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## Improved assessment of the impacts of plant protection products on certain soil ecosystem services requires better consideration of terrestrial microalgae and cyanobacteria

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#### Abstract:

There is growing scientific and societal consciousness that the environmental risks and impacts of plant protection products (PPPs) cannot be properly assessed without considering ecosystem services. However, the science on this issue remains incomplete and fragmented, as recently illustrated in a collective scientific assessment that pointed out the limited knowledge on the risks and impacts of PPPs on soil ecosystem services, which are clearly overlooked. Beside soil ecosystem services, certain key players involved in these services are largely overlooked in the scientific literature on the risks and impacts of PPPs, namely soil microbial photosynthetic communities. Here, we followed the principles of evidence-based logic chain approaches to show the importance of considering these microorganisms when studying the impacts of PPPs on certain services provided by soil ecosystems, with a focus on regulating and maintenance services that play a role in the regulation of baseline flows and extreme events. Terrestrial microalgae and cyanobacteria are ubiquitous photosynthetic microorganisms that, together with other soil micro- and macro-organisms, play key roles in the ecosystem functions that underpin these ecosystem services. There is an extensive literature on the ecotoxicological effects of PPPs on different organisms including soil microorganisms, but studies concerning soil microbial photosynthetic communities are very scarce. However, there is scientific evidence that herbicides can have both direct and indirect impacts on these microbial photosynthetic communities. Given that they play key functional roles, we argue that soil microbial photosynthetic communities warrant greater attention in efforts to assess the environmental risks and impacts of PPPs and, ultimately, help preserve or restore the regulating and maintenance services provided by soil ecosystems.

#### Effects of PPPs on the regulating and maintenance services provided by soils

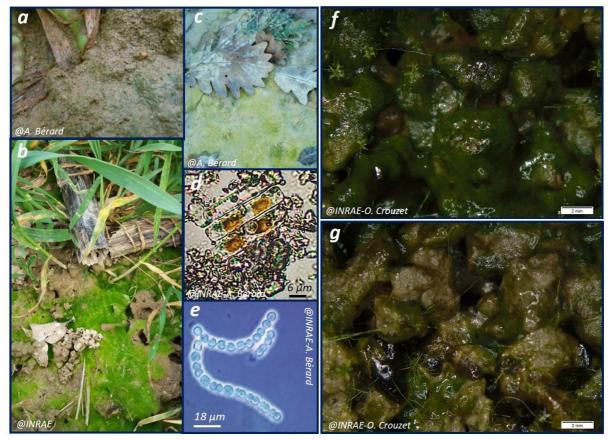
Prompted by recommendations made by the European Food Safety Authority (EFSA 2010; 2016), scientists, environmental managers, stakeholders, and regulators are increasingly conscious of the importance of protecting ecosystem services against the adverse effects of plant protection products (PPPs) (Nienstedt et al. 2012; Arts et al. 2015; Rumschlag et al. 2020). Several scientists in the fields of ecotoxicology and environmental risk assessment of chemicals have been working for years to develop research and propose conceptual and operational approaches to achieve this objective (e.g., Cairns and Niederlehner 1994; Forbes and Calow 2013; Brown et al. 2021; Faber et al. 2021; Maltby et al. 2022). However, there are still few studies providing hard information on the risks and effects of PPPs on ecosystem services, and the resulting science on this issue remains incomplete and fragmented (Pesce et al. 2023a). This was highlighted by the conclusions of a collective scientific assessment performed between 2020 and 2022 addressing the state of knowledge on the effects of PPPs on biodiversity and ecosystem services (Pesce et al. 2021). In particular, this collective scientific assessment pointed out the limited knowledge on the effects of PPP on services provided by soil ecosystems (Pesce et al. 2023b). According to the most recent version of the Common International Classification of Ecosystem Services (CICES, version 5.1.; Haines-Young and Potschin 2018), these services include regulating and maintenance services belonging to the group "regulation of baseline flows and extreme events," especially the classes "control of erosion rates," "buffering and attenuation of mass movement," and "hydrological cycle and water flow regulation, including flood control and coastal protection," which are largely neglected in the scientific literature dealing with the impacts of PPPs.

# Abundance, diversity, and functional role of soil microbial photosynthetic communities in soil ecosystems

Terrestrial microalgae and cyanobacteria communities (mainly designated hereafter as soil microbial photosynthetic communities) are ubiquitous photosynthetic microorganisms. They are pioneer autotrophic components of microbial communities growing on the surface of soils (Fig. 1). In temperate agricultural soils, they are highly abundant (from 1 × 104 to 1 × 107 cells/g soil, depending on season and land use, (Metting 1981; Zancan et al. 2006). Soil microbial phototroph communities can reach for around 25% of the microbial biomass of agricultural soils (Abinandan et al. 2019). These communities are highly diversified: eukaryotic microalgae are mainly represented by Chlorophyceae, Bacillariophyceae (Foets et al. 2021), Xanthophyceae and Chrysophyceae, while prokaryotic cyanobacteria include Oscillatoriales, Nostocales, and Chroococcales, many of which are filamentous (Metting 1981; Davies et al. 2013; Lentendu et al. 2014; Djemiel et al. 2020). While certain groups are thought to be specific to soils, others are also found in aquatic environments (Bérard et al. 2004), suggesting biological transfers between aquatic and terrestrial environments (Pfister et al. 2017).

In some crops alternating between flooding and drying out (e.g., rice paddies), cyanobacteria, and, to a much lesser extent, microalgae have been deeply studied on the surface of emerged and submerged soil (Singh et al. 2018; Kaushik et al. 2019). Soil microbial photosynthetic communities form a soil– atmosphere microbial interface (subjected to sprayed PPP), which makes them central to addressing the challenges of sustainable agriculture and preservation and management of ecosystem services, in particular soil health and agricultural production (Ramakrishnan et al. 2023).

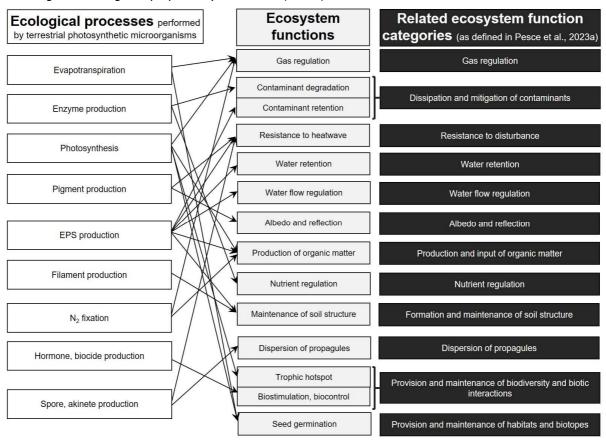
**Fig. 1**: Pictures illustrating soil microbial phototrophs. **a** Conventional cropland (maize), **b** conservation cropland (barley), and **c** forest (Mediterranean). Microscopy observations: **d** diatoms; **e** cyanobacteria; **f** and **g** photosynthetic microbial biofilms growing in 50 days incubated microcosms on soil aggregates from organic cropping system: incubation with the herbicide isoproturon (**g**) at field dose recommended as 2.4 L ha<sup>-1</sup> of the commercial formulation clearly inhibited the development of photosynthetic biofilms on soil aggregates, compared with incubated microcosms without isoproturon (**f**)



Indeed, these microbial communities are involved in many ecosystem functions, which are achieved via various ecological processes (Fig. 2), such as photosynthesis, pigment production, biofilm/filament, and exopolysaccharide production (Malam Issa et al. 2007; Chamizo et al. 2018), enzymes, hormone and biocide production (Abinandan et al. 2019; Poveda 2021; Santini et al. 2021; Righini et al. 2022; Osman et al. 2023), and N2 fixation (Peng and Bruns 2019). In croplands that experience frequent soil surface disturbances (e.g., tillage practices), microbial production by soil photosynthetic microorganisms can range from 80 to 157 g C m-2 year-1, which represent 6 to 7% of the net primary production of terrestrial vegetation (Shimmel and Darley 1985; Jassey et al. 2022). The presence of soil microbial photosynthetic communities has also been shown to reduce the net emission of CO2 from agricultural soils under experimental conditions (Sauze et al. 2017).

Experimental and field studies have already demonstrated that soil microbial photosynthetic communities play a role in maintaining soil structure by producing exopolysaccharides and filaments/biofilms that help to control (i) soil aggregation and thus limiting erosion (Metting 1987; Knapen et al. 2007; Kidron and Drahorad 2021; Keqiang et al. 2023) and (ii) soil water retention that contributes to hydrological cycle and water flow regulation (Cantón et al. 2020; Rossi et al. 2021; Giora et al. 2020; Kidron 2021). Native soil microbial photosynthetic communities can also enhance soil dissipation of some PPPs, suggesting a key role on soil surface as a bio-filter (Davies et al. 2013). The inoculation of biocrust organisms (particularly pioneer cyanobacteria) is increasingly used as a soil restoration solution (e.g., Chamizo et al. 2020; Zhao et al. 2021, Dhawi 2023). However, in the context of agroecosystems, the knowledge about the functional role of soil microbial photosynthetic communities is scarce and mostly outdated (Metting 1987; Knapen et al. 2007; Malam Issa et al. 2007; Peng and Bruns 2019), with the exception of rice fields, where microalgae and cyanobacteria are recognized for their beneficial effects on crops and are sometimes taken into account in rice cropping system simulations (Gaydon et al. 2012).

**Fig. 2**: Putative list of ecological processes performed by terrestrial photosynthetic microorganisms in soil ecosystems and possible unweighted contribution of each of these processes to soil ecosystem functions classified according to the categories proposed by Pesce et al. (2023a)



Given the diversity of ecological processes performed by soil microbial photosynthetic communities, they may be significantly involved in many of the 12 categories of ecosystem functions potentially impacted by PPPs, as recently proposed by Pesce et al. (2023a). Ultimately, these ecosystem functions are the pillars of numerous soil ecosystem services. This is why we call for better consideration of soil microbial photosynthetic communities to preserve or restore soil ecosystem services.

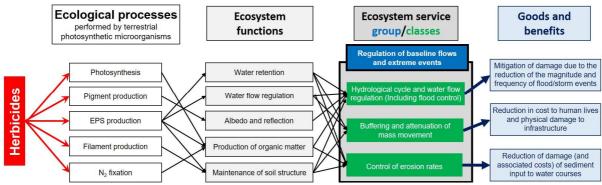
#### Limited knowledge about the effects of PPPs on terrestrial microalgae and cyanobacteria

There is an extensive literature on the ecotoxicological effects of PPPs on different taxonomic groups (including for instance terrestrial macro-invertebrates and heterotrophic microorganisms as well as aquatic microalgae and cyanobacteria), but soil microbial photosynthetic communities remain some of the least documented biological communities in ecotoxicology. However, a few studies have shown that pesticides can negatively impact these non-target communities (when considering the community level, less than 30 relevant articles were obtained from the bibliographic analysis performed on the Web of Science — WoS). For instance, herbicides have been shown to strongly affect these terrestrial photosynthetic microbial communities by decreasing their biomass and diversity (Pipe 1992; Zancan et al. 2006; Crouzet et al. 2013; Joly et al. 2014). Herbicide effects also disrupt ecological processes such as soil photosynthesis (Bérard et al. 2004), biomass production (Pipe 1992), N2 fixation (Dash et al. 2018; Wegener et al. 1985), and pigment and exopolysaccharide production (Crouzet et al. 2019; Joly et al. 2014; Zaady et al. 2013). Such effects can have further consequences on ecosystem functions. Zaady et al. (2013) reported that the triazine herbicide simazine increased soil runoff and decreased hydraulic conductivity due to its negative impact on soil chlorophyll biomass and exopolysaccharides. Crouzet et al. (2019) showed that the phenylurea herbicide isoproturon has a negative impact on soil aggregate stability by decreasing chlorophyll biomass and exopolysaccharide production and changing pigment composition.

# Potential impacts of PPPs on soil ecosystem services: "regulation of baseline flows and extreme events" as an example

As mentioned in the previous section, scientific evidence exists concerning the impact of some herbicides on several ecological processes performed by soil microbial photosynthetic communities, including photosynthesis, pigment, EPS, and possibly filament production (components of filamentous microalgae and cyanobacteria biomass impacted by herbicides), as well as N2 fixation. Herbicide effects on such ecological processes can have consequences on different ecosystem functions which themselves support soil ecosystem services.

**Fig. 3**: Schematic network of logic chains showing how three classes of the "Regulation of baseline flows and extreme events" ecosystem services and related goods and benefits can suffer cascading effects caused by ecotoxicological impacts of herbicides on selected ecological processes carried out by soil microbial phototrophs. EPS, exopolysaccharides excreted by microbial phototrophs in the soil



To improve the assessment of the risks and effects of PPPs (or other chemicals) on ecosystem services, several authors advocate the development of evidence-based logic chain approaches (Hayes et al. 2018; Faber et al. 2021; Maltby et al. 2021).

These approaches use logic chains on specific impacts, from effects on key lower-trophic-level organisms, which disrupt a wide range of ecological processes and functions, to consequences on broader ecosystem services (Hayes et al. 2018). In Fig. 3, this kind of approach was used to illustrate how ecotoxicological effects of PPPs on the abovementioned ecological processes performed by soil microbial photosynthetic communities can impact some ecosystem functions supporting soil ecosystem services belonging to the group "regulation of baseline flows and extreme events" and on associated goods and benefits (CICES, version 5.1.; Haines-Young and Potschin 2018).

This specific example, which shows how some regulation and maintenance services provided by soil ecosystems can suffer cascading effects caused by ecotoxicological impacts of PPPs on ecological processes carried out by soil microbial photosynthetic communities, highlights the need to consider these organisms in environmental assessments on the risks and effects of PPPs (and other chemicals), not only in order to protect these communities, their diversity, and the ecological processes they perform, but also to protect or restore the functioning of soil agroecosystems and the services they provide, as addressed in semi-arid contexts with biocrusts (Zhao et al. 2021). However, risk and effect assessment targeting more specifically selected soil ecosystem functions and services needs taking into consideration all the main biological actors involved, meaning that soil phototrophic microorganisms should be studied together with other soil microorganisms identified as key interacting players of a soil microbial ecosystem as well as macro-organisms identified as key players in the considered evidence-based logic chain.

#### **Conclusions and recommendations**

Very few studies have been carried out to characterize the risks and impacts of PPPs on the diversity and activities of soil photosynthetic microbial communities, and most were published in the 1970s-1990s, which explains the paucity of quantitative data on these effects (Mamy et al. 2022). Current research on this topic mainly concerns rice fields, with a focus on cyanobacteria and their diazotrophic activity contributing to N2 fixation (Kaushik et al. 2019). In order to significantly develop knowledge concerning the impact of PPPs on soil microbial photosynthetic communities, it is necessary to consolidate the methods for sampling and characterizing these communities in different soils and land use contexts (Barragan et al. 2018). Besides these methods, which have to be specific to the terrestrial environment, it is also important to draw on methodological and conceptual developments from research aimed at assessing the impact of PPPs on freshwater photosynthetic microorganisms, which are the subject of numerous studies at various biological levels (Vonk and Kraak 2020). For example, the use of functional traits, as applied to aquatic microalgae (Baert et al. 2017), could help to better understand the functional consequence of the ecotoxicological effects of PPPs on soil microbial photosynthetic communities, calling for further investigations about their own functional traits. The application of omics approaches (such as metabarcoding, Djemiel et al. 2020; Rivera et al. 2020; as well as transcriptomics, proteomics and metabolomics, Moisset et al. 2015, Lips et al. 2022), which are increasingly used to investigate the structural and functional responses of aquatic photosynthetic microorganisms to PPPs at different biological levels (from strains to communities) could provide new insights into the effects of PPPs on soil photosynthetic microorganisms. For instance, it could help to investigate their sensitivity to PPPs, based on their phylogeny, and to study alterations in community diversity and functions in response to PPP exposure. Fluorimetry techniques (PAM and spectral measurements), currently applied to aquatic environments, could also be used to characterize the responses of soil photosynthetic microorganisms to PPPs. Promisingly, it has thus been shown that spectral analyses based on reflectance or fluorescence can be applied to the study of soil biological crusts (Rodriguez-Caballero et al. 2015).

Flow cytometry, which is already widely used in aquatic environments, can also be applied to soil ecosystems (Menyhart et al. 2018). Finally, the development of imaging tools (Zhang et al. 2022, 2023) would also enable progress to be made in taxonomic recognition and more global observation on the scale of microbial crusts.

Moreover, relying on these different methods, terrestrial photosynthetic microbial communities could potentially serve as good indicators of soil quality. The biodiversity of freshwater diatoms is currently used for biomonitoring purposes, to assess the ecological quality of aquatic ecosystems, thanks to the use of biological indices (e.g., BDI, Coste et al. 2009) based on the large ecological amplitude of diatoms and their species-specific sensitivity to pollution. The connectivity between terrestrial and aquatic environments means that several photosynthetic microorganisms can be found both in soils and waters. One example is the taxonomic group of diatoms. Martínez-Carreras et al. (2015) and Klaus et al. (2015) demonstrated rapid transfers of terrestrial diatoms from soil surfaces to surrounding waters. The diatom group may therefore hold potential for use as an indicator of pollution (especially PPPs) throughout the terrestrial–aquatic continuum.

In general, there is a need to make progress on the links between diversity, function, and services impacted by PPPs (Pesce et al. 2023a). Going beyond the simple question of PPPs, this is clearly identified as a major challenge in microbial ecotoxicology (Hellal et al. 2023). Thus, efforts to improve the evaluation and quantification of the impacts of PPPs and other toxicants on soil ecosystem services demand greater interdisciplinarity, with more interaction between ecotoxicologists, environmental scientists in other disciplines, such as those that connect to physics and pedology (in order to take into account the specific context of soils) and scientists from the humanities and social sciences working in the field of ecosystem services. This nexus of sciences and disciplines will be an essential step towards sustainable use of PPPs that can help maintain or even restore the ecological quality and functioning of soil ecosystems and thus enable them to provide vital ecosystem services.

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#### References

- Abinandan S, Subashchandrabose SR, Venkateswarlu K, Megharaj M (2019) Soil microalgae and cyanobacteria: the biotechnological potential in the maintenance of soil fertility and health. Crit Rev Biotechnol 39:981–998. <u>https://doi.org/10.1080/07388551.2019.1654972</u>
- Arts GHP, Dollinger M, Kohlschmid E, Maltby L, Ochoa-Acuna H, Poulsen V (2015) An ecosystem services approach to pesticide risk assessment and risk management of non-target terrestrial plants: recommendations from a SETAC Europe workshop. Environ Sci Pollut Res 22:2350–2355. https://doi.org/10.1007/s11356-014-3637-6
- Baert JM, De Laender F, Janssen CR (2017) The consequences of nonrandomness in species-sensitivity in relation to functional traits for ecosystem-level effects of chemicals. Environ Sci Technol 51:7228–7235. https://doi.org/10.1021/acs.est.7b00527
- Barragan C, Wetzel CE, Ector L (2018) A standard method for the routine sampling of terrestrial diatom communities for soil quality assessment. J Appl Phycol 30:1095–1113. <u>https://doi.org/10.1007/s10811-017-1336-7</u>
- Bérard A, Rimet F, Capowiez Y, Leboulanger C (2004) Procedures for determining the pesticide sensitivity of indigenous soil algae – a possible bioindicator of soil contamination? Arch Environ Contam Toxicol 46:24–31. <u>https://doi.org/10.1007/s00244-003-2147-1</u>

- Brown AR, Marshall S, Cooper C, Whitehouse P, Van den Brink PJ, Faber JH, Maltby L (2021) Assessing the feasibility and value of employing an ecosystem services approach in chemical environmental risk assessment under the Water Framework Directive. Sci Total Environ 789:147857. https://doi.org/10.1016/j.scitotenv.2021
- Cairns J, Niederlehner BR (1994) Estimating the effects of toxicants on ecosystem services. Environ Health Perspect 102:936–939. <u>https://doi.org/10.1289/ehp.94102936</u>
- Cantón Y, Chamizo S, Rodriguez-Caballero E, Lázaro R, Roncero-Ramos B, Román JR, Solé-Benet A (2020) Water regulation in cyanobacterial biocrusts from drylands: negative impacts of anthropogenic disturbance. Water 12:720. https://doi.org/10.3390/w12030720
- Chamizo S, Mugnai G, Rossi F, Certini G, De Philippis R (2018) Cyanobacteria inoculation improves soil stability and fertility on different textured soils: gaining insights for applicability in soil restoration. Front Environ Sci 6:49. <u>https://doi.org/10.3389/fenvs.2018.00049</u>
- Chamizo S, Adessi A, Certini G, De Philippis R (2020) Cyanobacteria inoculation as a potential tool for stabilization of burned soils. Restor Ecol 28:S106–S114. <u>https://doi.org/10.1111/rec.13092</u>
- Coste M, Boutry S, Tison-Rosebery J, Delmas F (2009) Improvements of the Biological Index (BDI): description and efficiency of the new version (BDI-2006). Ecol Indic 9:621–650. https://doi.org/10.1016/j.ecolind.2008.06.003
- Crouzet O, Wiszniowski J, Donnadieu F, Bonnemoy F, Bohatier J, Mallet C (2013) Dose-dependent effects of the herbicide mesotrione on soil cyanobacterial communities. Arch Environ Contam Toxicol 64:23–31. https://doi.org/10.1007/s00244-012-9809-9
- Crouzet O, Consentino L, Petraud JP, Marrauld C, Aguer JP, Bureau S, Le Bourvellec C, Touloumet L, Bérard A (2019) Microalgae mediated soil aggregation in agricultural temperate soils: influence of cropping systems and an herbicide. Front Microbiol 10:1319. <u>https://doi.org/10.3389/fmicb.2019.01319</u>
- Dash NP, Kaushik MS, Kumar A, Abraham G, Singh PK (2018) Toxicity of biocides to native cyanobacteria at different rice crop stages in wetland paddy field. J Appl Phycol 30:483–493. <u>https://doi.org/10.1007/s10811-017-1276-2</u>
- Davies G, Schäfer H, Marshall S, Bramke I, Oliver RG, Bending GD (2013) Light structures phototroph, bacterial and fungal communities at the soil surface. PLoS One 8:e69048. https://doi.org/10.1371/journal.pone.0069048
- Dhawi F (2023) How can we stabilize soil using microbial communities and mitigate desertification? Sustainability 15:863. <u>https://doi.org/10.3390/su15010863</u>
- Djemiel C, Plassard D, Terrat S, Crouzet O, Sauze J, Mondy S, Nowak V, Wingate L, Ogee J, Maron PA (2020) mu green-db: a reference database for the 23S rRNA gene of eukaryotic plastids and cyanobacteria. Sci Rep 10:11. https://doi.org/10.1038/s41598-020-62555-1
- [EFSA] European Food Safety Authority (2010) EFSA panel on plant protection products and their residues (PPR): scientific opinion on the development of specific protection goal options for environmental risk assessment of pesticides, in particular in relation to the revision of the Guidance Documents on Aquatic and Terrestrial Ecotoxicology (SANCO/3268/2001 and SANCO/10329/2002). EFSA J 8:1821 [55 pp.]. https://doi.org/10.2903/j.efsa.2010.1821
- [EFSA] European Food Safety Authority (2016) EFSA Scientific Committee: Guidance to develop specific protection goals options for environmental risk assessment at EFSA, in relation to biodiversity and ecosystem services. EFSA J 14:4499 [50 pp.]. <u>https://doi.org/10.2903/j.efsa.2016.4499</u>
- Faber JH, Marshall S, Brown AR, Holt A, van den Brink PJ, Maltby L (2021) Identifying ecological production functions for use in ecosystem services-based environmental risk assessment of chemicals. Sci Total Environ 791:146409. <u>https://doi.org/10.1016/j.scitotenv.2021.146409</u>
- Foets J, Stanek-Tarkowska J, Teuling AJ, Van de Vijver B, Wetzel CE, Pfister L (2021) Autecology of terrestrial diatoms under anthropic disturbance and across climate zones. Ecol Indic 122:16. https://doi.org/10.1016/j.ecolind.2020.107248
- Forbes VE, Calow P (2013) Use of the ecosystem services concept in ecological risk assessment of chemicals. Integr Environ Assess Manag 9:269–275. <u>https://doi.org/10.1002/ieam.1368</u>
- Gaydon DS, Probert ME, Buresh RJ, Meinke H, Timsina J (2012) Modelling the role of algae in rice crop nutrition and soil organic carbon maintenance. Eur J Agron 39:35–43. <u>https://doi.org/10.1016/j.eja.2012.01.004</u>
- Giora J, Kidron GJ, Wang Y, Herzberg M (2020) Exopolysaccharides may increase biocrust rigidity and induce runoff generation. J Hydrol 588:125081. <u>https://doi.org/10.1016/j.jhydrol.2020.125081</u>
- Haines-Young R, Potschin MB (2018) Common international classification of ecosystem services (CICES) V5.1 and guidance on the application of the revised structure. <u>http://www.cices.eu</u> Accessed 5 May 2023
- Hayes F, Spurgeon DJ, Lofts S, Jones L (2018) Evidence-based logic chains demonstrate multiple impacts of tracemetalsonecosystemservices.JEnvironManag223:150–164.<a href="https://doi.org/10.1016/j.jenvman.2018.05.053">https://doi.org/10.1016/j.jenvman.2018.05.053</a>

- Hellal J, Barthelmebs L, Bérard A et al (2023) Unlocking secrets of microbial ecotoxicology: recent achievements and future challenges. FEMS Microbiol Ecol 99:1–21. <u>https://doi.org/10.1093/femsec/fiad102</u>
- Jassey VEJ, Walcker R, Kardol P, Geisen S, Heger T, Lamentowicz M, Hamard S, Lara E (2022) Contribution of soil algae to the global carbon cycle. New Phytol 234:64–76. <u>https://doi.org/10.1111/nph.17950</u>
- Joly P, Misson B, Perrière F, Bonnemoy F, Joly M, Donnadieu-Bernard F, Aguer JP, Bohatier J, Mallet C (2014) Soil surface colonization by phototrophic indigenous organisms in two contrasted soils treated by formulated maize herbicide mixtures. Ecotoxicology 23:1648–1658. <u>https://doi.org/10.1007/s10646-014-1304-9</u>
- Kaushik MS, Kumar A, Abraham G, Dash NP, Singh PK (2019) Field evaluations of agrochemical toxicity to cyanobacteria in rice field ecosystem: a review. J Appl Phycol 31:471–489. <u>https://doi.org/10.1007/s10811-018-1559-2</u>
- Keqiang Z, Zijia Z, Cui Z, Ling X, Delong M, Li W, Shaoxian S, Torres Sancheze RM, Farias M (2023) Rapid artificial biocrust development by cyanobacterial inoculation and clay amendment. Land Degrad Dev 34:3728–3743. <u>https://doi.org/10.1002/ldr.4716</u>
- Kidron GJ (2021) The role of biocrust-induced exopolymeric matrix in runoff generation in arid and semiarid zones a mini review. J Hydrol Hydromech 69:360–368. <u>https://doi.org/10.2478/johh-2021-0028</u>
- Kidron GJ, Drahorad SL (2021) Biocrust-induced extracellular polymeric substances are responsible for dune stabilization in the Negev. Land Degrad Dev 33:425–438. <u>https://doi.org/10.1002/ldr.4146</u>
- Klaus J, Wetzel CE, Martínez-Carreras N, Ector L, Pfister L (2015) A tracer to bridge the scales: on the value of diatoms for tracing fast flow path connectivity from headwaters to meso-scale catchments. Hydrol Process 29:5275–5289. <u>https://doi.org/10.1002/hyp.10628</u>
- Knapen A, Poesen J, Galindo-Morales P, De Baets S, Pals A (2007) Effects of microbiotic crusts under cropland in temperate environments on soil erodibility during concentrated flow. Earth Surf Process Landf 32:1884–1901. <u>https://doi.org/10.1002/esp.1504</u>
- Lentendu G, Wubet T, Chatzinotas A, Wilhelm C, Buscot F, Schlegel M (2014) Effects of long-term differential fertilization on eukaryotic microbial communities in an arable soil: a multiple barcoding approach. Mol Ecol 23:3341–55. <u>https://doi.org/10.1111/mec.12819</u>
- Lips S, Larras F, Schmitt-Jansen M (2022) Community metabolomics provides insights into mechanisms of pollution-induced community tolerance of periphyton. Sci Total Environ 824:153777. https://doi.org/10.1016/j.scitotenv.2022.153777
- Malam Issa O, Défarge C, Le Bissonnais Y, Marin B, Duval O, Bruand A, D'Acqui LP, Nordenberg S, Annerman M (2007) Effects of the inoculation of cyanobacteria on the microstructure and the structural stability of a tropical soil. Plant Soil 290:209–219. <u>https://doi.org/10.1007/s11104-006-9153-9</u>
- Maltby L, Brown R, Faber JH, Galic N, Van den Brink PJ, Warwick O, Marshall S (2021) Assessing chemical risk within an ecosystem services framework: implementation and added value. Sci Total Environ 791:148631. https://doi.org/10.1016/j.scitotenv.2021.148631
- Maltby L, Brown R, Wilkinson H (2022) Applying ecosystem services principles to the derivation of freshwater environmental quality standards. Front Environ Sci 10:932161. <u>https://doi.org/10.3389/fenvs.2022.932161</u>
- Mamy L, Pesce S, Sanchez W et al (2022) Impacts des produits phytopharmaceutiques sur la biodiversité et les services écosystémiques. Rapport de l'expertise scientifique collective. [Rapport de recherche] INRAE; IFREMER. 2022, p 1408. <u>https://doi.org/10.17180/0gp2-cd65</u>
- Martínez-Carreras N, Wetzel CE, Frentress J, Ector L, McDonnell JJ, Hoffmann L, Pfister L (2015) Hydrological connectivity inferred from diatom transport through the riparian-stream system. Hydrol Earth Syst Sci 19:3133–3151. <u>https://doi.org/10.5194/hess-19-3133-2015</u>
- Menyhart L, Nagy S, Lepossa A (2018) Rapid analysis of photoautotroph microbial communities in soils by flow cytometric barcoding and fingerprinting. Appl Soil Ecol 130:237–240. https://doi.org/10.1016/j.apsoil.2018.06.013
- Metting B (1981) The systematics and ecology of soil algae. Bot Rev 47:195–312. https://doi.org/10.1007/BF02868854
- Metting B (1987) Dynamics of wet and dry aggregate stability from a three-year microalgal soil conditioning experiment in the field. Soil Sci 143:139–143. <u>https://doi.org/10.1097/00010694-198702000-00009</u>
- Moisset S, Tiam SK, Feurtet-Mazel A, Morin S, Delmas F, Mazzella N, Gonzalez P (2015) Genetic and physiological responses of three freshwater diatoms to realistic diuron exposures. Environ Sci Pollut 22:4046–4055. https://doi.org/10.1007/s11356-014-3523-2
- Nienstedt KM, Brock T, Jv W, Montforts M, Hart A, Aagaard A, Alix A, Boesten J, Bopp SK, Brown C, Capri E, Forbes V, Köpp H, Liess M, Luttik R, Maltby L, Sousa JP, Streissl F, Hardy AR (2012) Development of a framework based on an ecosystem services approach for deriving specific protection goals for environmental risk assessment of pesticides. Sci Total Environ 415:31–38. <u>https://doi.org/10.1016/j.scitotenv.2011.05.057</u>

- Osman MEAH, Abo-Shady AM, Gaafar RM, Ismail GA, El-Nagar MMF (2023) Assessment of cyanobacteria and tryptophan role in the alleviation of the toxic action of brominal herbicide on wheat plants. Gesunde Pflanzen 75:785–799. <u>https://doi.org/10.1007/s10343-022-00785-1</u>
- Peng X, Bruns MA (2019) Development of a nitrogen-fixing cyanobacterial consortium for surface stabilization of agricultural soils. J Appl Phycol 31:1047–1056. <u>https://doi.org/10.1007/s10811-1597-9</u>
- Pesce S, Mamy L, Achard AL, Le Gall M, Le Perchec S, Réchauchère O, Leenhardt S, Sanchez S (2021) Collective scientific assessment as a relevant tool to inform public debate and policymaking: an illustration with the effects of plant protection products on biodiversity and ecosystem services. Environ Sci Pollut Res 28:38448– 38454. <u>https://doi.org/10.1007/s11356-021-14863-w</u>
- Pesce S, Bérard A, Coutellec M-A et al (2023a) Linking ecotoxicological effects on biodiversity and ecosystem functions to impairment of ecosystem services is a challenge: an illustration with the case of plant protection products. Environ Sci Pollut Res. <u>https://doi.org/10.1007/s11356-023-29128-x</u>
- Pesce S, Mamy L, Sanchez W et al (2023b) Main conclusions and perspectives from the collective scientific assessment of the effects of plant protection products on biodiversity and ecosystem services along the land–sea continuum in France and French overseas territories. Environ Sci Pollut Res. https://doi.org/10.1007/s11356-023-26952-z
- Pfister L, Wetzel CE, Klaus J, Martinez-Carreras N, Antonelli M, Teuling AJ, McDonnell JJ (2017) Terrestrial diatoms as tracers in catchment hydrology: a review. Wires Water 4:13. <u>https://doi.org/10.1002/wat2.1241</u>
- Pipe A (1992) Pesticide effects on soil algae and cyanobacteria. Rev Environ Contam Toxicol 127:95–170. https://doi.org/10.1007/978-1-4613-9751-9\_4
- Poveda J (2021) Cyanobacteria in plant health: biological strategy against abiotic and biotic stresses. Crop Prot 141:105450. <u>https://doi.org/10.1016/j.cropro.2020.105450</u>
- Ramakrishnan B, Raju MN, Venkateswarlu K, Megharaj M (2023) Potential of microalgae and cyanobacteria in improving soil health and agricultural productivity - a critical view. Environ Sci : Adv. <u>https://doi.org/10.1039/D2VA00158F</u>
- Righini H, Francioso O, Martel Quintana A, Roberti R (2022) Cyanobacteria: a natural source for controlling agricultural plant diseases caused by fungi and oomycetes and improving plant growth. Horticulturae 8:58. https://doi.org/10.3390/horticulturae8010058
- Rivera SF, Vasselon V, Bouchez A, Rimet F (2020) Diatom metabarcoding applied to large scale monitoring networks: optimization of bioinformatics strategies using Mothur software. Ecol Indic 109:13. https://doi.org/10.1016/j.ecolind.2019.105775
- Rodriguez-Caballero E, Knerr T, Weber B (2015) Importance of biocrusts in dryland monitoring using spectral indices. Remote Sens Environ 170:32–39. <u>https://doi.org/10.1016/j.rse.2015.08.034</u>
- Rossi F, Potrafka RM, Pichel FG, de Philippis R (2021) The role of the exopolysaccharides in enhancing hydraulic conductivity of biological soil crusts. Soil Biol Biochem 46:33–40. https://doi.org/10.1016/j.soilbio.2011.10.016
- Rumschlag SL, Mahon MB, Hoverman JT, Raffel TR, Carrick HJ, Hudson PJ, Rohr JR (2020) Consistent effects of pesticides on community structure and ecosystem function in freshwater systems. Nat Commun 11:6333. https://doi.org/10.1038/s41467-020-20192-2
- Santini G, Biondi N, Rodolfi L, Tredici MR (2021) Plant biostimulants from cyanobacteria: an emerging strategy to improve yields and sustainability in agriculture. Plants-Basel 10:22. <u>https://doi.org/10.3390/plants10040643</u>
- Sauze J, Ogée J, Maron PA, Nowak V, Wohl S, Kaisermann A, Jones SP, Crouzet O, Wingate L (2017) The interaction of soil phototrophs and fungi with pH and their impact on soil CO2, CO18O and OCS exchange. Soil Biol Biochem 115:371–382. <u>https://doi.org/10.1016/j.soilbio.2017.09.009</u>
- Shimmel KM, Darley WM (1985) Productivity and density of soil algae in an agricultural system. Ecology 66:1439–1447. <u>https://doi.org/10.2307/1938006</u>
- Singh AK, Singh PP, Tripathi V, Verma H, Singh SK, Srivastava AK, Kumar A (2018) Distribution of cyanobacteria and their interactions with pesticides in paddy field: a comprehensive review. J Environ Manag 224:361–375. https://doi.org/10.1016/j.jenvman.2018.07.039
- Vonk JA, Kraak MHS (2020) Herbicide Exposure and Toxicity to Aquatic Primary Producers. In: DeVoogt P (ed) Reviews of environmental contamination and toxicology, vol 250. Springer International Publishing Ag, Cham, pp 119–171. <u>https://doi.org/10.1007/398\_2020\_48</u>
- Wegener KE, Aldag R, Meyer B (1985) Soil algae: effects of herbicides on growth and C2H2 reduction (nitrogenase) activity. Soil Biol Biochem 17:641–644. <u>https://doi.org/10.1016/0038-0717(85)90041-0</u>
- Zaady E, Arbel S, Barkai D, Sarig S (2013) Long-term impact of agricultural practices on biological soil crusts and their hydrological processes in a semiarid landscape. J Arid Environ 90:5–11. https://doi.org/10.1016/j.jaridenv.2012.10.021
- Zancan S, Trevisan R, Paoletti MG (2006) Soil algae composition under different agro-ecosystems in North-Eastern Italy. Agr Ecosys Environ 112:1–12. <u>https://doi.org/10.1016/j.agee.2005.06.018</u>

- Zhang J, Li C, Yin Y, Zhang J, Grzegorzek M (2022) Applications of artificial neural networks in microorganism image analysis: a comprehensive review from conventional multilayer perceptron to popular convolutional neural network and potential visual transformer. Artif Intell Rev 56:1013–1070. https://doi.org/10.1007/s10462-022-10192-7
- Zhang J, Vieira DN, Cheng Q et al (2023) DiatomNet v1.0: A novel approach for automatic diatom testing for drowning diagnosis in forensically biomedical application. Comput Methods Programs Biomed 232:107434. https://doi.org/10.1016/j.cmpb.2023.107434
- Zhao Y, Wang N, Zhang Z, Pan Y, Jia R (2021) Accelerating the development of artificial biocrusts using covers for restoration of degraded land in dryland ecosystems. Land Degrad Dev 32:285–295. https://doi.org/10.1002/ldr.3714

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