Y = 93.7$, $x = 0.3160$, and $y = 0.3323$, using an 8 mm diameter
238 measurement area, a 10° observation angle, and A10 illumination. CIELAB coordinates
239 L^* (lightness), a^* (redness), and b^* (yellowness) were obtained by taking 5 readings at
240 random locations on the steak surface without connective tissue, after allowing 30 minutes
241 of oxygenation at 4°C . Chroma (C^*), hue angle (h^*), and the percentages of
242 metmyoglobin, deoxymyoglobin, and oxymyoglobin were calculated using equations
243 defined by American Meat Science Association [AMSA] (2012). The reflectance ratio at
244 630 nm and 580 nm (R630/580) was measured to estimate the stability of meat colour,
245 where a higher ratio indicates lower metmyoglobin accumulation on the beef surface and
246 thus greater colour stability (AMSA, 2012; Canto et al., 2016).

247

248 2.7.2 *Water-holding capacity (WHC)*

249 The water-holding capacity (WHC) of the refrigerated striploin steaks at 3, 14, and 28 days
250 *post-mortem* was determined using the method described by Grau and Hamm (1953),
251 modified by Hoffmann et al. (1982). Meat samples (0.5 g) were weighed on filter paper
252 and pressed with a 5 kg weight for 5 minutes. The WHC result was expressed as a
253 percentage based on the difference between the final and initial weights of the sample.

254

255 2.7.3 *Cooking losses*

256 Cooking losses for refrigerated striploin steaks at 3, 14, and 28 days *post-mortem* were
257 cooked on an electric plate (model SSE50, EDANCA, São Paulo, Brazil) at 200°C until
258 they reached an internal temperature of 71°C . The cooking loss was calculated by
259 subtracting the sample's weight after cooking (weight_2) from its weight before cooking
260 (weight_1) (Anne et al., 2022), and converting the result to a percentage using the following
261 equation:

262

$$263 \quad \text{Cooking loss (\%)} = \frac{\text{weight}_1 \text{ (g)} - \text{weight}_2 \text{ (g)} \times 100}{\text{weight}_1 \text{ (g)}} \quad (2)$$

264

265 2.7.4 *Shear force*

266 The Warner–Bratzler shear force of the refrigerated striploin steaks at 3, 14, and 28 days
267 *post-mortem* was determined according to Shackelford et al. (1991), with modifications.
268 Briefly, the LT steaks were cooked on an electric plate (model SSE50, EDANCA, São
269 Paulo, Brazil) at 200 °C until they reached an internal temperature of 71 °C (Martins et al.,
270 2021). After cooking, the steaks were refrigerated at 0±2.0 °C for 24 h. Using a drill and
271 leaker, between five and seven sections (1.27 cm in diameter) were extracted by cutting
272 parallel to the muscle fibres. Each section was sheared perpendicular to the fibre orientation
273 using a Warner-Bratzler shear blade (HDP/BS) attached to a TA-XT texturometer (Stable
274 Micro Systems, Godalming, United Kingdom) calibrated for a pre-test speed of 2 mm/s,
275 test speed of 2 mm/s, and post-test speed of 10 mm/s. The device was programmed to travel
276 30 mm at the end of the three phases. The maximum force peak recorded during the analysis
277 represented the shear force in Newtons (N) for hardness. The highest and lowest
278 measurements were excluded, and the mean of five sections was used for statistical
279 analysis.

280

281 2.8 *Data analysis*

282 Statistical analyses were carried out using XLSTAT (Student 2023.1.2.1406) and R project
283 version 4.3.1 (R core team, 2023).

284

285 2.8.1 *Optimised Descriptive Profile*

286 The Optimised Descriptive Profile (ODP) data were examined univariately using a
287 Student's t-test to identify potential sensory differences between the pHu ranges at a 5%
288 significance level in R software. Principal Component Analysis was conducted on the
289 correlation matrix from a multivariate standpoint in XLSTAT.

290

291 2.8.2 *Consumer testing*

292 Hedonic data from the meat acceptance tests were analysed using ANOVA, with pHu range
293 as a fixed factor and consumers as a random factor, treating both samples and consumers
294 as sources of variation. The frequency of terms by pH range from the Check-All-That-
295 Apply (CATA) data was compiled and then analysed using Correspondence Analysis (CA).
296 The correlation between liking and sensory attributes was explored through Principal
297 Coordinate Analysis, as recommended by Saldaña et al. (2020b). Additionally, a penalty
298 analysis was conducted to calculate the impact on liking when sensory attributes were

299 marked or unmarked as ideal, following the method of Selani et al. (2022). All these
300 analyses were performed using XLSTAT.

301

302 2.8.3 *Quality parameters of beef 3 days post-mortem*

303 For the instrumental analyses, data related to pH, drip loss, and texture profile analysis
304 (TPA) of the two different pHu ranges of steaks were compared using a Student's t-test in
305 R software to identify significant differences.

306

307 2.8.4 *Quality parameters of beef at 3, 14, and 28 days post-mortem*

308 Instrumental data assessing colour, water-holding capacity, cooking losses, and shear force
309 considered two pHu ranges and three refrigeration periods as sources of variation. ANOVA
310 was then performed at a 5% significance level, with pHu range as a fixed factor and animal
311 as a random factor, using R software. A Principal Component Analysis (PCA) was
312 conducted with combinations of pHu and storage time as individuals (rows) and
313 instrumental data as variables (columns), using the Pearson correlation matrix in XLSTAT.

314

315 3. Results

316

317 3.1 *Optimised Descriptive Profile*

318 The study revealed significant differences in the sensory profile only regarding juiciness
319 (**Table 3**). Specifically, the panel found steaks with intermediate pHu to be juicier ($P < 0.05$)
320 than those with normal pHu. When considering the overall impact of pHu on beef quality,
321 other sensory characteristics appeared similar. Vector size was used to analyse the positive
322 and/or negative relationships between the 16 sensory characteristics and intermediate and
323 normal pHu. Notably, the flavour attributes of roast beef, fat, connective tissue, and blood
324 odour were prominent in steaks within the intermediate pH range. Conversely, the surface
325 brown colour, chewiness, and hardness were more intense in steaks from the normal pHu
326 range (**Fig. 1A;B**).

327

328 3.2 *Consumer testing*

329 The relationship between striploin steak samples from intermediate and normal pHu ranges
330 and the sensory descriptors used by consumers in the CATA task is represented by the first
331 two dimensions of the Correspondence Analysis (CA), which captures 100% of the original
332 information (**Fig. 2A**). Steaks treated with intermediate pHu were characterised by the

333 attributes of blood odour, hardness, fibrousness, surface brown colour, and a meaty beef
334 flavour. In contrast, the sample with a normal pHu was described as having apparent
335 opacity, blood flavour, dryness, and an internal brown colour. Significantly, none of the
336 samples were near the ideal sensory profile, indicating substantial room for improvement
337 in order to satisfy consumers' ideal sensory preferences. It is worth noting that samples
338 with intermediate and normal pHu were perceived similarly, with no significant difference
339 in terms of acceptance ($P > 0.05$) and overall liking scores of 6.93 and 6.63, respectively
340 (data not shown). Overall liking was associated with the attributes of overall juiciness, fat
341 flavour, roast beef aroma, roast beef flavour, overall tenderness, chewiness, and saltiness
342 (**Fig. 2B**), suggesting that these attributes are instrumental in driving consumer liking.
343 Furthermore, it was identified that the characteristics of chewiness, saltiness, overall
344 tenderness, juiciness, aroma, and roast beef flavour were the main drivers of preference for
345 the optimal beef sample (**Fig. 2A**).

346

347 A penalty-lift analysis pinpointed attributes positively and negatively associated with
348 overall liking. Positively influencing acceptance were roast beef flavour, internal red
349 colour, juiciness, chewiness, tenderness, and roast beef aroma (**Fig. 3**). Conversely,
350 hardness, fibrousness, dryness, and apparent opacity were inversely related, indicating their
351 detrimental effect on beef acceptability.

352

353 3.3 *Quality parameters of beef 3 days post-mortem*

354 The results demonstrated that the differences between intermediate and normal pHu did
355 not significantly affect the drip loss or any texture parameters of the beef (**Table 4**).

356

357 3.4 *Quality parameters of beef at 3, 14, and 28 days post-mortem*

358 According to **Table 5**, there was a significant ($P < 0.05$) impact of pHu and ageing time on
359 the L^* , b^* parameters, specifically on chroma, oxymyoglobin, and deoxymyoglobin. Only
360 the ageing time had an effect on the a^* , hue, surface colour stability, metmyoglobin, and
361 water-holding capacity; without interaction between the two variables (pHu x ageing time).
362 However, only in the pHu ranges was a difference seen for shear force. Compared to meat
363 with a normal pHu, meat with an intermediate pHu had a lower shear force value.

364

365 Lastly, data collected for both intermediate and normal pHu samples during the ageing
366 period was subjected to Principal Component Analysis (PCA). 93.21% of the variation in

367 the total could be explained by the first two principal components, according to this
368 analysis. The first dimension was primarily formed by the intermediate pHu (70.96%),
369 whereas the second dimension was primarily formed by the normal pHu (22.25%) (**Fig. 4**).
370 There were noticeable variations in quality metrics between the two pHu ranges. In relation
371 to lightness (L^*), redness (a^*), yellowness (b^*), chroma, hue, oxymyoglobin,
372 metmyoglobin, and shear force for normal pHu, the intermediate pHu was positively
373 correlated with deoxymyoglobin, cooking losses, water-holding capacity, and surface
374 colour stability.

375

376 **4. Discussion**

377

378 *4.1 Optimised Descriptive Profile*

379 Amongst the range of sensory characteristics examined, pHu notably influenced only the
380 juiciness of meat. Meat juiciness, defined as the sensation of moisture released during
381 chewing, is a vital aspect of meat texture. While initial tenderness sensations diminish
382 rapidly, the experience of juiciness persists in various intensities throughout mastication.
383 This study found that meat samples with intermediate pHu were juicier ($P < 0.05$) than
384 those with normal pHu (Table 3), supporting the findings of Grayson et al. (2016) and
385 mirroring results from Yang et al. (2021) for high pHu (dark-cutting) meat.

386 A higher water-holding capacity (WHC), associated with greater juiciness, was noted in
387 meat with intermediate pHu (**Table 5**). Although there wasn't a significant statistical
388 difference in WHC across pHu ranges, a positive correlation with the intermediate pHu
389 group was evident (**Fig. 4**). This phenomenon can be attributed to the isoelectric point (IP)
390 of myofibrillar proteins, like myosin, near pH 5.4, where myofilaments and myofilament
391 spaces have zero net charge. As explained by Zhang et al. (2022), a pH shift above this
392 point results in myofibrillar proteins having a higher net charge, thus increasing
393 myofilament distances and improving WHC. This seems to be the case with intermediate
394 pHu muscle. Conversely, a rapid pH decline leading to low pHu is linked to reduced WHC,
395 likely due to increased surface hydrophobicity and decreased solubility of myofibrillar
396 proteins, impacting the myofilament structure and surface charges (Huff-Lonergan &
397 Lonergan, 2005; Sharedeh et al., 2015). Additionally, a decrease in pH affects the redox
398 stability of myoglobin, accelerating the oxidation of ferrous myoglobin to ferric, increasing
399 surface moisture, and altering the appearance of *post-mortem* muscle (Faustman & Suman,
400 2017). These findings for low/normal pHu are consistent with our study's observations.

401

402 Significantly, an improvement in water-holding capacity was noted during extended ageing
403 (**Table 5**). Changes in muscle structure during this period, such as proteolysis of
404 myofibrillar and cytoskeletal proteins and alterations in collagen, affect ion-protein
405 interactions, thereby increasing the space available for water retention (della Malva et al.,
406 2021).

407

408 The perception of fat and roast beef flavours appears positively influenced by the higher
409 lipid content in intermediate pHu (**Table 2**). The rich flavour of meat is the result of
410 complex interactions among various ingredients during thermal processing (Aaslyng et al.,
411 2014; Troy & Kerry, 2010), likely explaining the noted preference for roast beef flavour in
412 intermediate pHu samples. The differences in fatty acid profiles between pHu ranges
413 significantly impact the cooking process and the formation of aroma and flavour-active
414 compounds (Kerth & Miller, 2015). O'Quinn et al. (2012) observed a correlation between
415 meat flavour and fat content, suggesting that higher fat content in steaks enhances meat
416 flavour intensity. In summary, **Table 3** indicates that all sensory panel evaluators
417 consistently rated the same attributes - taste, colour, and aroma - despite the inherent
418 challenges in achieving a consensus on these aspects.

419

420 4.2 Consumer testing

421 Our study's findings, supported by trained sensory panel evaluations and consumer CATA
422 task results (Ares & Jaeger, 2023), are significant. Both panels characterized normal pHu
423 steaks as dry, and intermediate pHu steaks as tough (**Table 3; Fig. 2A**). This aligns with
424 Miller et al. (2001), who noted that flavour and juiciness significantly influence consumer
425 satisfaction as steak toughness increases (**Fig. 2B**). Key determinants for consumer
426 approval in beef are tenderness, juiciness, and flavour (Duarte et al., 2022). The primary
427 attributes influencing consumer preferences in our study include roast beef aroma, flavour,
428 chewiness, salty taste, overall tenderness, and juiciness (**Fig. 2A**). This aligns with Liu et
429 al. (2022), who highlighted juiciness as a vital factor in meat quality, with the American
430 Meat Science Association (AMSA) stating it accounts for 10% of consumer approval
431 variation (Watson et al., 2008). Future studies should focus on juiciness, especially in
432 under-researched regions like Brazil.

433

434 Concerning overall acceptance (**Fig. 2B**), previous research indicated a preference for
435 tender and juicy beef (Corcoran et al., 2023; Gomes et al., 2014). Lyford et al. (2010) and
436 Malheiros et al. (2022) suggested consumers are willing to pay more for tender steaks.
437 Zebu beef, compared to *Taurus* cattle, often has lower tenderness and marbling, potentially
438 due to feeding regimes and the rate of muscle pH decline (Antonelo et al., 2022). Our
439 findings affirm tenderness as a desirable quality in beef. Gomes et al. (2014) noted that
440 high cooking temperatures, such as grilling, can enhance meat's appearance, flavour, and
441 aroma. However, cooking meat above 70 °C typically results in drier and tougher meat
442 compared to lower temperatures (60 °C) (Obuz & Dikeman, 2003). The Nellore breed (*Bos*
443 *taurus indicus*) and the internal temperature used for cooking (71 °C) in our study,
444 recommended by Brazilian restaurants and AMSA, might have influenced the lack of
445 perceived tenderness in both pHu ranges (**Fig. 2A**).

446

447 Meat colour perception involves more than just brown or red (Corlett et al., 2021; Hopkins,
448 1996). Factors influencing myoglobin's thermal stability and internal colour of cooked
449 meat include the protein's redox state (Suman et al., 2016). Higher muscle pH helps
450 maintain myoglobin in the reduced ferrous state, offering protection against denaturation
451 (Faustman & Suman 2017; Hunt et al., 1999). Additionally, the brown colour on cooked
452 meats' outer surfaces, observed in the intermediate pHu group (**Fig. 2A**), involves Maillard
453 reaction pigments, which are separate from myoglobin pigments (Faustman & Suman
454 2017). The Maillard reaction, a heat-induced reaction between reducing sugars and
455 proteins, occurs more slowly at low pH (Feiner, 2006). Higher pH may increase non-
456 protonated amino acids in meat, enhancing reactivity in the Maillard reaction (Madruga &
457 Mottram, 1995). High pHu contributes to this reaction, leading to more pigments that affect
458 the external colour of beef.

459

460 Our study also shows that taste and aroma are major drivers of consumption and
461 satisfaction in cooked beef. Previous studies highlight the importance of identifying
462 common sensory attributes that satisfy consumer preferences and promote regular beef
463 consumption. The meat industry should focus on efficient technological solutions that
464 balance large-scale production with defined quality parameters, catering to Brazilian
465 consumers' demands for affordable, high-quality products meeting hygienic, nutritional,
466 and ethical standards.

467

468 The penalty-lift analysis suggests flavour and aroma significantly impact perceived sensory
469 quality, supporting the idea that juiciness enhances liking (**Fig. 3**). These factors
470 collectively increase overall liking, as beef taste and juiciness are perceived early in
471 mastication (Djekic et al., 2022), chewing affects juiciness perception (Aaslyng et al.,
472 2003), and meat flavour and aroma are released during this process (Watanabe et al., 2019).
473 Research indicates a negative correlation between beef pH and hardness (Troy & Kerry,
474 2010), leading to increased sensory hardness and dryness and decreased liking.

475

476 4.3 Parameters of beef quality during ageing (3, 14, and 28 days post-mortem)

477 Fresh meat colour intensity is shaped by various intrinsic and extrinsic factors during
478 muscle-to-meat conversion (Gagaoua et al., 2018). The pH value plays a pivotal role in
479 meat colour development, primarily by affecting oxygen consumption and the reducing
480 activity of myoglobin (Ramanathan & Mancini, 2018).

481

482 Important colour parameters like L^* , a^* , b^* , chroma, hue, oxymyoglobin, and
483 metmyoglobin were significantly influenced ($P < 0.05$) by the normal pHu class and ageing
484 period (3, 14, and 28 days) (**Fig. 4**). pHu values explain variations in L^* values, with higher
485 L^* values observed in steaks with normal pHu (**Table 5**). Increased pH levels lead proteins
486 to retain more water, causing muscle fibres to swell and reduce the distance between them,
487 resulting in a darker muscle surface due to decreased light scattering and increased light
488 absorption by myoglobin (Wu et al., 2020). On day three, steaks showed higher L^* values,
489 but after 14 and 28 days, the values were similar (**Table 5**). This supports Canto et al.
490 (2016), who suggested variations in L^* values might be due to higher myoglobin content
491 on day three, as myoglobin concentration in beef negatively correlates with surface
492 luminosity. Studies also observed higher L^* values in *Longissimus thoracis et lumborum*
493 on day 3 during cold storage (Jeong et al., 2009; Hwang et al., 2010).

494

495 No significant difference was found between pHu ranges in terms of redness (a^*),
496 suggesting either no effect of higher pH on colour stability or an insufficient pH increase
497 to cause additional darkening. However, the highest values ($P < 0.05$) at 14 and 28 days of
498 ageing occurred during the ageing period (**Table 5**). According to **Fig. 4**, low a^* values
499 were linked to high deoxymyoglobin content 3 days *post-mortem* in intermediate pHu beef,
500 while high a^* values were directly associated with oxymyoglobin content in beef with
501 normal pHu (Wu et al., 2020). Previous studies reported increased a^* values in *longissimus*

502 steaks during cold storage (Canto et al., 2016; Kim et al., 2009) due to limited oxygen
503 availability for myoglobin in the early *post-mortem* period (Li et al., 2020). The increase
504 of a^* during ageing may suggest increased colour stability and lessened susceptibility to
505 ageing-induced discolouration (della Malva et al., 2021). Stable-coloured *longissimus*
506 steaks exhibited a surfeit of glycolytic enzymes contributing to enhanced colour stability
507 (Suman et al., 2023). This may also elucidate the elevated oxymyoglobin content in the 14
508 and 28 day *post-mortem* samples.

509

510 The a^* value has been suggested as a simple and reliable indicator to predict consumer
511 acceptability of beef colour (Wang et al., 2020). According to Holman et al. (2017), meat
512 colour is considered acceptable when a^* values are equal to or greater than 14.5. In our
513 study, beef samples from both pH ranges at 3, 14, and 28 days displayed a^* values above
514 14.5, indicating likely consumer acceptance. Similarly, the yellowness values (b^*)
515 followed a similar pattern as the a^* values, with higher values ($P < 0.05$) at 14 and 28 days,
516 aligning with the results of Canto et al. (2016), Kim et al. (2009), and McKenna et al.
517 (2005).

518

519 Chroma and hue angles, derived from a^* and b^* values, are influenced by myoglobin
520 content and mitochondrial functions (Li et al., 2020). In our study, elevated chroma values
521 at 14 and 28 days associated with an increase in oxymyoglobin and a decrease in
522 metmyoglobin, consistent with a prior study by Kannan et al. (2001). The hue angle
523 demonstrated a positive association with oxymyoglobin at 14 and 28 days and a negative
524 association with metmyoglobin at 3 days, which corresponds to the study by Lindahl et al.
525 (2001).

526

527 Surface colour stability (R630/580) showed a significant difference related to ageing time
528 ($P < 0.05$), with lower values on day 3 compared to days 14 and 28. This observation of
529 surface colour stability is consistent with our findings on a^* values and supports a previous
530 study that noticed higher R630/580 values in *Longissimus lumborum* muscles during retail
531 exposure on days 5 and 9 (Joseph et al., 2012). McKenna et al. (2005) noted that muscles
532 with robust colour stability, like the *Longissimus lumborum*, tend to have higher b^* values
533 during storage, which aligns with our results. Also, the presence of sarcoplasmic proteins,
534 acting as antioxidants to inhibit lipid and myoglobin oxidation, may contribute to the
535 enhanced colour stability in *longissimus* steaks (Suman et al., 2023).

536

537 In our study, the relative content of deoxymyoglobin was influenced by both the pHu range
538 and ageing time ($P < 0.05$). Steaks with an intermediate pHu had higher deoxymyoglobin
539 content on day 3 than after 28 days of ageing (**Table 5**). This aligns with findings by Li et
540 al. (2020), indicating that meat with limited glycolysis and higher pH impacts the presence
541 of deoxymyoglobin and oxymyoglobin formation by promoting a higher rate of
542 mitochondrial respiration (Gagaoua et al., 2021b). Elevated muscle pH increases
543 mitochondrial respiration capacity, leading to more oxygen consumption by the
544 mitochondria, which in turn causes muscle darkening by reducing myoglobin's oxygen
545 (Kiyimba et al., 2022). Several pathways influence the activity of metmyoglobin reductase
546 (MRA), all relying on NADH, a crucial electron donor. Mitacek et al. (2019) highlighted
547 NADH's key role in reducing metmyoglobin in *post-mortem* muscle. These insights help
548 explain variations in colour stability between beef with normal and intermediate pHu.

549

550 No significant difference in cooking loss was observed during ageing (**Table 5**), though
551 the intermediate pHu group showed higher cooking loss values. This suggests that beef
552 muscles with more moisture undergo greater cooking losses (Jeremiah et al., 2003). Higher
553 fat content in meat corresponds to less bound water and protein (**Table 2**), impacting
554 cooking losses (Ueda et al., 2007). Protein denaturation during cooking, particularly at
555 temperatures above 70 °C, contributes to meat weight loss (Purslow et al., 2016). Our
556 findings agree with Wicklund et al. (2005) and Berger et al. (2018), who reported no
557 significant differences in cooking loss over varying ageing periods and ageing methods,
558 respectively.

559

560 Shear force results aligned with Gomes et al. (2014), showing a negative correlation
561 between normal pHu beef and shear force, as opposed to intermediate pHu beef, which
562 exhibited lower values. However, our study found no significant interaction between pHu
563 and storage period for shear force values. Liu et al. (2022) suggest that higher pH isn't
564 always the sole factor in reduced meat quality. Pre-slaughter stress, even if the final carcass
565 pH is acceptable (Ferguson & Warner, 2008), can increase shear force values (Gruber et
566 al., 2010). Heat treatment can also raise meat shear force, as it causes muscle fibre
567 disruption due to the thermal denaturation of actin and sarcoplasmic proteins at around 70
568 to 80 °C (Christensen et al., 2000). Various factors, including breed, species, diet, animal
569 age, pH rate, and myofibrillar protein breakdown, significantly affect meat tenderization

570 (Ponnampalam et al., 2017). These factors also influence shear force results. The role of μ -
571 calpain in breaking down larger proteins like titin and nebulin early *post-mortem* is thought
572 to delay meat tenderization in normal pHu meat (Lomiwes et al., 2014).

573

574 From a sensory science perspective, tenderness isn't just about the force needed to chew
575 but also involves mouthfeel sensations from moisture and fat. Our study, comparing the
576 pHu group to texture profile at 3 days *post-mortem* and shear force at 3, 14, and 28 days
577 *post-mortem*, found these differences to be non-significant. This is in line with Powell et
578 al. (2011), who found no impact of shear force on consumer ratings across seven muscles.
579 Liu et al. (2022) argue that the lack of significant correlation between shear force and
580 perceived tenderness indicates these are distinct aspects.

581

582 **5. Conclusion**

583 This study provides valuable insights into assessing beef quality attributes and consumer
584 perceptions based on variations in beef muscle pHu levels. We observed that beef with an
585 intermediate pHu showed increased juiciness compared to typical pHu beef. This
586 heightened juiciness is likely due to increased water-holding capacity. These findings align
587 with consumer preference assessments that identified steaks with intermediate pHu as
588 tougher and those with normal pHu as drier. Consumer feedback indicates that attributes
589 like tenderness and juiciness significantly influence overall beef appreciation. Other
590 important factors include roast beef flavour, chewability, saltiness, and juiciness. The study
591 also showed that steaks with intermediate pHu had higher deoxymyoglobin concentrations
592 on the third day compared to the twenty-eighth day of ageing. Instrumental colour
593 measurements, such as L^* , b^* , chroma, and oxymyoglobin, were significantly affected by
594 both pHu category and ageing duration (3, 14, and 28 days). This research enhances our
595 understanding of pHu variations in beef muscle and their impact on sensory attributes and
596 consumer preferences. Further studies with larger sample sizes are essential to support and
597 expand these findings, aligning with consumer expectations for premium-quality beef.

598

599 **Acknowledgements**

600 This work was supported by the São Paulo Research Foundation FAPESP (grant #
601 2017/26667-2; 2019/26026-2).

602

603 **Declarations of competing interest**

604 The authors declare that they have no known competing financial interests or personal
605 relationships that could have appeared to influence the work reported in this paper.

606

607 **References**

608

609 Aaslyng, M. D., Bejerholm, C., Ertbjerg, P., Bertram, H. C., & Andersen, H. J. (2003).
610 Cooking loss and juiciness of pork in relation to raw meat quality and cooking procedure.
611 *Food Quality and Preference*, *14*(4), 277-288. [https://doi.org/10.1016/S0950-](https://doi.org/10.1016/S0950-3293(02)00086-1)

612 [3293\(02\)00086-1](https://doi.org/10.1016/S0950-3293(02)00086-1)

613 Aaslyng, M. D., Meinert, L., Bejerholm, C., & Warner, R. (2014). Sensory assessment of
614 meat. In *Encyclopedia of Meat Sciences* (pp. 272-279). Elsevier Inc.

615 <https://doi.org/10.1016/B978-0-323-85125-1.00042-9>

616 AMSA. (2012). Guidelines for meat color measurement. American Meat Science
617 Association. [http://www.meatscience.org/publications-resources/printedpublicat](http://www.meatscience.org/publications-resources/printedpublications/amsa-meat-color-measurement-guidelines)
618 [ions/amsa-meat-color-measurement-guidelines](http://www.meatscience.org/publications-resources/printedpublications/amsa-meat-color-measurement-guidelines).

619 AMSA American Meat Science Association (2016). Research guidelines for cookery,
620 sensory evaluation, and instrumental tenderness measurements of fresh meat (2 ed,
621 version 1.02). American Meat Science Association, Champaign, IL, USA.

622 Anne, D., Thierry, A., Keisuke, S., & Michiyo, M. (2022). Transformation of highly
623 marbled meats under various cooking processes. *Meat Science*, *189*, Article 108810.
624 <https://doi.org/10.1016/j.meatsci.2022.108810>

625 Antonelo, D. S., Gómez, J. F., Silva, S. L., Beline, M., Zhang, X., Wang, Y., Pavan, B.,
626 Koulicoff, L. A., Rosa, A. F., Goulart, R. S., Li, S., Gerrard, D. E., Suman, S. P., Wes
627 Schilling, M., & Balieiro, J. C. (2022). Proteome basis for the biological variations in
628 color and tenderness of *longissimus thoracis* muscle from beef cattle differing in growth
629 rate and feeding regime. *Food Research International*, *153*, Article 110947.

630 <https://doi.org/10.1016/j.foodres.2022.110947>

631 AOAC. (1995). Official Methods of Analysis. Association of Official Analytical
632 Chemists International. Washington D.C.

633 Ares, G., & Jaeger, S. R. (2023). Check-all-that-apply (CATA) questions with consumers
634 in practice: Experimental considerations and impact on outcome. In *Rapid sensory*
635 *profiling techniques* (pp. 257-280). Woodhead Publishing. [https://doi.org/10.1016/B978-](https://doi.org/10.1016/B978-0-12-821936-2.00013-3)
636 [0-12-821936-2.00013-3](https://doi.org/10.1016/B978-0-12-821936-2.00013-3)

637 Bejerholm, C., Tørngren, M. A., & Aaslyng, M. D. (2014). Cooking of meat. In M., & C.

638 (Eds.), *Encyclopedia of meat sciences* (pp. 370-376) (2nd ed.). Oxford: Academic Press.

639 Bendall, J. R. (1973). *Postmortem* changes in muscle. The structure and function of
640 muscle, 2(Part 1), 243-309.

641 Berger, J., Kim, Y. H. B., Legako, J. F., Martini, S., Lee, J., Ebner, P., & Zuelly, S. M. S.
642 (2018). Dry-aging improves meat quality attributes of grass-fed beef loins. *Meat Science*,
643 *145*, 285-291. <https://doi.org/10.1016/j.meatsci.2018.07.004>

644 Calkins, C. R., & Hodgen, J. M. (2007). A fresh look at meat flavor. *Meat science*, *77(1)*,
645 63-80. <https://doi.org/10.1016/j.meatsci.2007.04.016>

646 Canto, A. C. V. C. S., Costa-Lima, B. R. C., Suman, S. P., Monteiro, M. L. G., Viana, F.
647 M., Salim, A. P. A. A., Mahesh, N. N., Silva, T. J. P., & Conte-Junior, C. A. (2016).
648 Color attributes and oxidative stability of *longissimus lumborum* and *psoas major*
649 muscles from Nellore bulls. *Meat Science*, *121*, 19-26.
650 <https://doi.org/10.1016/j.meatsci.2016.05.015>

651 Christensen, M., Purslow, P. P., & Larsen, L. M. (2000). The effect of cooking
652 temperature on mechanical properties of whole meat, single muscle fibres and perimysial
653 connective tissue. *Meat Science*, *55(3)*, 301-307. [https://doi.org/10.1016/S0309-](https://doi.org/10.1016/S0309-1740(99)00157-6)
654 [1740\(99\)00157-6](https://doi.org/10.1016/S0309-1740(99)00157-6)

655 Corcoran, L. C., Schlich, P., Moloney, A. P., O'Riordan, E., Millar, K., Botinestean, C.,
656 Gallagher, E., O'Sullivan, M. G., & Crofton, E. C. (2023). Comparing consumer liking of
657 beef from three feeding systems using a combination of traditional and temporal liking
658 sensory methods. *Food Research International*, *168*, Article 112747.
659 <https://doi.org/10.1016/j.foodres.2023.112747>

660 Corlett, M. T., Pethick, D. W., Kelman, K. R., Jacob, R. H., & Gardner, G. E. (2021).
661 Consumer perceptions of meat redness were strongly influenced by storage and display
662 times. *Foods*, *10(3)*, Article 540. <https://doi.org/10.3390/foods10030540>

663 Della Malva, A., Gagaoua, M., Santillo, A., De Palo, P., Sevi, A., & Albenzio, M. (2022).
664 First insights about the underlying mechanisms of Martina Franca donkey meat
665 tenderization during aging: A proteomic approach. *Meat Science*, *193*, Article 108925.
666 <https://doi.org/10.1016/j.meatsci.2022.108925>

667 Djekic, I. V., Ilic, J. G., Sołowiej, B. G., Djekić, R. I., & Tomasevic, I. B. (2022).
668 Application of Food Mechanics and Oral Processing in Modelling First Bite of Grilled
669 Meat. *Journal of Food Quality*, *2022*, Article ID 9176628.
670 <https://doi.org/10.1155/2022/9176628>

671 Duarte, T. L., Bolkenov, B., Klopatek, S. C., Oltjen, J. W., King, D. A., Shackelford, S.

672 D., Wheeler, T. L., & Yang, X. (2022). Evaluating the Shelf Life and Sensory Properties
673 of Beef Steaks from Cattle Raised on Different Grass Feeding Systems in the Western
674 United States. *Foods*, *11*(14), Article 2141. <https://doi.org/10.3390/foods11142141>

675 Faustman, C., & Suman, S. P. (2017). The Eating Quality of Meat: I—Color. Lawrie's
676 Meat Science (Eight Edition), 329-356. [https://doi.org/10.1016/B978-0-08-100694-
677 8.00011-X](https://doi.org/10.1016/B978-0-08-100694-8.00011-X)

678 Ferguson, D. M., & Warner, R. D. (2008). Have we underestimated the impact of pre-
679 slaughter stress on meat quality in ruminants? *Meat Science*, *80*(1), 12-19.
680 <https://doi.org/10.1016/j.meatsci.2008.05.004>

681 Feiner, G. (2006). Definitions of terms used in meat science. *Meat Products Handbook*,
682 46-71. <https://doi.org/https://doi.org/10.1533/9781845691721.1.46>

683 Font-i-Furnols, M., & Guerrero, L. (2014). Consumer preference, behavior and
684 perception about meat and meat products: An overview. *Meat science*, *98*(3), 361-371.
685 <https://doi.org/10.1016/j.meatsci.2014.06.025>

686 Gagaoua, M., Micol, D., Picard, B., Terlouw, C. E. M., Moloney, A. P., Juin, H., Meteau,
687 K., Scollan, N., Richardson, I., & Hocquette, J-F. (2016). Inter-laboratory assessment by
688 trained panelists from France and the United Kingdom of beef cooked at two different
689 end-point temperatures. *Meat Science*, *122*, 90-96.
690 <https://doi.org/10.1016/j.meatsci.2016.07.026>

691 Gagaoua, M., Picard, B., & Monteils, V. (2018). Associations among animal, carcass,
692 muscle characteristics, and fresh meat color traits in Charolais cattle. *Meat Science*, *140*,
693 145-156. <https://doi.org/10.1016/j.meatsci.2018.03.004>

694 Gagaoua, M., Terlouw, E. M. C., Mullen, A. M., Franco, D., Warner, R. D., Lorenzo, J.
695 M., Purslow, P. P., Gerrard, D., Hopkins, D. L., Troy, D., & Picard, B. (2021a).
696 Molecular signatures of beef tenderness: Underlying mechanisms based on integromics
697 of protein biomarkers from multi-platform proteomics studies. *Meat Science*, *172*, Article
698 108311. <https://doi.org/10.1016/j.meatsci.2020.108311>

699 Gagaoua, M., Warner, R. D., Purslow, P., Ramanathan, R., Mullen, A. M., López-
700 Pedrouso, M.,... & Terlouw, E. M. C. (2021b). Dark-cutting beef: A brief review and an
701 integromics meta-analysis at the proteome level to decipher the underlying pathways.
702 *Meat Science*, *181*, Article 108611. <https://doi.org/10.1016/j.meatsci.2021.108611>

703 Gomes, C. L., Pflanzler, S. B., Cruz, A. G., De Felício, P. E., & Bolini, H. M. A. (2014).
704 Sensory descriptive profiling and consumer preferences of beef strip loin steaks. *Food*
705 *Research International*, *59*, 76-84. <https://doi.org/10.1016/j.foodres.2014.01.061>

706 Grau, R., & Hamm, R. (1953). Eine einfache methode zur bestimmung der
707 wasserbindung im muskel. *Naturwissenschaften*, *40(1)*, 29-30.

708 Grayson, A. L., Shackelford, S. D., King, D. A., McKeith, R. O., Miller, R. K., &
709 Wheeler, T. L. (2016). The effects of degree of dark cutting on tenderness and sensory
710 attributes of beef. *Journal of Animal Science*, *94(6)*, 2583-2591.
711 <https://doi.org/10.2527/jas.2016-0388>

712 Gruber, S. L., Tatum, J. D., Engle, T. E., Chapman, P. L., Belk, K. E., & Smith, G. C.
713 (2010). Relationships of behavioral and physiological symptoms of preslaughter stress to
714 beef *longissimus* muscle tenderness. *Journal of Animal Science*, *88(3)*, 1148-1159.
715 <https://doi.org/10.2527/jas.2009-2183>

716 Hoffmann, H., Hamm, R., & Bluchel, E. (1982). Neus uber die bestimmung der
717 wasserbindung des nut hielf filter paper premethods. *Fleishwirtschaft*, *62(1)*, 87-94.

718 Holman, B. W., van de Ven, R. J., Mao, Y., Coombs, C. E., & Hopkins, D. L. (2017).
719 Using instrumental (CIE and reflectance) measures to predict consumers' acceptance of
720 beef colour. *Meat Science*, *127*, 57-62. <https://doi.org/10.1016/j.meatsci.2017.01.005>

721 Honikel, K. O. (1998). Reference methods for the assessment of physical characteristics
722 of meat. *Meat science*, *49(4)*, 447-457. [https://doi.org/10.1016/S0309-1740\(98\)00034-5](https://doi.org/10.1016/S0309-1740(98)00034-5)

723 Hopkins, D. L. (1996). Assessment of lamb meat colour. *Meat Focus*
724 *International*, *5(11)*, 400-401.

725 Huff-Lonergan, E., & Lonergan, S. M. (2005). Mechanisms of water-holding capacity of
726 meat: The role of postmortem biochemical and structural changes. *Meat Science*, *71(1)*,
727 194-204. <https://doi.org/10.1016/j.meatsci.2005.04.022>

728 Hunt, M. C., Sørhølm, O., & Slinde, E. (1999). Color and heat denaturation of myoglobin
729 forms in ground beef. *Journal of Food Science*, *64(5)*, 847-851.
730 <https://doi.org/10.1111/j.1365-2621.1999.tb15925.x>

731 Hwang, Y-H., Kim, G-D., Jeong, J-Y., Hur, S-J., & Joo, S-T. (2010). The relationship
732 between muscle fiber characteristics and meat quality traits of highly marbled Hanwoo
733 (Korean native cattle) steers. *Meat Science*, *86(2)*, 456-461.
734 <https://doi.org/10.1016/j.meatsci.2010.05.034>

735 Jaeger, S. R., Beresford, M. K., Paisley, A. G., Antúnez, L., Vidal, L., Cadena, R. S.,
736 Giménez, A., & Ares, G. (2015). Check-all-that-apply (CATA) questions for sensory
737 product characterization by consumers: Investigations into the number of terms used in
738 CATA questions. *Food Quality and Preference*, *42*, 154-164.
739 <https://doi.org/10.1016/j.foodqual.2015.02.003>

740 Jeong, J. Y., Hur, S. J., Yang, H. S., Moon, S. H., Hwang, Y. H., Park, G. B., & Joo, S. T.
741 (2009). Discoloration Characteristics of 3 Major Muscles From Cattle During Cold
742 Storage. *Journal of Food Science*, *74(1)*, C1-C5. [https://doi.org/10.1111/j.1750-](https://doi.org/10.1111/j.1750-3841.2008.00983.x)
743 [3841.2008.00983.x](https://doi.org/10.1111/j.1750-3841.2008.00983.x)

744 Jeremiah, L., Dugan, M., Aalhus, J., & Gibson, L. (2003). Assessment of the chemical
745 and cooking properties of the major beef muscles and muscle groups. *Meat Science*,
746 *65(3)*, 985-992. [https://doi.org/10.1016/S0309-1740\(02\)00308-X](https://doi.org/10.1016/S0309-1740(02)00308-X)

747 Jia, W., Wang, X., Zhang, R., Shi, Q., & Shi, L. (2022). Irradiation role on meat quality
748 induced dynamic molecular transformation: From nutrition to texture. *Food Reviews*
749 *International*, *1*, 1-23. <https://doi.org/10.1080/87559129.2022.2026377>

750 Joseph, P., Suman, S. P., Rentfrow, G., Li, S., & Beach, C. M. (2012). Proteomics of
751 muscle-specific beef color stability. *Journal of Agricultural and Food Chemistry*, *60(12)*,
752 3196-3203. <https://doi.org/10.1021/jf204188v>

753 Kannan, G., Kouakou, B., & Gelaye, S. (2001). Color changes reflecting myoglobin and
754 lipid oxidation in chevon cuts during refrigerated display. *Small Ruminant Research*,
755 *42(1)*, 67-74. [https://doi.org/10.1016/S0921-4488\(01\)00232-2](https://doi.org/10.1016/S0921-4488(01)00232-2)

756 Kerth, C. R., & R. K. Miller. 2015. Beef flavor: A review from chemistry to
757 consumer. *Journal of the Science of Food and Agriculture*, *95*, 2783-2798.
758 <https://doi.org/10.1002/jsfa.7204>

759 King, N. J., & Whyte, R. (2006). Does it look cooked? A review of factors that influence
760 cooked meat color. *Journal of food science*, *71(4)*, R31-R40.
761 <https://doi.org/10.1111/j.1750-3841.2006.00029.x>

762 Kim, Y. H., Keeton, J. T., Smith, S. B., Berghman, L. R., & Savell, J. W. (2009). Role of
763 lactate dehydrogenase in metmyoglobin reduction and color stability of different bovine
764 muscles. *Meat Science*, *83(3)*, 376-382. <https://doi.org/10.1016/j.meatsci.2009.06.009>

765 Kiyimba, F., Hartson, S. D., Rogers, J., Vanoverbeke, D. L., Mafi, G. G., & Ramanathan,
766 R. (2022). Dark-cutting beef mitochondrial proteomic signatures reveal increased
767 biogenesis proteins and bioenergetics capabilities. *Journal of Proteomics*, *265*, Article
768 104637. <https://doi.org/10.1016/j.jprot.2022.104637>

769 Kosowska, M., Majcher, M. A., & Fortuna, T. (2017). Volatile compounds in meat and
770 meat products. *Food Science and Technology (Brazil)*, *37(1)*, 1-7.
771 <https://doi.org/10.1590/1678-457X.08416>

772 Li, C., Wang, D., Xu, W., Gao, F., & Zhou, G. (2013). Effect of final cooked temperature
773 on tenderness, protein solubility and microstructure of duck breast muscle. *LWT - Food*

774 *Science and Technology*, 51(1), 266-274.
775 <https://doi.org/10.1016/j.lwt.2012.10.003>

776 Li, X., Zhang, D., Ijaz, M., Tian, G., Chen, J., & Du, M. (2020). Colour characteristics of
777 beef *longissimus thoracis* during early 72 h postmortem. *Meat Science*, 170, 108245.
778 <https://doi.org/10.1016/j.meatsci.2020.108245>

779 Li, Z., Ha, M., Frank, D., Hastie, M., & Warner, R. D. (2023). Muscle fibre type
780 composition influences the formation of odour-active volatiles in beef. *Food Research*
781 *International*, 165, Article 112468. <https://doi.org/10.1016/j.foodres.2023.112468>

782 Lindahl, G., Lundström, K., & Tornberg, E. (2001). Contribution of pigment content,
783 myoglobin forms and internal reflectance to the colour of pork loin and ham from pure
784 breed pigs. *Meat Science*, 59(2), 141-151. [https://doi.org/10.1016/S0309-1740\(01\)00064-](https://doi.org/10.1016/S0309-1740(01)00064-X)
785 [X](https://doi.org/10.1016/S0309-1740(01)00064-X)

786 Liu, J., Ellies-Oury, M. P., Stoyanchev, T., & Hocquette, J. F. (2022). Consumer
787 perception of beef quality and how to control, improve and predict it? Focus on eating
788 quality. *Foods*, 11(12), Article 1732. <https://doi.org/10.3390/foods11121732>

789 Lomiwes, D., Farouk, M. M., Wu, G., & Young, O. A. (2014). The development of meat
790 tenderness is likely to be compartmentalised by ultimate pH. *Meat Science*, 96(1), 646-
791 651. <https://doi.org/10.1016/j.meatsci.2013.08.022>

792 Loudon, K. M. W., Lean, I. J., Pethick, D. W., Gardner, G. E., Grubb, L. J., Evans, A. C.,
793 & McGilchrist, P. (2018). On farm factors increasing dark cutting in pasture finished beef
794 cattle. *Meat science*, 144, 110-117. <https://doi.org/10.1016/j.meatsci.2018.06.011>

795 Lyford, C., Thompson, J., Polkinghorne, R., Miller, M., Nishimura, T., Neath, K., Allen,
796 P., & Belasco, E. (2010). Is willingness to pay (WTP) for beef quality grades affected by
797 consumer demographics and meat consumption preferences? *Australasian Agribusiness*
798 *Review*, 18, 1-17.

799 Madruga, M. S., & Mottram, D. S. (1995). The effect of pH on the formation of maillard-
800 derived aroma volatiles using a cooked meat system. *Journal of the Science of Food and*
801 *Agriculture*, 68(3), 305-310. <https://doi.org/10.1002/jsfa.2740680308>

802 Malheiros, B. A., Spers, E. E., Silva, H. M. R. D., & Contreras Castillo, C. J. (2022).
803 Southeast Brazilian Consumers' Involvement and Willingness to Pay for Quality Cues in
804 Fresh and Cooked Beef. *Journal of Food Products Marketing*, 28(6), 276-293.
805 <https://doi.org/10.1080/10454446.2022.2129539>

806 Martins, M. M., Saldaña, E., Teixeira, A. C. B., Selani, M. M., & Contreras-Castillo, C. J.
807 (2021). Going beyond sensory and hedonic aspects: A Brazilian study of emotions

808 evoked by beef in different contexts. *Meat Science*, 180, Article 108536.
809 <https://doi.org/10.1016/j.meatsci.2021.108536>

810 McKenna, D., Mies, P., Baird, B., Pfeiffer, K., Ellebracht, J., & Savell, J. (2005).
811 Biochemical and physical factors affecting discoloration characteristics of 19 bovine
812 muscles. *Meat Science*, 70(4), 665-682. <https://doi.org/10.1016/j.meatsci.2005.02.016>

813 Miller, M. F., Carr, M. A., Ramsey, C. B., Crockett, K. L., & Hoover, L. C. (2001).
814 Consumer thresholds for establishing the value of beef tenderness. *Journal of animal*
815 *science*, 79(12), 3062-3068. <https://doi.org/10.2527/2001.79123062x>

816 Mitacek, R. M., Ke, Y., Prenni, J. E., Jadeja, R., VanOverbeke, D. L., Mafi, G. G., &
817 Ramanathan, R. (2019). Mitochondrial degeneration, depletion of NADH, and oxidative
818 stress Decrease color stability of wet-aged beef Longissimus steaks. *Journal of Food*
819 *Science*, 84(1), 38-50. <https://doi.org/10.1111/1750-3841.14396>

820 Murray, J. M., Delahunty, C. M., & Baxter, I. A. (2001). Descriptive sensory analysis:
821 past, present and future. *Food research international*, 34(6), 461-471.
822 [https://doi.org/10.1016/S0963-9969\(01\)00070-9](https://doi.org/10.1016/S0963-9969(01)00070-9)

823 Obuz, E., & Dikeman, M. E. (2003). Effects of cooking beef muscles from frozen or
824 thawed states on cooking traits and palatability. *Meat Science*, 65(3), 993–997.
825 [https://doi.org/10.1016/S0309-1740\(02\)00314-5](https://doi.org/10.1016/S0309-1740(02)00314-5)

826 O'Quinn, T. G., Brooks, J. C., Polkinghorne, R. J., Garmyn, A. J., Johnson, B. J., Starkey,
827 J. D., Rathmann, R. J., & Miller, M. F. (2012). Consumer assessment of beef strip loin
828 steaks of varying fat levels. *Journal of Animal Science*, 90(2), 626-634.
829 <https://doi.org/10.2527/jas.2011-4282>

830 Patinho, I., Saldaña, E., Selani, M. M., Camargo, A. C., Merlo, T. C., Menegali, B. S.,
831 Silva, A. P. S., & Contreras-Castillo, C. J. (2019). Use of *Agaricus bisporus* mushroom in
832 beef burgers: Antioxidant, flavor enhancer and fat replacing potential. *Food Production,*
833 *Processing and Nutrition*, 1, 1-15. <https://doi.org/10.1186/s43014-019-0006-3>

834 Patinho, I., Selani, M. M., Saldaña, E., Bortoluzzi, A. C. T., Rios-Mera, J. D., Silva, C.
835 M., Kushida, M. M., & Contreras-Castillo, C. J. (2021). *Agaricus bisporus* mushroom as
836 partial fat replacer improves the sensory quality maintaining the instrumental
837 characteristics of beef burger. *Meat science*, 172, Article 108307.
838 <https://doi.org/10.1016/j.meatsci.2020.108307>

839 Picard, B., & Gagaoua, M. (2020). Muscle fiber properties in cattle and their relationships
840 with meat qualities: An overview. *Journal of Agricultural and Food Chemistry*, 68(22),
841 6021-6039. <https://doi.org/10.1021/acs.jafc.0c02086>

842 Poleti, M. D., Moncau, C. T., Silva-Vignato, B., Rosa, A. F., Lobo, A. R., Cataldi, T. R.,
843 Negrão, J. A., Silva, S. L., Eler, J. P., & Balieiro, J. C. C. (2018). Label-free quantitative
844 proteomic analysis reveals muscle contraction and metabolism proteins linked to ultimate
845 pH in bovine skeletal muscle. *Meat science*, *145*, 209-219.
846 <https://doi.org/10.1016/j.meatsci.2018.06.041>

847 Ponnampalam, E. N., Hopkins, D. L., Bruce, H., Li, D., Baldi, G., & Bekhit, A. E.
848 (2017). Causes and contributing factors to “dark cutting” meat: Current trends and future
849 directions: A review. *Comprehensive Reviews in Food Science and Food Safety*, *16*(3),
850 400-430. <https://doi.org/10.1111/1541-4337.12258>

851 Powell, L., Nicholson, K. L., Huerta-Montauti, D., Miller, R. K., & Savell, J. W. (2011).
852 Constraints on establishing threshold levels for Warner–Bratzler shear-force values based
853 on consumer sensory ratings for seven beef muscles. *Animal production science*, *51*(10),
854 959-966. <https://doi.org/10.1071/AN10267>

855 Purslow, P., Oiseth, S., Hughes, J., & Warner, R. (2016). The structural basis of cooking
856 loss in beef: Variations with temperature and ageing. *Food Research International*, *89*,
857 739-748. <https://doi.org/10.1016/j.foodres.2016.09.010>

858 Ramanathan, R. & Mancini, R. A. (2018). Role of Mitochondria in Beef Color: A
859 Review, *Meat and Muscle Biology* *2*(1), 309-320.
860 <https://doi.org/10.22175/mmb2018.05.0013>

861 Ramanathan, R., Kiyimba, F., Gonzalez, J., Mafi, G., & DeSilva, U. (2020). Impact of Up
862 and Down regulation of Metabolites and Mitochondrial Content on pH and Color of the
863 *Longissimus Muscle* from Normal-pH and Dark-Cutting Beef. *Journal of Agricultural*
864 *and Food Chemistry*, *68*, 7194-7203. <https://doi.org/10.1021/acs.jafc.0c01884>

865 Saldaña, E., Martins, M. M., Behrens, J. H., Valentin, D., Selani, M. M., & Contreras-
866 Castillo, C. J. (2020a). Looking at non-sensory factors underlying consumers' perception
867 of smoked bacon. *Meat science*, *163*, Article 108072.
868 <https://doi.org/10.1016/j.meatsci.2020.108072>

869 Saldaña, E., Serrano-León, J., Selani, M. M., & Contreras-Castillo, C. J. (2020b). Sensory
870 and hedonic impact of the replacement of synthetic antioxidant for pink pepper residue
871 extract in chicken burger. *Journal of food science and technology*, *57*, 617-627.
872 <https://doi.org/10.1007/s13197-019-04093-x>

873 Saldaña, E., Merlo, T. C., Patinho, I., Rios-Mera, J. D., Contreras-Castillo, C. J., &
874 Selani, M. M. (2021). Use of sensory science for the development of healthier processed
875 meat products: A critical opinion. *Current opinion in food science*, *40*, 13-19.

876 <https://doi.org/10.1016/j.cofs.2020.04.012>

877 Selani, M. M., Ramos, P. H. B., Patinho, I., França, F., dos Santos Harada-Padermo, S.,
878 Contreras-Castillo, C. J., & Saldaña, E. (2022). Consumer's perception and expected
879 liking of labels of burgers with sodium reduction and addition of mushroom flavor
880 enhancer. *Meat Science*, 185, Article 108720.

881 <https://doi.org/10.1016/j.meatsci.2021.108720>

882 Shackelford, S. D., M. Koohmaraie, M. F. Miller, J. D. Crouse, and J. O. Reagan. 1991.
883 An evaluation of tenderness of the *longissimus* muscle of Angus by Hereford versus
884 Brahman crossbred heifers. *Journal of Animal Science*, 69, 171-177.

885 <https://doi.org/10.2527/1991.691171x>

886 Sharedeh, D., Gatellier, P., Astruc, T., & Daudin, J. D. (2015). Effects of pH and NaCl
887 levels in a beef marinade on physicochemical states of lipids and proteins and on tissue
888 microstructure. *Meat Science*, 110, 24-31. <https://doi.org/10.1016/j.meatsci.2015.07.004>

889 Spanier, A. M., Flores, M., McMillin, K. W., & Bidner, T. D. (1997). The effect of post-
890 mortem aging on meat flavor quality in Brangus beef. Correlation of treatments, sensory,
891 instrumental and chemical descriptors. *Food Chemistry*, 59(4), 531-538.

892 [https://doi.org/10.1016/S0308-8146\(97\)00003-4](https://doi.org/10.1016/S0308-8146(97)00003-4)

893 Suman, S. P., Nair, M. N., Joseph, P., & Hunt, M. C. (2016). Factors influencing internal
894 color of cooked meats. *Meat Science*, 120, 133-144.

895 <https://doi.org/10.1016/j.meatsci.2016.04.006>

896 Suman, S. P., Wang, Y., Gagaoua, M., Kiyimba, F., & Ramanathan, R. (2023). Proteomic
897 approaches to characterize biochemistry of fresh beef color. *Journal of Proteomics*, 281,
898 Article 104893. <https://doi.org/10.1016/j.jprot.2023.104893>

899 Torres Filho, R. D. A., Cazedey, H. P., Fontes, P. R., Ramos, A. D. L. S., & Ramos, E.
900 M. (2017). Drip loss assessment by different analytical methods and their relationships
901 with pork quality classification. *Journal of Food Quality*, 2017, Article ID 9170768.

902 <https://doi.org/10.1155/2017/9170768>

903 Troy, D. J., & Kerry, J. P. (2010). Consumer perception and the role of science in the
904 meat industry. *Meat science*, 86(1), 214-226.

905 <https://doi.org/10.1016/j.meatsci.2010.05.009>

906 Ueda, Y., Watanabe, A., Higuchi, M., Shingu, H., Kushibiki, S., & Shinoda,
907 M. (2007). Effects of intramuscular fat deposition on the beef traits of Japanese Black
908 steers (Wagyu). *Animal Science Journal*, 78(2), 189-194. [https://doi.org/10.1111/j.1740-](https://doi.org/10.1111/j.1740-0929.2007.00424.x)
909 [0929.2007.00424.x](https://doi.org/10.1111/j.1740-0929.2007.00424.x)

910 Wakeling, I. N., & MacFie, H. J. (1995). Designing consumer trials balanced for first and
911 higher orders of carry-over effect when only a subset of k samples from t may be
912 tested. *Food Quality and Preference*, 6(4), 299-308. [https://doi.org/10.1016/0950-](https://doi.org/10.1016/0950-3293(95)00032-1)
913 [3293\(95\)00032-1](https://doi.org/10.1016/0950-3293(95)00032-1)

914 Wang, F., Holman, B. W., Zhang, Y., Luo, X., Mao, Y., & Hopkins, D. L. (2020).
915 Investigation of colour requirements of frozen beef rolls by Chinese consumers for hot
916 pot. *Meat Science*, 162, Article 108038. <https://doi.org/10.1016/j.meatsci.2019.108038>

917 Warner, R. & Miller, R. & Ha, M. & Wheeler, T. L. & Dunshea, F. & Li, X. & Vaskoska,
918 R. & Purslow, P. (2021). Meat Tenderness: Underlying Mechanisms, Instrumental
919 Measurement, and Sensory Assessment. *Meat and Muscle Biology* 4(2), 1-25.
920 <https://doi.org/10.22175/mmb.10489>

921 Watanabe, G., Motoyama, M., Orita, K., Takita, K., Aonuma, T., Nakajima, I., Tajima,
922 A., Abe, A., & Sasaki, K. (2019). Assessment of the dynamics of sensory perception of
923 Wagyu beef strip loin prepared with different cooking methods and fattening periods
924 using the temporal dominance of sensations. *Food Science & Nutrition*, 7(11), 3538-
925 3548. <https://doi.org/10.1002/fsn3.1205>

926 Watson, R., Polkinghorne, R., & Thompson, J. M. (2008). Development of the Meat
927 Standards Australia (MSA) prediction model for beef palatability. *Australian Journal of*
928 *Experimental Agriculture*, 48(11), 1368-1379. <https://doi.org/10.1071/EA07184>

929 Wicklund, S. E., Homco-Ryan, C., Ryan, K. J., Mckeith, F. K., Mcfarlane, B. J., &
930 Brewer, M. S. (2005). Aging and Enhancement Effects on Quality Characteristics of Beef
931 Strip Steaks. *Journal of Food Science*, 70(3), S242-S248. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2621.2005.tb07164.x)
932 [2621.2005.tb07164.x](https://doi.org/10.1111/j.1365-2621.2005.tb07164.x)

933 Wu, S., Han, J., Liang, R., Dong, P., Zhu, L., Hopkins, D. L., Zhang, Y., & Luo, X.
934 (2020). Investigation of muscle-specific beef color stability at different ultimate pHs.
935 *Asian-Australasian Journal of Animal Sciences*, 33(12), 1999-2007.
936 <https://doi.org/10.5713/ajas.19.0943>

937 Yang, X., Wang, J., Holman, B. W., Liang, R., Chen, X., Luo, X., Zhu, L., Hopkins, D.
938 L., & Zhang, Y. (2021). Investigation of the physicochemical, bacteriological, and
939 sensory quality of beef steaks held under modified atmosphere packaging and
940 representative of different ultimate pH values. *Meat Science*, 174, Article 108416.
941 <https://doi.org/10.1016/j.meatsci.2020.108416>

942 Zhang, Y., Kim, Y., Puolanne, E., & Ertbjerg, P. (2022). Role of freezing-induced
943 myofibrillar protein denaturation in the generation of thaw loss: A review. *Meat Science*,

944 190, Article 108841. <https://doi.org/10.1016/j.meatsci.2022.108841>
945 Zhu, Y., Gagaoua, M., Mullen, A. M., Viala, D., Rai, D. K., Kelly, A. L., Sheehan, D., &
946 Hamill, R. M. (2021). Shotgun proteomics for the preliminary identification of
947 biomarkers of beef sensory tenderness, juiciness and chewiness from plasma and muscle
948 of young Limousin-sired bulls. *Meat science*, 176, Article 108488.
949 <https://doi.org/10.1016/j.meatsci.2021.108488>
950 Zimoch, J., & Gullett, E. A. (1997). Temporal aspects of perception of juiciness and
951 tenderness of beef. *Food Quality and Preference*, 8(3), 203-211.
952 [https://doi.org/10.1016/S0950-3293\(96\)00049-3](https://doi.org/10.1016/S0950-3293(96)00049-3)
953

Table 1. Attributes, definitions, assessment techniques, and scale endpoints used in the Optimised Sensory Profile of beef.

*Attribute	Definition	Technique	Scale endpoints
Connective tissue presence	Visual presence of white-coloured gelatinous matter	The assessor should be seated at a 90° angle and can approach the sample without touching the plate, at a minimum distance of ~40 cm	Lowest: beef knuckle Highest: beef shin
Apparent opacity	Opacity outside colour (opaque)	The assessor should be seated at a 90° angle and can approach the sample without touching the plate, at a minimum distance of ~40 cm	Lowest: roasted Chester™ Highest: grilled broiler breast fillet
Surface brown colour	Brown pigmentation on meat surface, characteristic of cooked beef	The assessor should be seated at a 90° angle and can approach the sample without touching the plate, at a minimum distance of ~40 cm	Lowest: boiled beef Highest: grilled beef
Internal colour	Internal colour of beef, ranging from red to brown	The assessor should be seated at a 90° angle and can approach the sample without touching the plate, at a minimum distance of ~40 cm	Lowest: rare grilled beef Highest: well-done grilled beef
Roast beef aroma	Characteristic aroma of roast beef	Raise the sample container to the nose and perform up to 3 long inhalations	Lowest: meat cooked in water Highest: meat baked in the oven
Blood aroma	Characteristic aroma of raw beef	Raise the sample container to the nose and perform up to 3 long inhalations	Lowest: medium steak Highest: raw beef
Overall tenderness	Little force required for chewing	Chewing with molar teeth until sample is swallowed	Lowest: grilled beef outside flat Highest: grilled beef tenderloin
Overall juiciness	Release of liquid during chewing	Chewing with molar teeth until sample is swallowed	Lowest: grilled beef knuckle Highest: Grilled sirloin cap steak
Hardness	Force required to achieve certain product deformation	Chewing with molar teeth until sample is swallowed	Lowest: medium rump steak (63 °C) Highest: well-done outside flat steak (79 °C)
Dryness	No liquid release during chewing	Chewing with molar teeth until sample is swallowed	Lowest: rare sirloin cap steak (60 °C) Highest: cooked pork chop (79 °C)
Fibrousness		Place the sample in the mouth and generate a perception within 7 chews	

*Attribute	Definition	Technique	Scale endpoints
	Presence of fibres that persists throughout chewing		Lowest: grilled tenderloin steak Highest: grilled outside flat steak
Chewiness	The number of chews required for the product to be ready for swallowing	Place the sample in the mouth and generate a perception within 7 chews	Lowest: medium tenderloin (63 °C) Highest: well-done pork chop (79 °C)
Roast beef flavour	Characteristic roast meat flavour	Chewing with molar teeth until sample is swallowed	Lowest: Boiled beef Highest: baked beef
Meaty beef flavour	Characteristic beef flavour	Chewing with molar teeth until sample is swallowed	Lowest: Boiled beef Highest: grilled beef
Fat flavour	Perception of fat in the mouth during chewing	Place the sample in the mouth and generate a perception within 7 chews	Lowest: cooked chicken breast fillet (79 °C) Highest: cooked beef rib (69 °C)
Blood flavour	Flavour associated with blood in cooked meat products; closely related to a metallic flavour	Place the sample in the mouth and generate a perception within 7 chews	Lowest: well-done sirloin cap steak (82 °C) Highest: rare sirloin steak (55 °C)

* For each attribute, a 9-cm unstructured linear intensity scale was utilised, with intensity levels ranging from 'Lowest' at 0 to 'Highest' at 9, anchored at both ends of the scale

Table 2. Proximate composition of intermediate and normal pHu beef 3 days *post-mortem*

Treatment	Moisture (%)	Protein (%)	Lipid (%)	Ash (%)
Intermediate pHu	74.95	20.89	2.67	1.03
Normal pHu	73.49	24.02	1.40	1.06

Proximate composition was determined by method (AOAC, 1995) and (ISO 1871:2009; 936:1998).

Table 3 – Mean ratings* of the sensory attributes evaluated in the intermediate and normal pHu beef 3 days *post-mortem*. Measurements on a 9-cm unstructured linear scale (0 = none/Lowest; 9= Highest)

Sensory attributes	Intermediate pHu	Normal pHu	t-value	P-value**
Connective tissue presence	1.90±2.21	1.62±2.21	0.68	0.50
Apparent opacity	2.36±2.31	3.02±2.31	-1.57	0.12
Surface brown colour	7.18±1.57	7.44±1.57	-0.90	0.37
Internal colour	6.35±1.98	6.22±1.98	0.36	0.72
Roast beef aroma	6.92±2.29	7.10±2.29	-0.43	0.67
Blood aroma	0.97±1.46	0.81±1.46	0.60	0.55
Overall tenderness	4.19±2.88	4.06±2.88	0.23	0.82
Overall juiciness	5.88±2.50	4.48±2.50	3.18	0.00
Hardness	4.35±2.94	4.21±2.94	0.27	0.79
Dryness	2.35±2.35	3.02±2.35	-1.56	0.12
Fibrousness	3.66±2.80	3.64±2.80	0.03	0.97
Chewiness	5.42±2.85	5.36±2.85	0.12	0.91
Roast beef flavour	6.92±2.26	6.89±2.26	0.06	0.95
Meaty beef flavour	7.95±1.30	7.87±1.30	0.34	0.73
Fat flavour	1.76±2.02	1.78±2.02	-0.04	0.97
Blood flavour	1.71±1.71	1.21±1.71	1.64	0.10

* Measurements on a 9-cm unstructured linear scale (0 = none/Lowest; 9= Highest)

**Sensory attributes with a significant ($P < 0.05$) effect on responses are marked in bold. Results are expressed as mean \pm standard deviation.

Table 4. Quality parameters of intermediate and normal pHu beef 3 days *post-mortem*

Parameters	Intermediate pHu	Normal pHu	P-value*
Ultimate pH	5.89±0.04	5.58±0.04	0.01
Drip loss (%)	2.12±0.40	3.32±0.40	0.10
TPA			
Hardness (N)	62.19±7.01	81.94±7.01	0.12
Springiness	0.01±0.00	0.01±0.00	0.66
Cohesiveness	0.01±0.00	0.01±0.00	0.19
Chewiness	21.83±3.65	32.08±3.65	0.12
Gumminess	34.13±3.96	48.73±3.96	0.06

The parameters which have a significant ($P < 0.05$) effect on responses are marked in bold. Results are expressed as mean \pm standard error of the mean. TPA, texture profile analysis. N = Newtons.

Table 5. Effect of ultimate pH (pHu) and aging days on quality parameters of intermediate and normal pHu beef at 3, 14, and 28 days *post-mortem*

Parameters	<i>L</i> *	<i>a</i> *	<i>b</i> *	Chroma	Hue	Surface colour stability	%OMb	%MMb	%DMb	%WHC	%Cooking losses	Shear force (N)
Source of variation												
Intermediate pHu	38.50 ^b	25.81 ^a	18.12 ^b	31.55 ^b	34.70 ^a	5.94 ^a	68.22 ^b	17.17 ^a	14.51 ^a	34.22 ^a	28.43 ^a	85.19 ^b
Normal pHu	41.90 ^a	28.06 ^a	20.60 ^a	34.87 ^a	35.61 ^a	6.00 ^a	73.38 ^a	17.92 ^a	8.66 ^b	32.74 ^a	26.00 ^a	102.43 ^a
SEM	0.44	0.74	0.76	1.05	0.39	0.31	1.27	0.65	1.21	0.77	1.14	5.34
Aging days	<i>L</i> *	<i>a</i> *	<i>b</i> *	Chroma	Hue	Surface colour stability	%OMb	%MMb	%DMb	%WHC	%Cooking losses	Shear force (N)
3	42.13 ^a	21.83 ^b	13.55 ^b	25.71 ^b	31.59 ^b	3.64 ^b	61.78 ^b	23.06 ^a	15.16 ^a	30.92 ^b	25.10 ^a	104.89 ^a
14	38.60 ^b	28.79 ^a	21.63 ^a	36.03 ^a	36.75 ^a	6.99 ^a	73.09 ^a	14.05 ^b	12.68 ^a	33.86 ^{ab}	29.44 ^a	92.67 ^a
28	39.87 ^b	30.17 ^a	22.90 ^a	37.89 ^a	37.13 ^a	7.28 ^a	77.53 ^a	15.52 ^b	6.91 ^b	35.65 ^a	27.10 ^a	83.86 ^a
SEM	0.53	0.91	0.93	1.28	0.48	0.38	1.55	0.79	1.48	0.95	1.40	6.54

Different letters in the same column indicate significant differences ($P < 0.05$) according to the Tukey's test. SEM: Standard error of the mean. Luminosity (*L**), Redness (*a**), Yellowness (*b**), Chroma and Hue. Omb: oxymyoglobin, MMb: metmyoglobin. DMb: deoxymyoglobin. Surface colour stability, Water-holding capacity (WHC), Cooking losses and Shear force (N = Newtons).

Fig. 1. Principal Component Analysis of ODP data from intermediate and normal pHu beef at 3 days *post-mortem*. **A)** Individual data points. **B)** Variables plots.

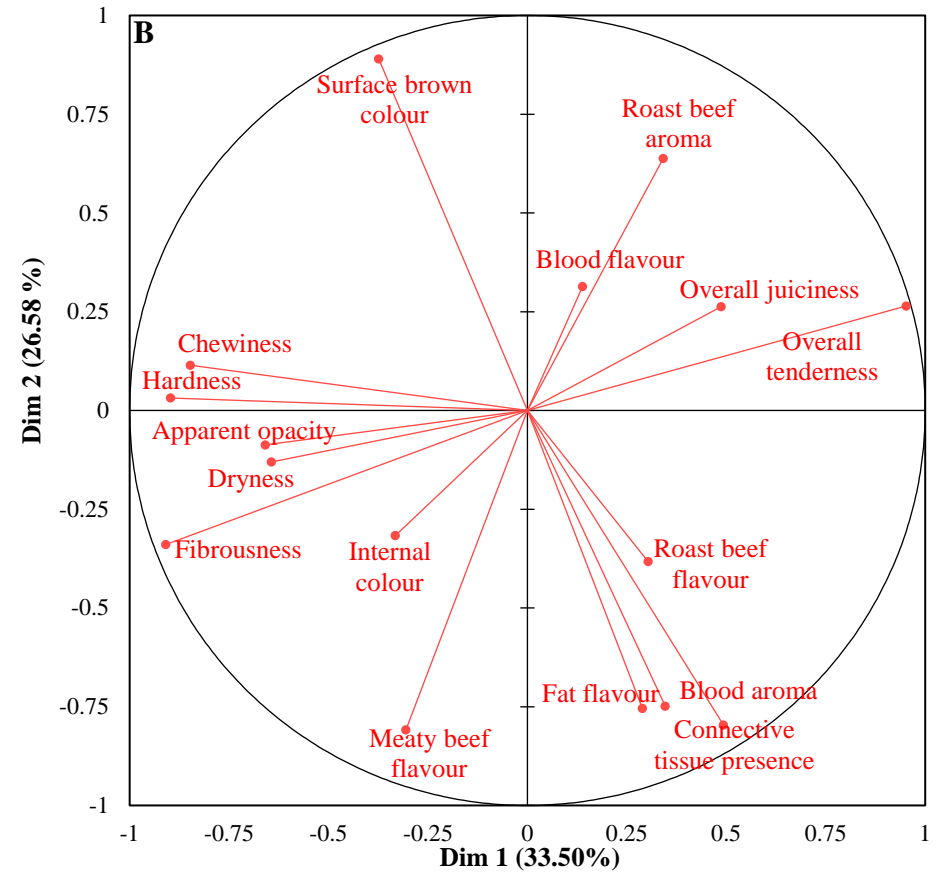
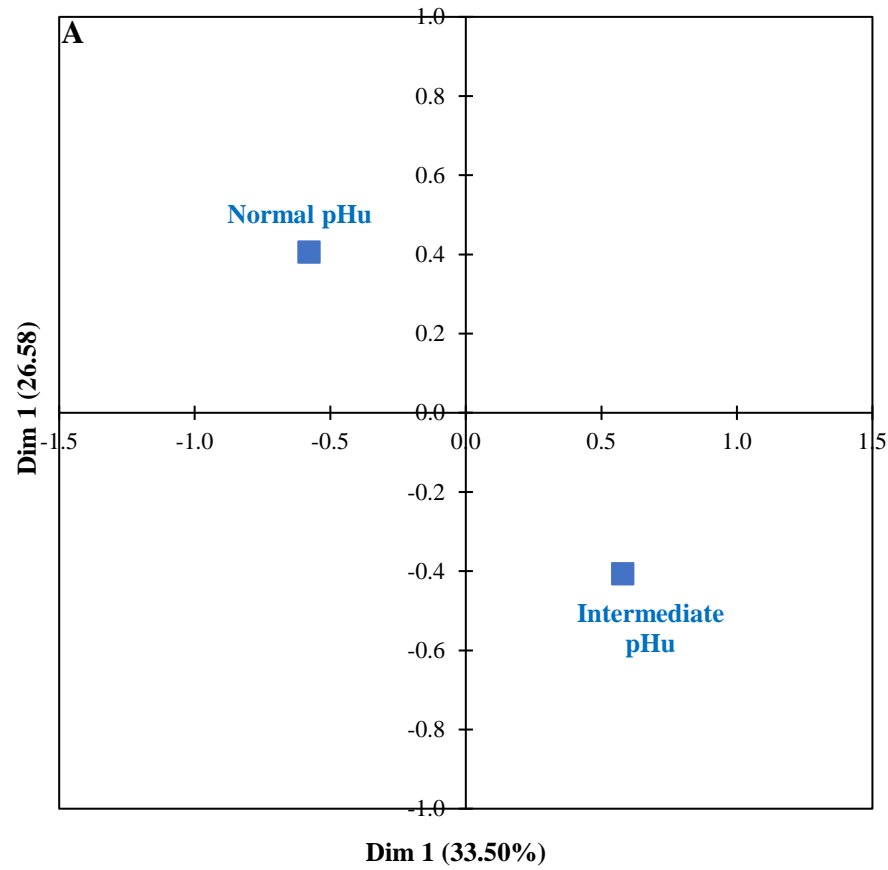


Fig. 2. Correspondence and Principal Coordinate Analysis of Sensory Descriptors. **A)** Correspondence Analysis of sensory descriptors for intermediate and normal pHu beef at 3 days post-mortem, depicted in the first two dimensions derived from PCA of the Check-All-That-Apply (CATA) questions and the ideal sample. **B)** Representation of the relationship between overall liking and the sensory descriptors by Principal Coordinate Analysis.

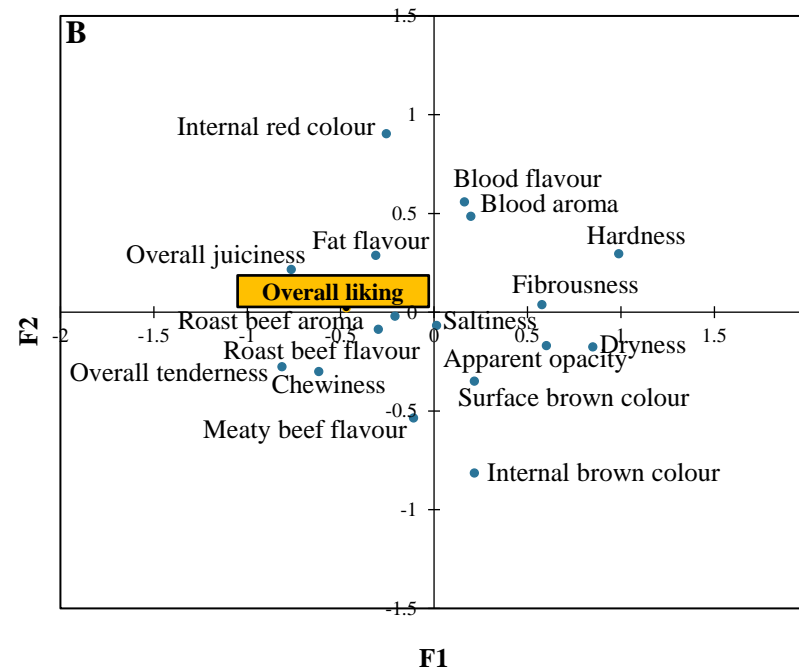
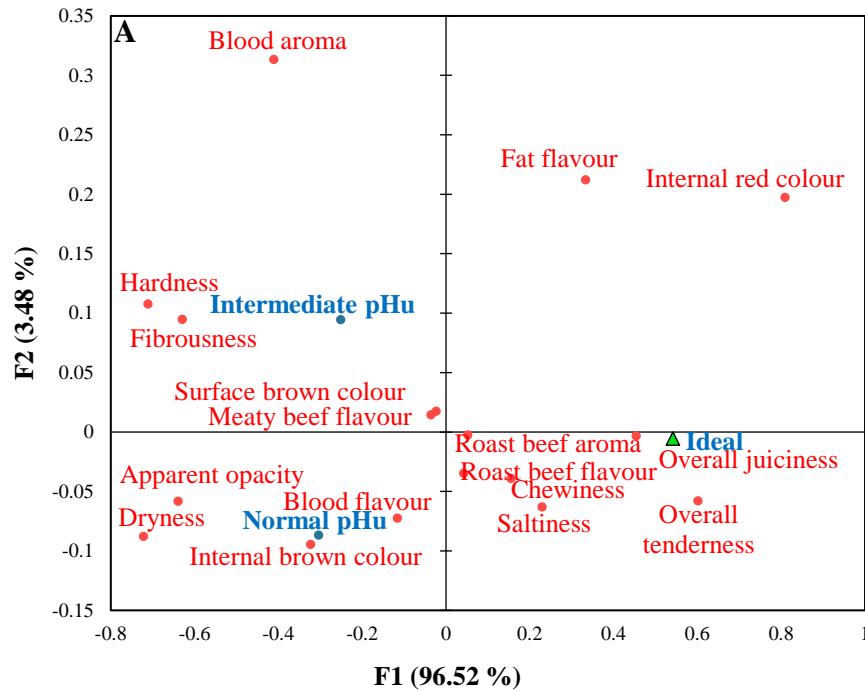


Fig. 3. Penalty-Lift Analysis on Sensory Attributes and Overall Liking Scores. Penalty-lift analysis showing the impact of sensory attributes on the mean overall liking scores for beef samples with intermediate and normal pHu, 3 days post-mortem.

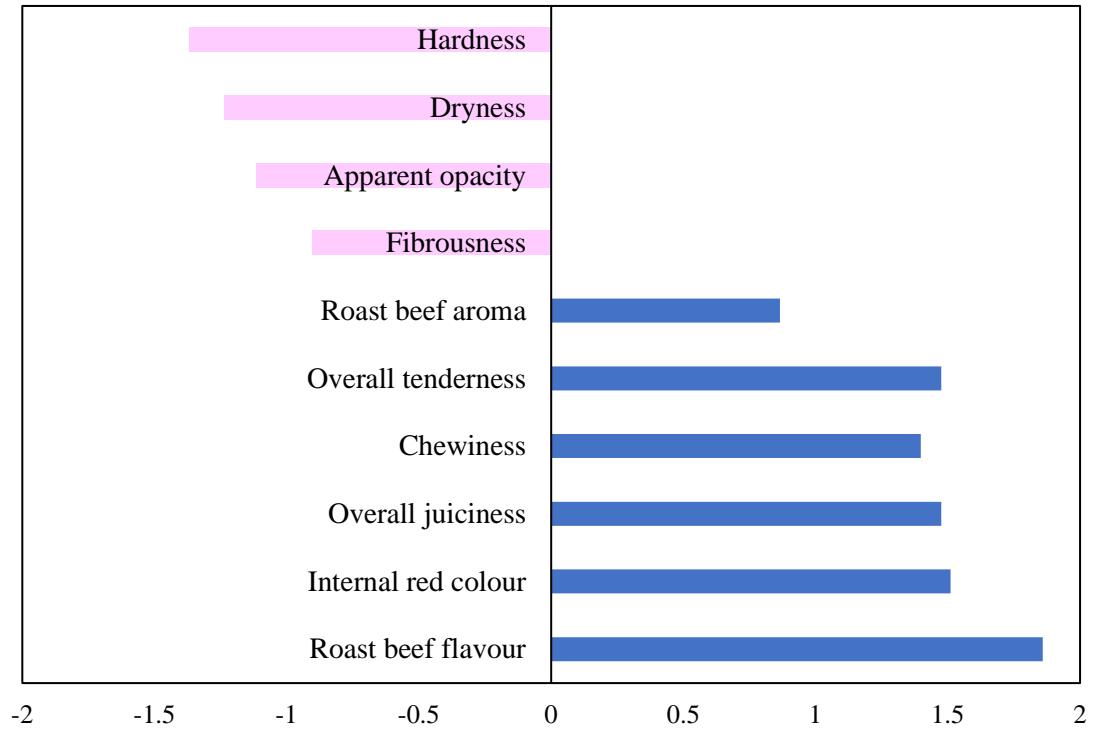
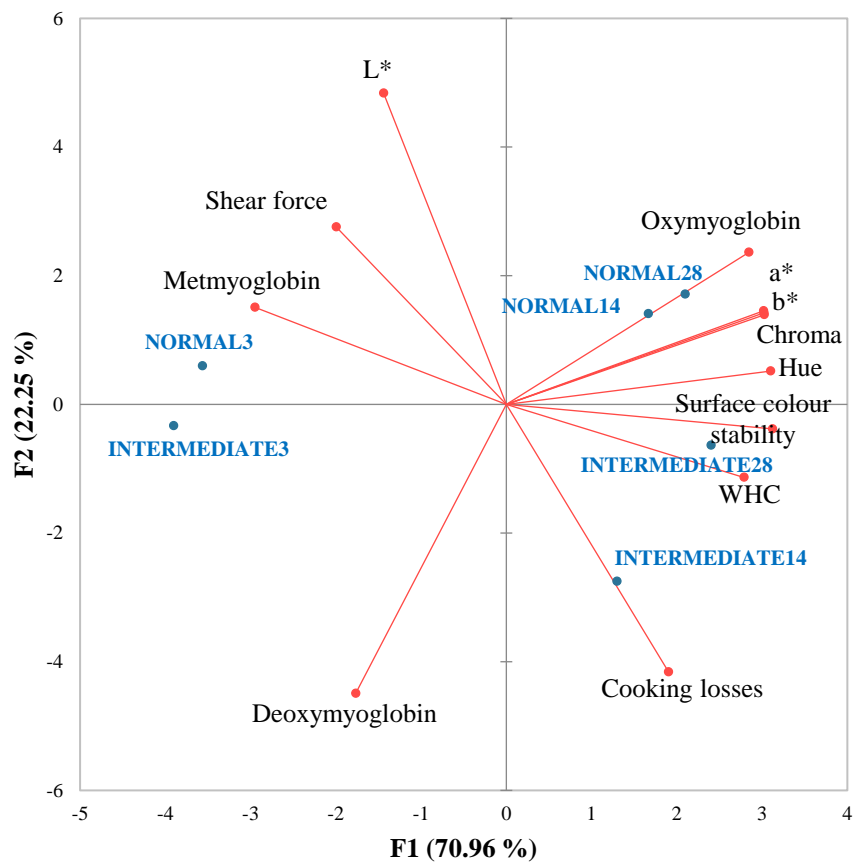


Fig. 4. Principal Component Analysis of Instrumental Quality Parameters in Beef Samples.

Principal Component Analysis of various instrumental quality parameters measured in beef samples with intermediate and normal pHu at 3, 14, and 28 days post-mortem. The parameters evaluated include luminosity (L^*), redness (a^*), yellowness (b^*), chroma, hue angle, and concentrations of oxymyoglobin, metmyoglobin, and deoxymyoglobin, alongside assessments of surface color stability, water holding capacity (WHC), cooking losses, and shear force.



Highlights

- Sensory analysis of Nellore beef comparing normal and intermediate pHu
- Intermediate pHu beef scored higher for juiciness according to trained panellists
- Consumers could not differentiate between intermediate and normal pHu beef
- Higher deoxymyoglobin on day 3 than on day 28 in intermediate pHu beef
- Beef colour is influenced by pHu and ageing: L^* , b^* , Chroma, and Oxymyoglobin