

Impacts of neonicotinoids on biodiversity: a critical review

Laure Mamy, Stéphane Pesce, Wilfried Sanchez, Stéphanie Aviron, Carole Bedos, Philippe Berny, Colette Bertrand, Stéphane Betoulle, Sandrine Charles, Arnaud Chaumot, et al.

▶ To cite this version:

Laure Mamy, Stéphane Pesce, Wilfried Sanchez, Stéphanie Aviron, Carole Bedos, et al.. Impacts of neonicotinoids on biodiversity: a critical review. Environmental Science and Pollution Research, 2024, 31 (1), pp.90-108. 10.1007/s11356-023-31032-3. hal-04321892

HAL Id: hal-04321892 https://hal.inrae.fr/hal-04321892

Submitted on 4 Dec 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Impacts of neonicotinoids on biodiversity: a critical review

2 Environmental Science and Pollution Research 3 DOI: 10.1007/s11356-023-31032-3 4 5 Published: 01 December 2023 6 Special Issue "Key learnings from a collective scientific assessment on the effects of plant 7 protection products on biodiversity and ecosystem services along the land to sea continuum" 8 9 Laure Mamy¹ · Stéphane Pesce² · Wilfried Sanchez³ · Stéphanie Aviron⁴ · Carole Bedos¹ · Philippe Berny⁵ · Colette 10 Bertrand¹ · Stéphane Betoulle⁶ · Sandrine Charles⁷ · Arnaud Chaumot² · Michael Coeurdassier⁸ · Marie-Agnès 11 $Coutellec^9 \cdot Olivier \ Crouzet^{10} \cdot Juliette \ Fabur\'e^1 \cdot Cl\'ementine \ Fritsch^8 \cdot Patrice \ Gonzalez^{11} \cdot Mickael \ Hedde^{12} \cdot Fabur\'e^1 \cdot Cl\'ementine \ Fritsch^8 \cdot Patrice \ Gonzalez^{11} \cdot Mickael \ Hedde^{12} \cdot Fabur\'e^1 \cdot Cl\'ementine \ Fritsch^8 \cdot Patrice \ Gonzalez^{11} \cdot Mickael \ Hedde^{12} \cdot Fabur\'e^1 \cdot Fabur$ 12 13 Christophe Leboulanger¹³ · Christelle Margoum² · Christian Mougin¹ · Dominique Munaron¹³ · Sylvie Nélieu¹ · $C\'{e} line Pelosi^{14} \cdot Magali \ Rault^{15} \cdot Elliott \ Sucr\'{e}^{13,16} \cdot Marielle \ Thomas^{17} \cdot Julien \ Tournebize^{18} \cdot Anne-Laure \ Achard^{19}$ 14 15 · Morgane Le Gall²⁰ · Sophie Le Perchec²¹ · Sophie Leenhardt²² 16 17 Corresponding author: Laure Mamy (laure.mamy@inrae.fr) 18 19 ¹ Université Paris-Saclay, INRAE, AgroParisTech, UMR ECOSYS, 91120 Palaiseau, France 20 ² INRAE, UR RiverLy, 69625 Villeurbanne, France 21 ³ Ifremer, Direction Scientifique, 34200 Sète, France 22 ⁴ INRAE, Institut Agro, ESA, UMR BAGAP, 35042, Rennes, France 23 ⁵ UR ICE Vetagro Sup, Campus Vétérinaire, 69280 Marcy-L'Etoile, France 24 ⁶ Université de Reims Champagne-Ardenne, Normandie Université, ULH, INERIS, SEBIO, 51100 Reims, France 25 ⁷ Université Lyon 1, UMR CNRS 5558, 69100 Villeurbanne, France 26 8 Laboratoire Chrono-Environnement, UMR 6249 CNRS - Université de Franche-Comté, 25000 Besançon, France 27 ⁹ DECOD (Ecosystem Dynamics and Sustainability), INRAE, L'Institut Agro, Ifremer, 35042 Rennes, France

¹⁰ OFB, Direction de la Recherche et Appui Scientifique (DRAS), 78610, Auffargis, France

28

- 29 ¹¹ Univ. Bordeaux, CNRS, Bordeaux INP, EPOC, UMR 5805, 33600 Pessac, France
- 30 12 Eco&Sols, Univ. Montpellier, INRAE, IRD, CIRAD, Institut Agro Montpellier, 34060 Montpellier, France
- 31 ¹³ MARBEC, Univ Montpellier, CNRS, Ifremer, IRD, 34200 Sète, France
- 32 ¹⁴ INRAE, Avignon Université, UMR EMMAH, 84000, Avignon, France
- 33 ¹⁵ Université d'Avignon, Université Aix-Marseille, CNRS, IRD, IMBE, Pôle Agrosciences, 84916 Avignon,
- 34 France
- 35 le Centre Universitaire de Formation Et de Recherche de Mayotte (CUFR), 97660 Dembeni, Mayotte, France
- 36 ¹⁷ Université de Lorraine, INRAE, UR AFPA, 54000 Nancy, France
- 37 ¹⁸ Université Paris-Saclay, INRAE, UR HYCAR, 92160 Antony, France
- 38 ¹⁹ INRAE, AQUA Division, IST, 69625 Villeurbanne, France
- 39 ²⁰ Ifremer, Information Scientifique et Technique, Bibliothèque La Pérouse, 29280 Plouzané, France
- 40 ²¹ INRAE, DipSO, UAR1266, 35042 Rennes, France
- 41 ²² INRAE, DEPE, 75338 Paris Cedex 07, France

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

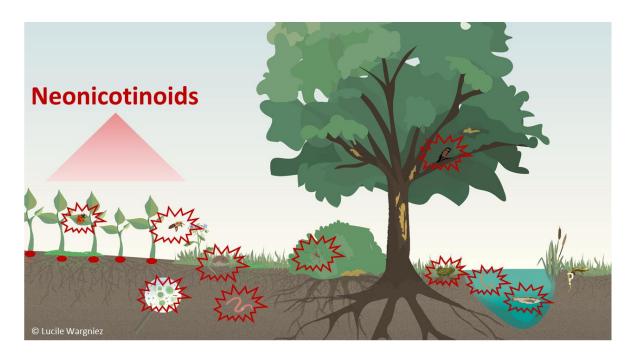
42

Abstract

Neonicotinoids are the most widely used class of insecticides in the world but they have raised numerous concerns regarding their effects on biodiversity. Thus, the objective of this work was to do a critical review of the contamination of the environment (soil, water, air, biota) by neonicotinoids (acetamiprid, clothianidin, imidacloprid, thiacloprid, thiamethoxam) and of their impacts on terrestrial and aquatic biodiversity. Neonicotinoids are very frequently detected in soils and in freshwater, and they are also found in the air. They have only been recently monitored in coastal and marine environments, but some studies already reported the presence of imidacloprid and thiamethoxam in transitional or semi-enclosed ecosystems (lagoons, bays and estuaries). The contamination of the environment leads to the exposure and to the contamination of non-target organisms, and to negative effects on biodiversity. Direct impacts of neonicotinoids are mainly reported on terrestrial invertebrates (e.g., pollinators, natural enemies, earthworms) and vertebrates (e.g., birds), and on aquatic invertebrates (e.g., arthropods). Impacts on aquatic vertebrate populations and communities, as well as on microorganisms, are less documented. In addition to their toxicity to directly exposed organisms, neonicotinoid induce indirect effects via trophic cascades as demonstrated in several species (terrestrial and aquatic

invertebrates). However, more data are needed to reach firmer conclusions and to get a clearer picture of such indirect effects. Finally, we identified specific knowledge gaps that need to be filled to better understand the effects of neonicotinoids on terrestrial, freshwater and marine organisms, as well as on ecosystem services associated with these biotas.

Graphical abstract



Keywords Pesticides · Plant protection products · Ecotoxicity · Ecotoxicology · Agrosystems · Collective scientific assessment

Introduction

Neonicotinoids are systemic insecticides (i.e., they diffuse throughout the treated plants to protect them from pests) that act on the central nervous system of insects by targeting nicotinic acetylcholine receptors (nAChRs) in the brain (Simon-Delso et al. 2015; Thompson et al. 2020). They are the world's fastest-growing and currently the most widely used class of insecticides against a broad spectrum of sucking and chewing insects (plant hoppers, thrips, micro-lepidopteras), and they are also involved in veterinary medicine (e.g., against fleas in pets) and in biocidal products such as those used for the treatment of livestock buildings or in pest baits for domestic use (Klingelhöfer et al. 2022; Thompson et al. 2020). In agriculture, neonicotinoids are mainly applied through seed

treatments, but they are also employed as granular application, spraying or soil treatment (Simon-Delso et al. 2015; Thompson et al. 2020). The five most used active substances are acetamiprid, clothianidin, imidacloprid, thiacloprid and thiamethoxam (clothianidin is also the main transformation product of thiamethoxam). Among these substances, only acetamiprid is still approved in the European Union (EU Pesticides database 2023). Clothianidin and thiamethoxam were withdrawn in 2019, while imidacloprid and thiacloprid were withdrawn in 2020 (European Commission 2023). However, for example in France, derogations have been granted in 2021 and 2022 for the use of coated seeds treated with imidacloprid or thiamethoxam in the context of the infestation of beet crops by aphids (JORF 2021; JORF 2022). Consequently, because of their wide use all over the world, and because of the high persistence of clothianidin, imidacloprid and thiamethoxam (average half-life in soils is 121 days for thiamethoxam (PPDB 2023), 187 days for imidacloprid (PPDB 2023) and 545 days for clothianidin (PPDB 2023) which could reach 20 years (Thompson et al. 2020)), neonicotinoids are likely to be ubiquitous in the environment, and present a potential environmental health concern (Bonmatin et al. 2015; Goulson 2013; Humann-Guilleminot et al. 2019a; Morrissey et al. 2015).

Neonicotinoids were first presented as having key attributes such as systemic nature, versatility in application (especially as seed treatments), selective toxicity to arthropods, lower binding efficiencies to vertebrate compared to invertebrate receptors, and assumed lower impacts on non-target aquatic and terrestrial organisms (Simon-Delso et al. 2015; Thompson et al. 2020). Neonicotinoids should also theoretically not target organisms lacking nAChRs and thus nervous systems, such as protists, fungi, prokaryotes and plants (Simon-Delso et al. 2015).

However, neonicotinoids appeared to have lethal and sublethal effects on non-target organisms, including pollinators, insect predators and vertebrates (especially birds) (Alsafran et al. 2022; Mineau and Kern 2023; Mineau and Palmer 2013; Simon-Deslo et al. 2015). Thus, for many years, the use of neonicotinoid-based products in agriculture has raised concerns in several countries, particularly because of their effects on pollinators (Demortain 2021; Suryanarayanan 2013), and EFSA (2018) concluded that most uses of neonicotinoid substances do represent a risk to wild bees and honeybees. In addition, as more than 80% of neonicotinoid seed treatments can remain in the soil (Alford and Krupke 2017; Sur and Stork 2003), soil invertebrates may be exposed to high doses of neonicotinoids, with recognized lethal and sublethal effects (Gunstone et al. 2021). Neonicotinoids also contaminate freshwater ecosystems worldwide and could impact aquatic invertebrates, over broad spatial scales (Cavallaro et al. 2019; Hallmann et al. 2014; Morrissey et al. 2015). Moreover, they were demonstrated to exert

negative effects on terrestrial and aquatic vertebrates (Gibbons et al. 2015; Thompson et al. 2020; Wood and Goulson 2017).

In this context, the objective of this work was to do a critical review of (1) the contamination of the environment (soil, water, air, biota) by neonicotinoids and (2) their impacts on terrestrial and aquatic biodiversity. Although the literature focused on the ecotoxicological effects of neonicotinoids is abundant, to the best of our knowledge, no review has been published on the overall impacts of these substances on the whole biodiversity.

Bibliographic corpus

The review of the literature on the impacts of neonicotinoids on biodiversity was performed under the framework of a French collective scientific assessment focused on the impacts of plant protection products (PPPs) on biodiversity and ecosystem services (Pesce et al. 2023). Collective scientific assessment seeks to inform public policy and to foster public debate by analyzing the literature, but it is neither a meta-analysis nor a systematic review (Pesce et al. 2021). Though not quantitative, this review gives a detailed and complete overview of the impacts of neonicotinoids on the whole biodiversity.

In this framework, the bibliographic corpus was adapted and constructed as follows: six queries (Q) focused on neonicotinoids (Q1), ecotoxicology (Q2), biodiversity (Q3), terrestrial ecosystems (Q4), freshwater ecosystems (Q5) and marine ecosystems (Q6) were defined with related keywords (Table SI1). The literature search was conducted on the Web of ScienceTM, from 2000 to 2020.

The corpus of publications was then built by combining Q1 with Q2, Q3, Q4, Q5 or Q6. The combination of Q1*Q2 yielded 7349 references; that of Q1*Q3, 457 references; Q1*Q4, 3309 references; Q1*Q5, 841 references; and Q1*Q6, 252 references. After removing duplicates, the total number of references was 7697.

The time course of the 7697 references showed a strong increase in the number of publications related to the impacts of neonicotinoids from 2000 to 2020 (Fig. 1). Among the five neonicotinoids retained in this review, imidacloprid was the most studied one (4218 occurrences in titles and abstracts), well above thiamethoxam (1672), acetamiprid (1176), clothianidin (887) and thiacloprid (674) (Fig. 2). The bibliometric measurements also demonstrated that terrestrial invertebrates were the most studied organisms and especially honeybees (Fig. 3). Apart from terrestrial invertebrates, fish come at the thirty second place (Fig. 3). In the first 35 occurrences, there are no other taxonomic group.

The categorization of references was based on titles and abstracts. The selected corpus was then divided according to the expertise of the different authors who proceeded to in-depth analysis of each reference. The

literature search was focused on the most integrative and ecologically realistic studies as possible. The results of single-species tests were not systematically reviewed, and were only used if they provided explanatory elements for processes observed under realistic environmental conditions.

The corpus was finally manually completed by various documents, papers and books known to the authors and which were not present in the 7697 references, and over time until April 2023. At the end, a total of 308

142

143

144

145

146

136

137

138

139

140

141

Terrestrial ecosystems

Contamination of soils, plants and air

publications were retained and cited in this work.

Neonicotinoids are found in all environments: soil, water (see section "Contamination of freshwater and marine

environments" below), plants and air.

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

Contamination of soils

Soil contamination by neonicotinoids has been studied under various climates, soil types, and agricultural practices (Table 1). A large study conducted on 74 French cultivated soils showed that imidacloprid (limit of quantification LOQ = 1 μg/kg) was present in 91% of the soil samples (excluding seven organically grown soils, with no detectable traces) although only 15% of the sites had been planted with treated seeds the year of the monitoring (Bonmatin et al. 2005a). In addition, imidacloprid was detected in 100% of the soils which received treated seeds (corn, wheat or barley) during the sampling year, and in 97% of the soils which received the same treatment one or two years before the study. Concentrations were higher in the soils which had been treated consecutively during two years before the monitoring than in those that received treated seeds only one year before, indicating that imidacloprid accumulates in soils over time. Silva et al. (2019) found that imidacloprid was present in 7% of the examined European topsoil samples (LOQ = 10 µg/kg, one order of magnitude higher than the above study) with a maximum content of 60 µg/kg, while Pelosi et al. (2021) found imidacloprid in 90 % of French sampled soils (n=180, 26 % when considering concentrations >10 μg/kg, LOQ = 0.4 μg/kg) and concentrations reaching 160 μg/kg (Table 1). Thiamethoxam was present in 20% of the French soils at low concentrations (maximum of 2 $\mu g/kg$, $LOQ = 0.4 \mu g/kg$) (Pelosi et al. 2021). In Switzerland, imidacloprid ($LOQ = 0.9 \ 10^{-3} \mu g/kg$) was quantified in 94% of cultivated field soils (n=82) and in 71% of ecological focus area soils (annual, biennial and perennial herbaceous plant species; n=68) (Humann-Guilleminot et al. 2019a). Clothianidin (LOQ = 1.6 10⁻³ μg/kg) was

also frequently observed in the sampled soils (77% of cultivated fields and 46% of ecological focus areas); followed by thiacloprid (LOQ = $1.6\ 10^{-3}\ \mu\text{g/kg}$; 28% and 13%), thiamethoxam (LOQ = $1.9\ 10^{-3}\ \mu\text{g/kg}$; 27% and 6%) and acetamiprid (LOQ = 2.0 10⁻³ μg/kg; 13% and 3%) (Humann-Guilleminot et al. 2019a). Similarly, Riedo et al. (2021) repeatedly observed imidacloprid (59% of soils, maximum concentration of 24 µg/kg, LOQ = 0.14 μ g/kg), clothianidin (55%, 57 μ g/kg, LOQ = 0.15 μ g/kg), thiamethoxam (21%, 24 μ g/kg, LOQ =0.15 μ g/kg) and thiacloprid (10%, 14 µg/kg, LOQ = 0.073 µg/kg) in various Swiss agricultural soils (Table 1). The highest concentration of imidacloprid in Switzerland was measured by Chiaia-Hernandez et al. (2017) and was found to be 138 µg/kg (LOQ = 3 µg/kg) (Table 1). Recently, Froger et al. (2023) monitored 111 PPP residues (48 fungicides, 36 herbicides, 25 insecticides and/or acaricides, and two safeners) in 47 soils sampled across France under various land uses (arable lands, vineyards, orchards, forests, grasslands, brownfields). The most frequently quantified neonicotinoid was clothianidin (17% of the soil samples, maximum concentration of 2.7 μ g/kg, LOQ = 0.5 μ g/kg) followed by imidacloprid (9%, 13.8 μ g/kg, LOQ = 2 μ g/kg), thiacloprid (6%, 0.26 μ g/kg, LOQ = 0.05 μ g/kg) and acetamiprid (2%, $0.48 \mu g/kg$, LOQ = $0.01 \mu g/kg$) (Table 1). Thiamethoxam was not quantified (LOQ = $0.5 \mu g/kg$). In English arable soils, where neonicotinoids have been used as seed treatments, the concentrations of clothianidin ranged from < 0.02 to $13.6 \,\mu\text{g/kg}$ (LOQ = $0.02 \,\mu\text{g/kg}$), that of imidacloprid from < 0.09 to $10.7 \,\mu\text{g/kg}$ (LOQ = $0.09 \,\mu\text{g/kg}$). μ g/kg) and that of thiamethoxam from < 0.02 to 1.5μ g/kg (LOQ = 0.02μ g/kg) (Jones et al. 2014). Overall, most of the reviewed works focusing on the presence of neonicotinoids in soils is centered on imidacloprid, while the other substances are much less targeted. The environmental conditions, crops, agricultural practices, analytical methods and sampling time and strategies may explain the differences observed between the reviewed studies but, in general, they show the ubiquitous contamination of soils by neonicotinoids (Bonmatin et al. 2015; Froger et al. 2023).

185186

187

188

189

190

191

192

193

194

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

Contamination of plants

Neonicotinoids enter plants through the roots and/or leaves, and are transported into various organs, including foliage, flowers, pollen and nectar (Bonmatin et al. 2015). They are frequently detected in cultivated plants, as well as in wild plants. Bonmatin et al. (2005b) measured imidacloprid concentrations in corn whose seeds have been treated with this insecticide and observed that 76% of stem and leaf samples at flowering contained more than 1 μ g/kg of the substance (LOQ = 0.1 μ g/kg). They also quantified from 1 to 10 μ g/kg of imidacloprid in sunflower flower heads, with notable variations depending on crop stage and seed variety. In the 29 analyzed samples of sunflower pollens, only two contained traces of imidacloprid. In parallel, imidacloprid was detected in

untreated sunflower heads grown on soil treated in previous years (from 0.1 to $2 \mu g/kg$). In sugar beet crop treated with 90 g/ha of imidacloprid as seed coating, the concentration of imidacloprid in leaves initially reached 12.4 mg/kg (fresh weight), then decreased but remained above 1 mg/kg 80 days after sowing, and was below the limit of detection (LOD = $10 \mu g/kg$) at harvest (Rouchaud et al. 1994). Humann-Guilleminot et al. (2019a) analyzed imidacloprid, clothianidin, thiamethoxam, thiacloprid and acetamiprid in plant samples taken from 79 cultivated fields (mainly from cereals and beetroots, but also from potatoes, rapeseed, maize, peas and flax) and 69 ecological focus areas over Switzerland. The neonicotinoids were detected in 97% of plant samples taken in cultivated fields, and in 93% of plant samples from ecological focus areas. The most frequently detected substance was imidacloprid (87% in cultivated fields and 84% in ecological focus areas), followed by thiacloprid (43% and 59%), clothianidin (39% and 12%), acetamiprid (34% and 45%) and thiamethoxam (19% and 7%).

Neonicotinoid residues were also detected in various wildflowers present in non-treated area surrounding crops grown from treated seeds, with residues in foliage ranging from 0.06 to 106 µg/kg (LOQ ranged from 0.06 to 0.60 µg/kg) (Botias et al. 2015; Botias et al. 2016). The authors pointed that these residues may overlap with lethal toxicity levels for some insect species (e.g., *Aphis glycines*). In addition, the widespread contamination of wild plants in agricultural landscape likely increases the exposure duration of pollinators though it is often supposed to be restricted to the crop flowering time (Botias et al. 2015).

Finally, in guttation droplets, potentially consumed by non-target species, works conducted in various European countries showed neonicotinoid concentrations of hundreds of mg/L at the emergence of plant, but only of a few µg/L one month after its emergence (Bonmatin et al. 2015; Tapparo et al. 2011).

Contamination of air

Neonicotinoids may also reach the atmosphere. The measurement of their concentrations relies on active air sampling systems and by trapping compounds on a sorbent from which the compounds are extracted and analyzed. Most of the time, the measured concentrations represent the sum of the compounds present in the atmosphere in both particulate and gaseous forms. Désert et al. (2018) monitored PPP concentrations in ambient air samples collected from February 2012 to December 2017 at one rural and six urban sites in the French Provence-Alpes-Côte d'Azur region. Imidacloprid was quantified in four locations, with concentrations higher than 1 ng/m³ (LOD = 0.081 ng/m³), but with a low frequency of quantification (1 to 2% depending on the site). As it was detected both in the rural and urban sampling sites, the authors suggested an atmospheric transport from agricultural areas to cities given the air mass retro-trajectories. In the French Phytatmo database (2023), which synthetizes the data

obtained by the French Approved Air Quality Monitoring Associations (AASQAs) from 2002 up to now, the average imidacloprid concentration, calculated from 18 quantifications, was equal to 0.39 ng/m³, with a maximum of 2.3 ng/m³ (Table 1), which was higher than the range of concentrations reported by Coscollà and Yusà (2016) (from 0.012 to 0.014 ng/m³) or by Raina-Fulton (2015) (from 0.01 to 0.36 ng/m³ in the particulate phase, LOD = 0.0039 ng/m³) in Canada. The analysis of the Phytatmo database (2023) also showed that acetamiprid and thiamethoxam were only detected once, while thiacloprid was found at an average concentration of 0.17 ng/m³ out of 17 quantifications, and at a maximum concentration of 0.47 ng/m³. In Canada, for the particulate phase, Raina-Fulton (2015) and Coscollà and Yusà (2016) reported acetamiprid concentrations of 0.006 ng/m³ and 0.018 ng/m³, respectively, and Raina-Fulton (2015) observed clothianidin concentrations ranging from 0.01 to 0.09 ng/m³.

Impacts on terrestrial biodiversity

Terrestrial heterotrophic microorganisms

Most studies devoted to the effects of neonicotinoids on functional activities and biodiversity of terrestrial heterotrophic microorganisms concerned imidacloprid. Acetamiprid, clothianidin and thiamethoxam were scarcely addressed, while there was no data for thiacloprid.

In laboratory experiments, Cycoń and Piotrowska-Seget (2015a) evaluated the impact of imidacloprid on soil microbial activities in soils spiked at the agricultural dose and at ten times this dose (1 and 10 mg/kg, respectively). At the agricultural dose, imidacloprid decreased microbial respiration, total bacterial count, and dehydrogenase, phosphatase and urease activities after 14 days. However, these effects were transient and the measured microbial functions recovered after 56 days of exposure. At ten times the agricultural dose, imidacloprid decreased the microbial parameters but no recovery was observed after 56 days suggesting irremediable impacts on communities. Consistently, nitrate concentration decreased while ammonium concentration increased, in agreement with the high sensitivity of nitrifying and nitrogen-fixing bacteria to imidacloprid. Under the same experimental conditions, the effect of imidacloprid on the structure of ammonia-oxidizing archea (AOA) and bacteria (AOB) communities was analyzed using Denaturing Gradient Gel Electrophoresis (DGGE) (Cycoń and Piotrowska-Seget, 2015b). At the agricultural dose, imidacloprid did not affect the α diversity of the bacterial communities. However, at ten times the dose, imidacloprid decreased the α diversity of the AOA community in a durable way, and temporarily that of the AOB community. In addition, at the highest dose, imidacloprid decreased nitrification and increased ammonification. To determine the role of the microbial community diversity in the fate and impact of imidacloprid and acetamiprid, Zhang et al. (2017) used soil microcosms cropped with *Brassica*

chinensis L. They showed that the diversity of the microbial community did not affect the amount of imidacloprid or acetamiprid remaining in the soil but, when microbial diversity decreased, the amount of insecticide exported from the soil to the plant increased. Finally, a study conducted on microbial strains isolated from soil and exposed to imidacloprid or thiamethoxam in Petri dishes showed that both neonicotinoids altered the functions of *Klebsellia* sp. strain 19, a phosphate-solubilizing rhizobacterium exhibiting Plant Growth Promoting Rhizobacteria (PGPR) properties (Ahemad and Khan 2011). Thus, these two insecticides could compromise the PGPR activity of microbial inoculant used to decrease crop dependence on chemically derived fertilizers.

In field conditions, soybean imidacloprid treated seeds decreased the number of *Rhizobia* by a factor of three, while the number of *Rhizobia* was not affected after foliar application (Sarnaik et al. 2006). In contrast, regardless of the mode of application, the insecticide had no effect on phosphate solubilizing bacteria (Sarnaik et al. 2006). Li et al. (2018) studied the impact of imidacloprid or clothianidin treated seeds on the wheat rhizosphere microbial communities over nine months. The analysis of 16S rRNA and ITS amplicons generated from soil-extracted DNA revealed changes in the α and β diversities of bacterial and fungal communities during plant development, but did not reveal any change due to seed treatment with each of the two insecticides. Furthermore, under these conditions, no effect of imidacloprid or clothianidin on some biocontrol agents (*Bacillus*, *Pseudomonas*, *Streptomyces*...) was observed in the wheat rhizosphere.

Two studies examined the impact of thiamethoxam on the taxonomic and metabolic diversity of soil bacterial communities using a laboratory setting. In forest land soils spiked with different amounts of thiamethoxam, an altered composition of the community was observed (Yu et al. 2020): the relative abundance of *Gemmatimonadetes* and OD1 decreased when compared to the control while the relative abundance of *Chloroflexi* and *Nitrospirae* increased. On the other hand, the catabolic diversity of the microbial community in soils treated with the lowest dose (0.02 mg/kg) of thiamethoxam was higher than that of the control while it was lower at the highest doses (0.2 mg/kg and 2 mg/kg). Analyzing soil samples from experimental plots where thiamethoxam was applied in field conditions, Filimon et al. (2015) showed that the insecticide only slightly reduced the phosphatase activity but reduced the number of nitrifying bacteria by about 60%.

In general, studies concerning the effects of neonicotinoids on terrestrial heterotrophic microorganisms revealed contradictory results depending on whether they were conducted in the laboratory (often under unrealistic agricultural conditions), showing impacts on the structure and on different microbial activities, or in the field (in more realistic conditions), showing no or very little effect of these substances.

Terrestrial invertebrates

Neonicotinoids have negative impacts on terrestrial invertebrates (pollinators, natural enemies, earthworms...) in agricultural environments despite variable responses depending on the traits and groups considered, as summarized below.

Neonicotinoids are likely to have greater effects on insect pollinators than other insecticides because they are

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

285

286

287

288

Pollinators

systemic insecticides regularly found in pollen, nectar, and other vegetative parts of plants throughout their flowering period (Krupke et al. 2012; Krupke and Long 2015), leading to risks of pollinators exposure via the oral route as well as through contact for a longer period of time. In addition, during their application, neonicotinoids can also contaminate the surrounding environments (Krupke et al. 2012; Krupke and Long 2015). Comparative toxicity studies among the different categories of neonicotinoids are scarce, but Arena and Sgolastra (2014) provided some insights. They showed that nitro-substituted neonicotinoids ("N-nitroguanidines"; including imidacloprid, thiamethoxam or clothianidin) were generally more toxic to pollinators than cyano-substituted neonicotinoids ("N-cyanoamidines"; including acetamiprid or thiacloprid). Honeybees. Exposure of honeybees (Apis mellifera) to neonicotinoids has been repeatedly demonstrated (e.g., Bonmatin et al. 2015; Hladik et al. 2016; Mitchell et al. 2017; Zhang et al. 2023). In pollens sampled in 2002-2003 before spring, summer, autumn and winter, in apiaries located in five French departments, imidacloprid and/or its 6-chloronicotinic acid transformation product were detected in 69% of the 81 samples, and quantified in 13.5% and 34.6% of the samples, respectively (Chauzat et al., 2006). The frequency of detection did not vary much according to the sampling period. This study was then continued until the end of 2005 (Chauzat et al. 2011): imidacloprid was detected in 11.2% of the bees (average concentration of 1.2 µg/kg) and in 40.5% of the pollen samples (0.9 µk/kg), and 6-chloronicotinic acid was detected in 18.7% of the bees (1.0 µg/kg) and in 33% of the pollen (1.2 µg/kg). In different sites cultivated with a corn/rapeseed rotation whose seeds were treated with thiamethoxam (or not), residues of thiamethoxam and clothianidin in pollens were close to the LOQ (1 µg/kg) in both corn and oilseed rape (from 1 to 2 µg/kg), and the amounts in oilseed rape nectar were lower than 1 µg/kg (LOQ = 0.5 µg/kg) (no corn nectar was analyzed) (Pilling et al. 2013). Wiest et al. (2011) detected imidacloprid in 1% of pollen and 2% of honey but nothing in bees sampled from hives located in the French Pays de la Loire region. Thiamethoxam and clothianidin were not detected in any of these samples. The multiple potential exposure pathways and the size of the pollinator activity zone make it challenging to fully identify and quantify the exposure of pollinators to neonicotinoids (van der Sluijs et al. 2013).

In parallel of the awareness raised by exposure data on the possible role of neonicotinoids in the massive decline of insects, honeybees have been the subject of extensive research focused on the toxicological effects of neonicotinoids. Particular concern resulted from studies focused on honeybee behavior which revealed neonicotinoid-induced impairment of memory and learning abilities (Tison et al. 2019; Willemsen and Hailey 2001) because such impairment is likely to affect navigation parameters and the ability to return to the hive (Henry et al. 2012; Henry et al. 2014). With regard to interaction with other factors or stressors, neonicotinoids were found to increase the susceptibility of honeybees to pathogens (*Nosema*) (Grassl et al. 2018; Müller 2018; Pettis et al. 2013; Uhl and Brühl 2019). Furthermore, the effects of neonicotinoids were demonstrated to increase with decrease in temperature: the ability of bees to return to the hive following exposure to thiamethoxam decreased at lower temperatures (< 28°C) (Henry et al. 2014; Monchanin et al. 2019). Finally, neonicotinoids can interact with other PPPs as observed for clothianidin and propiconazole (fungicide) which impact honeybee survival via synergistic effects (Sgolastra et al. 2017).

However, the issue of the effects of neonicotinoids on honeybees has been the subject of much controversy. In their large-scale monitoring study, Rolke et al. (2016) showed that honeybee colonies placed in clothianidin-treated oilseed rape crops exhibited developmental and reproduction performances similar to those of non-exposed colonies. Under the same crop treatment, clothianidin was not found to pose a risk to colonies in terms of health, development, and overwintering success of honeybee colonies (Belsky and Joshi 2020). This result was also found by Rundlöf et al. (2015) for clothianidin-rapeseed treated seed in combination with non-systemic pyrethroid (beta-cyfluthrin) treatments. Conversely, Samson-Robert et al. (2017) observed an increased mortality of honeybee colonies located in environments dominated by clothianidin-treated grain corn. More recently, Schott et al. (2021) demonstrated lethal effects of clothianidin on honeybee larvae, but found short-term resilience of colonies to treatments, which may result from compensation mechanisms (increased brood size). As to adults, seed treatments with clothianidin, thiamethoxam or imidacloprid resulted in increased worker bees mortality, but effects on colony growth were not observed thereafter (Lin et al. 2021). Actually, the effects of neonicotinoids on colony size vary across study areas (Woodcock et al. 2017). Spatial features, such as landscape characteristics and especially landmarks density (landscape elements that are used as visual cues for the orientation of bees), as well as the bee experience in the studied area (e.g., homing experiments carried out with foragers familiar or not with

the release point), influence the performance of individuals and therefore of colonies, which in turn can either limit or exacerbate the neonicotinoid-induced effects (Henry et al. 2014).

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

To go further into toxicity mechanisms and their consequences for bee colony survival, LaLone et al. (2017) built a network of six Adverse Outcome Pathways (AOPs) and used weight of evidence (WoE) evaluation to describe plausible causal relationships between neonicotinoid mechanisms of action (activation of nicotinic acetylcholine receptor as molecular initiating event and downstream molecular, cellular, or organism-level key events) and colony death, as adverse outcome of regulatory concern. However, WoE assessment identified uncertainty, and thereby need for further research, in some upstream-to-downstream key-event relationships (e.g., between mitochrondrial dysfunction and learning/memory, or between role change in the colony and further larval development). Wild bees. Beside works on the emblematic species Apis mellifera, some studies have focused on wild bees. In ground nesting species (Eucera pruinosa), soil treatment with imidacloprid was found to affect reproduction (decreased number of nests and larvae) and pollen consumption whereas no effect was observed with thiamethoxam used as seed treatment (Cucurbita pepo) (Chan and Raine 2021). However, seed treatments may lead to soil contamination, even in fields adjacent to crops and in non-cropped borders, and affect native bee nesting and richness (Main et al. 2020; Rundlöf et al. 2015). In the field, exposure to various neonicotinoids and/or other PPPs have lethal and sublethal effects, as shown for the solitary bee Osmia bicornis: clothianidin or thiamethoxam, used in combination with other insecticides (beta-cyfluthrin) or fungicides (fludioxonil and metalaxyl-M) impaired the reproduction (Woodcock et al. 2017), as did the mixture of thiacloprid and prochloraz (fungicide) (Alkassab et al. 2020), while clothianidin and propiconazole (fungicide) induced mortality (Sgolastra et al. 2017). In a multistress context, the effects of neonicotinoids on wild bees can be exacerbated by food resource limitation (Stuligross and Williams, 2020). Indeed, the diversification of non-crop floral resources can provide complementary resources, counteracting the negative effects of neonicotinoids as shown on O. bicornis reproduction and larval development (Klaus et al. 2021). With regard to other physiological mechanisms underlying population-level responses under field conditions, the negative effects of neonicotinoids observed on Osmia cornuta reproduction (Stuligross and Williams 2020) or at population level (fitness, density; Sandrock et al. 2014) may have a male component (thiamethoxam-altered male fertility; Strobl et al. 2021a) or not (clothianidin unaffected male survival, emergence and reproductive physiology; Strobl et al. 2021b). Using simple

generalized and linear mixed models (GLMM), Stuligross and Willams (2021) demonstrated how past and current

exposure to neonicotinoids profoundly impact both individual reproduction and population growth rate of orchard blue bees (*Osmia lignaria*).

The impact of neonicotinoids on wild social bees of the *Melipona* group is very little studied. However, the meta-analysis of Botina et al. (2020) highlighted lethal effects on both larvae and adults, especially marked for imidacloprid.

In bumblebees, the effects of neonicotinoids were found to be expressed at the organism level (molecular, cellular, and physiological responses; lethal and sublethal effects) as well as at the population level (mortality, altered colony structure and turnover) (Camp and Lehmann 2021). Colonies of *Bombus terrestris* and *Bombus impatiens* exposed to acetamiprid, clothianidin or imidacloprid exhibited lower growth rates and decreased production of new queens (Camp et al. 2020; Rundlöf et al. 2015; Whitehorn et al. 2012). In addition, a suite of effects was observed, including increased mortality of new queens, delayed nest foundation (Wu-Smart and Spivak 2018), acute and chronic effects on worker foraging activity (Gill and Raine 2014), reduced fecundity and brood production (imidacloprid; Laycock et al. 2012), disruption of their flight activity and endurance (imidacloprid; Kenna et al. 2019), and altered queen condition upon overwintering (thiamethoxam and clothianidin; Fauser et al. 2017). Some works also showed that seed treatments affect *Bombus* spp. densities in adjacent fields and in non-cropped borders (Main et al. 2020; Rundlöf et al. 2015). With respect to interactions with other stressors, no synergistic nor additive effects could be detected between neonicotinoids (mixture of thiamethoxam and clothianidin) and the trypanosome parasite *Crithidia bombi* on post hibernation performances (queen survival and body mass) of *B. terrestris* (Fauser et al. 2017).

With a multi-species dynamic Bayesian occupancy model, Woodcock et al. (2016) highlighted the high impact of neonicotinoid seed treatments as use in oilseed rape on the extinction of 62 species of wild bee populations. Their model was spatially and temporally explicit and related population persistence to exposure over a wide time period of 18 years. This paper identifies the need of developing national scale management strategies to support wild bee populations persistence over the long-term.

Butterflies. The impacts of neonicotinoids on lepidopterans are very little investigated, but the few studies addressing this issue underline a critical role of the timing and mode of exposure. In the monarch butterfly (*Danaus plexippus*), exposure of young adults to realistic doses of imidacloprid did not affect oocyte production, but significantly decreased insect longevity, with likely consequences for population development, migration, and overwintering (James 2019). On the contrary, under exposure to clothianidin-treated plants in the larval stage, there was no significant effect on parameters characterizing monarch migration (flight orientation, movement

speed; Wilcox et al. 2021). Using a linear mixed effect random slope model, Gilburn et al. (2015) demonstrated that the populations of 15 butterfly species commonly occurring at farmland sites in England declined due to the use of neonicotinoids.

Overview of the effects of neonicotinoids on pollinators. In 2018, EFSA (2018) confirmed that the use of neonicotinoids causes a risk to wild bees and honeybees. Although results appeared sometimes contradictory, many studies highlighted negative effects of neonicotinoids on pollinators. The contradictions occasionally observed can be explained by several methodological biases (Walters 2016): (1) laboratory experiments consider exposure conditions (in particular doses and durations) to neonicotinoids that are not really representative of those observed in natura in relation to agricultural practices; (2) most of the studies focus on honeybees or bumblebees, whereas susceptibility to insecticides varies greatly among the different groups of pollinators (Lundin et al. 2015; Rundlöf et al. 2015); (3) studies are most often focused on one type of neonicotinoid which makes generalization difficult. Furthermore, there is a need to combine laboratory and field approaches, and to address the effects of neonicotinoids at the sub-individual and individual levels, as well as the consequences for colonies and populations (see LaLone et al. 2017). For example, Henry et al. (2015) showed that the mortality in honeybee colonies near neonicotinoid (thiamethoxam and imidacloprid)-treated oilseed rape fields was higher than in colonies surrounded by less treated fields. However, this effect was not observable at the colony level during and after the flowering period of oilseed rape, because the impact of this loss was buffered by the colonies' demographic regulation response. While very few models exist that are devoted to the effects of neonicotinoids at the bee colony/population levels, this research area appears promising given the difficulty of actually detecting unintended effects of neonicotinoids in the field using conventional risk assessment methods (Lundin et al. 2015). In particular, Henry et al. (2017) advocated the potentialities of mechanistic models in a multiple stressor context. Since then, the honeybee colony model (BEEHAVE, Becher et al. 2014) has been extended to the colony development of bumblebees in a realistic landscape (Becher et al. 2018), and to translate results from standard laboratory studies to relevant parameters and processes for simulating bee colony dynamics (Preuss et al. 2022). On a regulatory point of view, significant efforts have been undertaken at the EU level to improve risk assessment of the effects of neonicotinoid on bees with, among others, the development of the ApisRAM population model (Adriaanse et al. 2023; EFSA PPR Panel 2015; EFSA Scientific Committee et al. 2021).

429

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

430

431

Natural enemies

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

457

458

459

460

Overall, neonicotinoids have negative impacts on natural enemies such as predators (mites, ladybugs) and parasitoids, especially in field crops (Douglas and Tooker 2016). By disrupting prey-predatory and host-parasitoid interactions, neonicotinoid-treated seeds also alter arthropod communities as a whole (Chen et al. 2016; Disque et al. 2019; Dubey et al., 2020). Ants. In Tetramorium caespitum, increased mortality and disruption of locomotion without loss of hunting behavior was observed after exposure to imidacloprid (Penn and Dale 2017). In other ant species (Pogonomyrmex occidentalis, Lasius niger, Lasius flavus), imidacloprid was also found to alter socio-behavioral traits (e.g., foraging, nest building, competition behavior) at environmentally relevant concentrations under experimental exposure (Sappington 2018; Thiel and Kohler 2016). Bugs. Prey consumption was reduced in predatory bugs (Pentatomidae) feeding on herbivorous preys previously exposed to imidacloprid-treated plants, even when prey density increased (lack of a type II functional response) (Resende-Silva et al. 2019). Studies with Orius insidiosus concluded that imidacloprid was moderately to highly toxic when applied as seed treatment, while foliar toxicity showed conflicting results (Naranjo 2001). In *Podisus* nigrispinus predatory bugs, sublethal effects of thiamethoxam treatments resulted in longer larval development, decreased adult body weight and delayed oviposition (Torres et al. 2003). Imidacloprid may also alter the predatory behavior of spined soldier bugs (Podisus maculiventris), with negative consequences in terms of weight gain (Resende-Silva et al. 2019). However, some of these effects were only seen at certain treatment doses (> 0.5 mg/plant) (Torres et al. 2003), and were sometimes transient (Pekar and Kocourek 2004). Carabids. When fed with slugs contaminated with thiamethoxam, Chlaenius tricolor carabid beetles displayed altered mobility twitching and mild motor difficulties, up to partial to extensive paralysis (Douglas et al. 2015). Forficulidae. As dominant earwig species in temperate orchards, Forficula auricularia is the most studied forficulidae species in the laboratory. Shaw and Wallis (2010) demonstrated impaired mobility and movement coordination in 70 % of earwigs exposed to thiacloprid, and that more than 80 % of them died after 10 days exposure. Thiacloprid was also shown to reduce larval growth and to decrease adult foraging behavior (Fountain and Harris 2015). Acetamiprid significantly decreased the predation behavior of adult males by 28 % but not of females nor nymphs when applied in apple orchards at the agricultural rate (Malagnoux et al. 2015). Lacewings. Survival of the green lacewings Chrysoperla carnea reduced when adults feed on imidacloprid-treated

plants (Rogers et al. 2007). In addition, imidacloprid was found to disrupt the mobility of individuals (appearance

of tremors; Rogers et al. 2007). It has to be underlined that, upon multigeneration exposure, this species was able to develop strong resistance to acetamiprid (Mansoor and Shad 2020). Ladybugs. Ladybugs are impacted by neonicotinoids via prey ingestion, especially at early larval stage in Coleomegilla maculata feeding on cereal aphids exposed to thiamethoxam (Bredeson et al. 2015). Thiamethoxam reduces the mobility of ladybugs (the time to turn around when placed on their backs increases with the concentration of ingested insecticide) but not the number of eggs, while a negative correlation between the increase in the concentration of the insecticide and the number of developing eggs has been shown (Bredeson and Lundgren 2018). Wang et al. (2018a) evaluated the toxicity of thiamethoxam to Harmonia axyridis, a predator of the Myzus persicae aphid, and its effect in term of functional response, by three exposure routes: direct contact of H. axvridis with thiamethoxam residues; cabbage leaves infested with M. persicae treated systematically with thiamethoxam which exposed H. axyridis to the insecticide indirectly (referred as systemic application, mimicking direct soil drench or seed treatments); and cabbage leaves infested with M. persicae treated with thiamethoxam by leaf-dip which exposed H. axyridis to thiamethoxam residues on both cabbage leaves and thiamethoxam-treated M. persicae (referred as leaf dip treatment, mimicking foliar spray application). Predation was negatively affected under the three conditions, but particularly when ladybugs were exposed following leaf dipping. For all exposure routes, H. axyridis rapidly recovered predatory ability, however, sublethal effects of thiamethoxam may reduce the population growth of *H. axyridis* and, therefore, impair the biological control of *M. persicae*, especially after leaf or contact exposure. Parasitoid hymenoptera. Acetamiprid was demonstrated to cause significant reductions in the abundances of various groups of parasitoids (Aphelinidae, Braconidae, Encytidae, Eulophidae, Eupelmidae, Ichneumonidae, Mymaridae, Platygastridae, Proctotrupidae, Pteromalidae, Scelionidae, Trichogrammatidae) (Khans and Alhewairini 2019), and these losses were generally accompanied by an increase in pest infestation levels (Saito et al. 2008). In various parasitoid species, systemic applications of imidacloprid were often minimally detrimental, whereas foliar applications could be highly toxic (Naranjo 2001). Predatory mites. In the presence of neonicotinoids (acetamiprid, clothianidin, imidacloprid, thiacloprid or thiamethoxam), disruption of mite behavior (Panonychus ulmi, Tetranychus urticae), without loss of abundance, resulted in loss of biological control activity (Beers et al. 2005). Predatory mites (Phytoseiidae) are affected by acetamiprid, but studies have shown that they can develop resistance (Fountain and Medd 2015) which led to a growing interest in their use in sustainable agriculture (Duso et al. 2014; Fountain and Medd 2015).

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

Spiders. For several spider families (Araneidae, Lycosidae), contact exposure to neonicotinoids (acetamiprid, imidacloprid) appeared to be the most toxic pathway (compared to consumption of treated prey) inducing lethal and sublethal effects such as disruption of web construction (Pekar 2012). Furthermore, neonicotinoids (acetamiprid, thiacloprid) were demonstrated to affect the richness of spider communities (Rosas-Ramos et al. 2020).

Detritivorous arthropods

In a three-year field experiment, Pearsons and Tooker (2021) showed that seed treatments (corn, soybean) with neonicotinoids (clothianidin, imidacloprid) reduced saprophagous arthropod (millipede, springtails, oribatid mites) density and activity (litter decomposition) by more than 10%.

Earthworms

Earthworms are likely to be exposed to neonicotinoids in soils. For example, in a French arable landscape, Pelosi et al. (2021) observed residues of imidacloprid in 79% of the sampled earthworms (*Allolobophora chlorotica*, n=155; maximum concentration of 777 μ g/kg; 43 % of the earthworms contained imidacloprid concentrations >100 μ g/kg, LOQ = 0.4 μ g/kg), while thiacloprid was found in 34% of the earthworms (maximum concentration of 42.1 μ g/kg, LOQ = 0.1 μ g/kg).

Neonicotinoids (e.g., acetamiprid, clothianidin, imidacloprid, thiamethoxam) have negative effects on several endpoints of various earthworm species (e.g., *Eisenia fetida*, *Lumbricus terrestris*, *Aporrectodea caliginosa*), from sub-individual to community levels: tissue integrity, physiological activity, behavior, growth, reproduction, and survival (Dittbrenner et al. 2010; Dittbrenner et al. 2011a; Dittbrenner et al. 2011b; Qi et al. 2018; Tu et al. 2011; Wang et al. 2015). They are also known to be toxic to compost worms (*E. fetida*) in laboratory conditions: they affect reproduction, cellulase activity and tissues, among others (Wang et al. 2015).

Nematodes

Compared to arthropods, nematodes tend to be less sensitive to neonicotinoids (Kudelska et al. 2017; Neury-Ormanni et al. 2019; Bradford et al. 2020). In entomopathogenic species (*Steinernema glaseri*, *Steinernema carpocapsae*, *Steinernema feltiae*, *Heterorhabditis bacteriophora*, *Heterorhabditis megidis*), a positive effect of imidacloprid was observed at low dose on reproduction (Koppenhöfer et al. 2003).

Terrestrial vertebrates

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

Birds (excluding raptors)

Numerous studies demonstrated that bird decline in agroecosystems is related to the use of neonicotinoids (Ertl et al. 2018; Lennon et al. 2019; Li et al. 2020; Mineau and Palmer 2013; Mineau and Kern 2023).

In agricultural areas and other environments across Europe and North America, the analyses of neonicotinoid residues in various biological components (eggs, feathers, livers, plasmas) of several avian trophic groups such as nectarivores, granivores, insectivores and carnivores showed ubiquitous exposure of birds (gamebirds, house sparrows, hummingbirds, songbirds...) (Bishop et al. 2020; Bro et al. 2016; Fuentes et al. 2023; Humann-Guilleminot et al. 2019b; Humann-Guilleminot et al. 2021; Lennon et al. 2020a; Lennon et al. 2020b; Poisson et al. 2021; Prouteau 2021; Roy et al. 2020). The prevalence of exposure greatly varies from one study to another and among species, but, even if some studies detected neonicotinoids only in a few individuals (e.g., Graves et al. 2022), the vast majority of works underlined pervasive exposure of numerous species and pointed out high frequencies of detection.

Granivorous birds are directly exposed to neonicotinoids following the consumption of neonicotinoid treated seeds (Lopez-Antia et al. 2016; Prosser and Hart 2005; Roy et al. 2019). For example, Lennon et al. (2020b) demonstrated that the detection of clothianidin in the plasma of several farmland bird species increased from 11% before sowing to 51% after sowing. In French cereal dominated landscape, where neonicotinoid treated seeds were widely used, the eggs or livers of grey partridge (Perdix perdix) and of some Columba species were found to be contaminated by neonicotinoids (Bro et al. 2016; Millot et al. 2017). In Ontario fields (Canada), the analysis of carcasses of wild turkey (Meleagris gallopavo silvestris), which consumes neonicotinoid-coated seeds, showed detectable levels of clothianidin and/or thiamethoxam in 22.5% of individuals (detection of both substances in 5%) (MacDonald et al. 2018). These studies underlined that the crop sowing periods are the most at risk (especially in autumn compared to early spring, Millot et al. 2017) for bird exposure through neonicotinoid treated seeds, because it also corresponds to a period of low food availability and of migration stopover for some species. Along agricultural gradients in Minnesota (USA), at least one neonicotinoid among the seven compounds screened (acetamiprid, clothianidin, dinotefuran, imidacloprid, nitenpyram, thiacloprid, thiamethoxam) was detected in 93 % and 80 % of fecal pellets of sharp-tailed grouse (Tympanuchus phasianellus) and greater prairie-chickens (T. cupido), respectively, and in 90 % and 76 % of their livers, respectively (Roy and Chen 2023). Imidacloprid and clothianidin were the most detected substances. To document the exposure of wild bird communities, Anderson et al. (2023) analyzed seven neonicotinoids (acetamiprid, clothianidin, dinotefuran, imidacloprid, nitenpyram,

thiacloprid, thiamethoxam) in plasma samples from 55 species across 17 avian families, in four counties in Texas (USA). Imidacloprid was detected in 36 % of samples (n=294), and two birds contained imidacloprid, acetamiprid and thiacloprid. Clothianidin, and thiamethoxam were not detected but their LOD (0.3 µg/L, 0.05 µg/L, respectively) were higher than that of imidacloprid (0.005 µg/L). Temporal variations have been evidenced, with lower frequencies of detection in summer and winter than in spring and fall which correspond to the usual planting days for the most common crops across the state. Some species showed higher prevalence of exposure such as the American robin (Turdus migratorius) and the red-winged blackbird (Agelaius phoeniceus). Importantly, the study evidenced a chronic or repeated exposure of wildlife since six birds out of seven re-sampled over time exhibited at least one detection of neonicotinoid, and three exhibited multiple exposure at different time points (Anderson et al. 2023). In Europe, several measurements of neonicotinoid residues in bird carcasses (livers or gizzards) revealed very large numbers of accidental direct bird poisonings (passerine, Columba and game species) following the ingestion of neonicotinoid-treated seeds, especially with imidacloprid (Berny et al. 1999; Bro et al. 2010; Buchweitz et al. 2019; Millot et al. 2017; Mineau and Kern 2023; Mineau and Palmer 2013). Despite biases in the detection of carcasses in the field survey (de Snoo et al. 1999; Vyas 1999), a significant number of birds have been categorically identified as victims of acute and lethal poisoning induced by neonicotinoids used in seed treatments. Nevertheless, these lightning mortality events would likely not be the primary cause of the significant decline of some bird species (gray partridge) in agricultural environments, but they are undeniably an aggravating factor (Millot et al. 2017). This is all the more since many other direct sublethal (physiological and behavioral) and indirect effects of neonicotinoids have been demonstrated, for many more species than just granivores (Gibbons et al. 2015; Wood and Goulson 2017). Improved seeding techniques can limit the risk of direct poisoning by ensuring that treated seeds are effectively buried so that the proportion of seeds on the surface after planting is low (McGee et al. 2018). However, the effectiveness of these methods depends on planting techniques and on seed type and are not generalizable to all coated seed situations (McGee et al. 2018). Coatings have been suggested to induce an aversion which limits ingestion to a few coated seeds, representing only a small fraction of the neonicotinoid LD50 (Lethal Dose causing the death of 50% of exposed organisms) (Avery et al. 1994), but these results have been shown to depend on the experimental context, including the availability of alternative food resources or the state of food stress (Millot et al. 2017; Mineau and Kern 2023; Mineau and Palmer 2013). Furthermore, the repellent effect results from the induction of a physiological disorder following initial ingestions of treated seeds, involving that significant sublethal effects can occur well before ingestion of a lethal dose (Lopez-Antia et al. 2014; Lopez-Antia et al. 2015; Mineau 2017). It has to be underlined that some passerine species,

550

551

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574

575

576

577

578

579

especially *Fringillidae*, can de-husk seeds which lowers their direct exposure by ingestion (Prosser and Hart 2005). Other contexts of neonicotinoid poisoning of passerines (American goldfinches *Spinus tristis*) have also been identified in public spaces in California (Rogers et al. 2019): the mortality of birds was due to the ingestion of natural elm seeds remaining on the ground which were contaminated with imidacloprid during the drench application.

While neonicotinoids were initially thought to be less harmful to birds than insects due to their lower affinity for vertebrate nicotinic receptors, mounting evidence now challenges this view and birds appear to be more sensitive to neonicotinoids than other vertebrates (Mineau and Kern 2023; Mineau and Palmer 2013). The acute toxicity of neonicotinoids was reported to be underestimated by a factor of ten for some wild bird species compared to the one determined on model species of mallard or bobwhite quail (*Colinus virginianus*) (Mineau and Kern 2023; Mineau and Palmer 2013). Chronic toxicity is poorly taken into account, as well as sublethal effects which are scarcely investigated.

Several reviews of the individual and sub-individual effects of neonicotinoids on birds have been published (Gibbons et al. 2015; Moreau et al. 2022; Pisa et al. 2015; Wood and Goulson 2017). The literature shows that imidacloprid induces weight loss or reduces energy reserves (fat mass) in the white-crowned sparrow (Zonotrichia leucophrys) (Eng et al. 2017; 2019). In hummingbirds (Selasphorus rufus), the consumption of imidacloprid in flower nectar induces underactivity and decreased energy expenditure (-25%), with no other effect detected on feeding activity or immune response (Bishop et al. 2018; English et al. 2021). On the contrary, some studies showed an impact of imidacloprid on the immune status of adult (Lopez-Antia et al. 2013) and juvenile (Lopez-Antia et al. 2015) red-legged partridges (Alectoris rufa). These contrasting results could be explained by interspecific variability and various exposure conditions (dose x species x biomarkers x duration) (English et al. 2021; Gibbons et al. 2015; Lopez-Antia et al. 2015). Behavioral alterations were also observed (Eng et al. 2019), and disruption of flight and/or navigation efficiency emerged as a sensitive and relevant endpoint of imidacloprid exposure and sublethal effect on the white-crowned sparrow (Eng et al. 2017). These effects have been associated with loss of energy reserves. Thus, even if transient under the tested conditions, these sublethal effects can likely lead to impaired migration success of white-crowned sparrows using agricultural environments as staging areas (Eng et al. 2017; 2019). Furthermore, reductions in feeding and activity most often resulting in weight loss and risk to survival have been demonstrated in migratory birds exposed to sublethal doses of imidacloprid (Eng et al. 2017; 2019). Finally, exposure to sublethal dose of acetamiprid has been associated to reduced sperm density in the house sparrow (Passer domesticus) (Humann-Guilleminot et al. 2019c).

In controlled experiments on red-legged partridges (*Alectoris rufa*) fed with control seeds or seeds treated with imidacloprid at 20%, 100% or 200% of the recommended dose, analyses in livers showed an increase in the accumulation of imidacloprid with exposure time, and mortality of 50% of the females within five days even at agricultural or lower doses (Lopez-Antia et al. 2013; Lopez-Antia et al. 2015). Moreover, breeding investment was lowered with reduced clutch size, eggs size and fertilization rate, and chick survival was diminished when birds were exposed to imidacloprid.

Sabin and Mora (2022) performed an ecological risk assessment to evaluate the potential effects of neonicotinoids (acetamiprid, clothianidin, imidacloprid, thiamethoxam) on populations of the northern bobwhite (*C. virginianus*) in the South Texas Plains Ecoregion (USA). The assessment of the exposure of both juveniles and adults showed levels which can induce adverse effects on growth, reproduction success, and long-term survival.

The analysis of the literature thus demonstrated that neonicotinoids are one of the factors responsible of the decline in the abundance and diversity of birds. Depending on the bird species and their diet, this impact results mainly either from a direct effect (e.g., ingestion of treated seeds), or from an indirect effect (e.g., reduction in food resources following the decline of prey). Such indirect effects are addressed hereafter in the "Food webs" section.

Raptors

Several works showed the presence of neonicotinoids in raptors. Imidacloprid was detected in the blood of Eurasian eagle owl (*Bubo bubo*) in Spain (Taliansky-Chamudis et al. 2017), imidacloprid and thiacloprid in the blood of honey buzzards (*Pernis apivorus*) in Finland (Byholm et al. 2018), and acetamiprid, clothianidin, thiacloprid, and thiamethoxam in the feathers of barn owls (*Tyto alba*) in Switzerland (Humann-Guilleminot et al. 2021). The detection frequencies were contrasted: 3% of the analyzed samples were positive in the Eurasian eagle owl, whereas in the insectivorous honey buzzard, imidacloprid and thiacloprid were detected in 40 and 70% of the samples, respectively. In the barn owl, more than 80% of the individuals were positive, notably for thiacloprid, the frequent detection in chicks suggesting a trophic exposure. The feeding specialization of the barn owl on insects would not be sufficient to explain the high detection frequency of neonicotinoids. In northern Germany, Badry et al. (2021) investigated the impregnation of the livers of three raptor species (red kite *Milvus milvus*, common buzzard *Buteo buteo*, Montagu's harrier *Circus pygargus*; n=186). Among the neonicotinoids, only thiacloprid was detected in two red kites. Recently, no neonicotinoid was detected in the blood of chicks of the same three

raptor species in Germany (Badry et al. 2022). No study examining the toxicity of neonicotinoids on raptors has been identified.

641

642

643

644

645

646

647

648

649

650

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

639

640

Mammals (excluding chiropterans)

One of the largest mammalian studies conducted to date resulted in the simultaneous analysis of 480 substances in muscle of 42 wild boars (Sus scrofa), 79 roe deer (Capreolus capreolus) and 15 deer (Cervus elaphus) in Poland (Kaczynski et al. 2021). The five neonicotinoids were among the most frequently detected compounds (imidacloprid and thiacloprid showing mean concentrations in the top five values). They were detected in 100% of the wild boar samples, while acetamiprid was detected in three deer, and thiacloprid and clothianidin were detected in two deer. Acetamiprid, clothianidin and thiacloprid were detected in 13 roe deer (16.5%). The mean residue concentrations ranged from 0.6 µg/kg (thiamethoxam) to 4.3 µg/kg (imidacloprid) in the liver. In France, multi-residues analyses targeting 140 PPPs (67 withdrawn and 73 currently used PPPs) and transformation products were performed in hair samples of small omnivorous rodents (wood mouse Apodemus sylvaticus) and insectivorous shrews (greater white-toothed shrew Crocidura russula) sampled in arable landscapes (Fritsch et al. 2022). Again, acetamiprid, imidacloprid and thiacloprid were among the most frequently detected substances (more than 80% of individuals) and/or quantified at high concentrations (up to 70.7 μg/kg) (Fritsch et al. 2022). The ubiquity of exposure to neonicotinoids was demonstrated as residues were detected in all animals regardless of the type of habitat (hedgerows, cereal crops, grasslands) or of the agricultural practices (conventional or organic farming) (Fritsch et al. 2022). Assessing the exposure of wild raccoons (Procyon lotor) captured in Hokkaido (Japan) to neonicotinoids (acetamiprid, imidacloprid, clothianidin, dinotefuran, thiacloprid, thiamethoxam, and desmethyl-acetamiprid), Shinya et al. (2022) showed that either one of the six screened neonicotinoids or one transformation product was detected in the urine of 90% of the raccoons. Neonicotinoids were also found in the hair of red fox (Vulpes vulpes) in Italia; acetamiprid, clothianidin, and imidacloprid being detected in 100% of the analyzed individuals (n=11), and thiacloprid in 91% of them (Picone et al. 2023).

The toxicity of neonicotinoids to mammals have been reviewed by Tomizawa (2004) and Gibbons et al. (2015), showing the potential for various deleterious effects on growth, development and reproduction as well as other sub-lethal effects such as genotoxic and cytotoxic effects, immunotoxicity, neuro-behavioral disorders and changes in behaviors related to anxiety and fear, impairments of the thyroid and retina, and reduced movement. The study of the effects of imidacloprid (112 and 225 mg/kg, daily gavage for 60 days, which is above realistic environmental exposure concentrations) on rat reproduction, a mammal model organism, showed a decrease in

sperm vitality and number, a reduction in sex organ mass, and a decrease in the production of sex hormones FSH and LH in males (Nafaji et al. 2010; Tetsatsi et al. 2019). A significant impact of imidacloprid on the rat body weight was also reported but no published evidence of reproductive disorders in relation to neonicotinoid exposure in wild mammals was found. However, most of the research on mammals have been performed on rats or mice, and under laboratory conditions, hampering the assessment of direct toxicity to wild mammals which may exhibit different sensitivity and may be exposed to other chemical or biological stressors. As for birds, Gibbons et al. (2015) emphasized that neonicotinoids can also impact terrestrial mammals via indirect effects which are reviewed in the "Food webs" section.

Chiropterans

The exposure of wild bats to clothianidin, imidacloprid and thiamethoxam was demonstrated through the detection of the three substances in the hair of big brown bats (*Eptesicus fuscus*) sampled in Missouri (USA) (Hooper et al. 2022). Imidacloprid showed the highest frequency of detection and was found in all samples (Hooper et al. 2022). In Turkey, in a large screening targeting 322 PPPs and organic contaminants in adult bat carcasses of *Pipistrellus pipistrellus* and *Myotis myotis*, 87 compounds were detected but they didn't include neonicotinoids (Kuzukiran et al. 2021). Habitat preferences of these bats (urban and forest species) may limit their exposure to neonicotinoids. Several studies mention a risk of exposure of chiropterans to neonicotinoids by the trophic route, based on the monitoring of chiropteran activities and dosages in their prey present in the foraging sites (Stahlschmidt and Brühl 2012; Stahlschmidt et al. 2017).

In rare experimental studies, Hsiao et al. (2016) and Wu et al. (2020) reported the neurotoxic effects of imidacloprid (at 20 mg/kg/day) on the echolocation ability of insectivorous bats (*Hipposideros terasensis*). Memory loss in bats has been associated with apoptosis lesions in certain areas of the hippocampus (Hsiao et al. 2016). Another study supports these behavioral data and suggests that altered echolocation movements likely affects bat movement and hunting activities (Wu-Smart and Spivak 2018). In addition, neonicotinoid use appears to be associated with an increased frequency of white-nose syndrome, caused in chiropterans by a fungal infection, in both the USA and Europe (Bayat et al. 2014; Oliveira et al. 2021). Upon awakening, bats experience a massive inflammatory response phase with destruction of part of the immune tissue before reconstruction making them particularly vulnerable to infection (Mineau and Callaghan 2018). Neonicotinoids can thus come as an aggravating factor during this critical period. In their review, Mineau and Callaghan (2018) concluded that there is sufficient evidence to support the assert that bats are being negatively affected by neonicotinoids, directly through functional

impairment, and indirectly through reduction in insect abundance (trophic cascades are detailed in the "Food webs" section): the levels of neonicotinoid residues in the environment are high enough to put bats at risk of motor impairment and death. Knowledge remains currently too incomplete to be able to thoroughly characterize the impacts of neonicotinoids on chiropterans.

Reptiles

Neonicotinoids (imidacloprid and thiamethoxam) have been detected in several Mongolian racerunner (*Eremias argus*) organs and tissues (blood, brain, heart, lungs, stomach, intestine, liver, kidney, skin, fat, and gonads), showing different internal distributions and post-exposure temporal variations depending on the substance considered. However, the limited number of individuals which were analyzed prevents any attempt at generalization (Wang et al. 2018b; Wang et al. 2019).

The exposure of *E. argus* to thiamethoxam and imidacloprid under controlled conditions led to variations in thyroid, stress or sex hormone levels, endocrine gland damage, or changes in expression of genes involved in endocrine functions (Wang et al. 2019; Wang et al. 2020). Yang et al. (2020) also reported the endocrine disrupting effect of imidacloprid to *E. argus* with decreased levels of testosterone and estradiol in plasma. Further research is required to better characterize the impacts of neonicotinoids on reptiles.

Amphibians

Amphibians are one of the biological groups most affected by the collapse of biodiversity on a planetary scale, in particular because of the use of PPPs (Hayes et al. 2010). However, the number of studies of the effects of neonicotinoids on the terrestrial stages of amphibians is low. Comparing dermal exposure of *Hyla gratiosa* and *Hyla cinerea* to imidacloprid via direct exposure of the frog present on the soil at the time of insecticide spraying, and via indirect exposure following soil contact after application, van Meter et al. (2015) showed that cumulative concentrations and bioconcentration factors were significantly higher for the direct exposure. In the Pampa region of Argentina, imidacloprid was detected in the terrestrial *Leptodactylus latinasus* frog living in close association with row crops (soybean, corn, wheat) (Brodeur et al. 2022).

Thompson et al. (2022) used both aquatic mesocosms, and terrestrial locomotor and behavior trials to study the effects of sublethal exposure of the wood frog (*Rana sylvatica* or *Lithobates sylvaticus*) to imidacloprid. The results showed a decrease in larval survival to metamorphosis under imidacloprid exposure in interaction with shorter hydroperiod. However, the effect of imidacloprid depends on the frog stage: terrestrial locomotor

performances were improved following aquatic exposure of the larvae, while an important loss in these performances was observed after terrestrial exposure to imidacloprid. In addition, high effects on population sex structure and sexual development were observed: a skewed juvenile sex ratio was evidenced in imidacloprid treatments with about 10% fewer males than in controls, and 15.7% of individuals exposed to imidacloprid could not be assigned to either sex (ambiguous reproductive organ morphology) (Thompson et al. 2022). A great deal of research remains to be done.

Aquatic ecosystems

Contamination of freshwater and marine environments

Freshwater environment

Neonicotinoids used in agricultural fields can enter surface waters (from rivers to lakes) through spray drift, dust from coated seeds, runoff, subsurface flow (for example, subsurface tile drainage), input of treated leaves, and/or plant decomposition in water (Alford and Krupke 2019; Stehle et al. 2018; Wang et al. 2023). The primary routes of transfer are direct contamination due to spray drift or to dust abrasion of coated seeds, and re-distribution from surface runoff or subsurface drainage (Schaafsma et al. 2019; Wettstein et al. 2016). Neonicotinoids are stable in water, and because of their high mobility, they are mainly transported in the dissolved phase (Bonmatin et al. 2015; Morrissey et al. 2015; PPDB 2023).

After neonicotinoid applications, the delivery ratio to surface water was estimated to be less than 2% for thiamethoxam and clothianidin together, and 0.48% for imidacloprid (Frame et al. 2021; Wettstein et al. 2016). The detection rates in surface water are higher after seed treatment than after spraying (Wettstein et al. 2016). In North America, clothianidin was found before, during and after planting (i.e., in 98% of the samples), while the detection of thiamethoxam mainly occurred in the post-plant season (54% of the samples), and that of imidacloprid during the planting season (48% of the samples) (Evelsizer and Skopec 2018). Clothianidin is both a PPP and a transformation product of thiamethoxam which could explain its higher frequency of detection (Wang et al. 2023).

Neonicotinoids have been quantified in various types of surface waters including wetlands, ditches, ponds and rivers (Table 1). Acetamiprid, imidacloprid, and thiamethoxam are the most frequently detected substances (Pietrzak et al. 2019). Overall, maximum concentrations of neonicotinoids in surface waters were found to be 9.14 μ g/L for imidacloprid, 6.90 μ g/L for thiamethoxam, 4.00 μ g/L for acetamiprid, 3.50 μ g/L for clothianidin, and 1.37 μ g/L for thiacloprid (Alford and Krupke 2019; de Araújo et al. 2022; Criquet et al. 2017; Evelsizer and Skopec

2018; Kuechle et al. 2019; Nélieu et al. 2021; Pietrzak et al. 2019; Schaafma et al. 2019; Wang et al. 2023) (Table 1). Most of these reported maximum concentrations exceed the ecological thresholds for neonicotinoid water concentrations (0.2 μ g/L for short-term acute exposure and 0.035 μ g/L for long-term chronic exposure) which were defined to avoid lasting effects on aquatic invertebrate communities (Morrissey et al. 2015). A recent review provided a meta-analysis of neonicotinoid concentrations in water, based on more than 40 papers published in ten countries (Wang et al. 2023). It reported mean concentrations of 0.222 μ g/L (n=1056) for clothianidin, 0.120 μ g/L (n=879) for imidacloprid, 0.059 μ g/L (n=863) for thiamethoxam, 0.023 μ g/L (n=428) for acetamiprid, and 0.011 μ g/L (n=295) for thiacloprid.

Some mitigation measures could consist in improving the application material to prevent dust during planting of treated seeds, and to improve water interception of surface and subsurface flow thanks to buffer zones such as wetlands. For example, in constructed wetlands, removal of neonicotinoids due to direct accumulation in macrophytes and to enhanced biodegradation was estimated to range from 10 to 100% in 28 days (Liu et al. 2021; Main et al. 2017).

Marine environment

Neonicotinoids have only been recently monitored in coastal and marine environments. Consequently, data are just available for imidacloprid and thiamethoxam, which are generally searched for using passive integrative POCIS samplers or directly in water. In mainland France, these substances were not found in the Channel/North Sea coast (Menet-Nedelec et al. 2018). On the contrary, on the other two maritime facades (Bay of Biscay and Mediterranean), imidacloprid and thiamethoxam were quantified quite frequently (with maximum frequencies of detection of 20%) in the coastal waters of the Arcachon Basin (maximum of 0.14 µg/L and 0.0039 µg/L for imidacloprid and thiamethoxam, respectively, in spot samples) (Auby et al. 2011; Tapie and Budzinski 2018) (Table 1), in transitional waters of the Gironde estuary (maximum imidacloprid concentration of 0.0053 µg/L with integrative sampling) (Levesque et al. 2018), in Marennes-Oléron bay (maximum of 0.0238 µg/L and 0.0004 µg/L for imidacloprid and thiamethoxam, respectively, with integrative sampling) (Pepin et al. 2017), and in Mediterranean lagoons (maximum of 0.028 µg/L and 0.0025 µg/L, for imidacloprid and thiamethoxam, respectively with integrative sampling) (Munaron et al. 2020; Munaron et al. 2023). Imidacloprid has also been detected in the Charente estuary and in the Loire estuary since 2006 (GIP Loire Bretagne 2013). According to ecotoxicological data collected in the OBSLAG (Observatory of the Mediterranean Lagoons) study, only imidacloprid would cause a chronic risk for the biota of lagoon ecosystems (exceeding its chronic marine predicted

no effect concentration PNEC in several lagoons since the beginning of the monitoring in 2017) (Munaron et al. 2022). This risk can be extended to the Arcachon basin and Marennes-Oléron bay given the reported data. No neonicotinoid was found in French marine sediments and no reference from the French overseas territories mentions their research in the water of the marine environment.

Only scarce information is available evidencing the contamination of marine waters worldwide. In the Queensland region of Australia, streams flowing into the marine waters of the Great Barrier Reef were found to be contaminated with imidacloprid at levels ranging from 0.0005 to 1.3 μ g/L (Warne et al. 2022). The contamination concerned observation sites located in downstream sectors near the mouths of large rivers (Warne et al. 2022). This pattern appeared similar in the Bohai Sea (China), where Naumann et al. (2022) observed the seasonal variation in neonicotinoid concentrations in rivers and marine water. In their study, the detection frequency of acetamiprid was 100% in both river (n=72) and marine (n=81) waters in summer and fall. Despite dilution in the coastal environment, the risk quotient associated with the contamination levels were reported as high risk for marine organisms regarding imidacloprid, thiamethoxam and acetamiprid (Naumann et al. 2022). Due to their slow degradation rates in the environment and binding properties to particulate organic matter (PPDB 2023), neonicotinoids are likely to accumulate in sediments: Chen et al. (2022) reported contamination of marine sediments in East China Sea, due to the Yangtze River inputs, several tenths of kilometer from the river mouth. The mean concentration of total neonicotinoids was 11.9 μ g/kg (dry weight). The authors concluded that marine sediments were a major sink for neonicotinoids, highly used in continental China as PPPs (Chen et al. 2022).

Impacts on aquatic biodiversity

Aquatic microorganisms

Few studies have been published on the effects of neonicotinoids on aquatic microorganisms. They suggest that imidacloprid does not affect the activity and respiration of aquatic microbial decomposers (Kreutzweiser et al. 2007; Kreutzweiser et al. 2008). With the exception of the study of Neury-Ormanni et al. (2020a), who observed that an exposure of the freshwater diatoms *Planothidium lanceolatum* and *Gomphonema gracile* to 5 µg/L imidacloprid resulted in indirect effects via competition and predation, effects of neonicotinoids on different microalgae (e.g., *Desmodesmus subspicatus*; Malev et al., 2012) and cyanobacteria (e.g., *Synechocystis* sp.; Li et al., 2010) were only observed at very high concentrations (i.e., several mg/L), irrelevant to environmental contamination levels. Using a quantitative structure activity-toxicity modeling approach, Gökçe and Saçan (2019) also predicted an absence of effects of acetamiprid on microalgae exposed to up to 100 mg/L. Neonicotinoids are

therefore unlikely to be toxic to aquatic microbes, including primary producers, except under extreme events of contamination.

820

821

822

823

824

825

826

827

828

829

830

831

832

833

834

835

836

837

838

839

840

841

842

843

844

845

846

818

819

Aquatic invertebrates

Works focused on the effects of neonicotinoids on aquatic invertebrates are increasingly investigated (as compared to other insecticide classes, such as carbamates and organophosphates) due to the relative recentness of their use (first homologations date back from the 1990s), and to the risk specifically posed to aquatic invertebrates because of the levels of water contamination reported (see above). Morrissey et al. (2015) highlighted strong evidence that water-borne neonicotinoid exposure is frequent, long-term and at concentrations which commonly exceed several existing water quality guidelines. In addition, several monitoring studies of watercourses in either agricultural or urban landscapes demonstrated a significant contamination of freshwater amphipods (*Gammarus pulex*) by neonicotinoids (e.g., Shahid et al. 2018a; Švara et al. 2021).

Despite awareness of these contamination levels, works devoted to the effects of neonicotinoids on aquatic invertebrate biodiversity are still limited. A first review published in 2015 noted the weak level of knowledge available on the effect of neonicotinoids on the invertebrate fauna of freshwater and marine environments (Pisa et al. 2015). Since then, various field case studies have provided data and documented/predicted effects of neonicotinoids on aquatic invertebrate communities. For example, in Canadian wetlands near treated rapeseed crops, a correlation was established between neonicotinoids (acetamiprid, clothianidin, imidacloprid, thiamethoxam) transfer during rainfall events and changes in emergent insect (Diptera) diversity (Cavallaro et al. 2019). Through an experimental rice mesocosm study, imidacloprid was found to significantly reduce populations of various insects (dragonfly, bug, beetle) (Kobashi et al. 2017). A drastic decline in zooplankton biomass in Japanese brackish lakes also coincided with the introduction of neonicotinoids (clothianidin, imidacloprid, thiamethoxam) in rice agriculture since the 1990s, followed by collapse of predator fish populations (Yamamuro et al. 2019). In the Netherlands, where imidacloprid residues in water are particularly high, correlations between these residues and decline in arthropod taxa such as mayflies, odonates, diptera, and some crustaceans were revealed on a national scale (van Dijk et al. 2013). This was also observed in a study adopting a PAF (Potentially Affected Fraction) approach, but with much lower proportions of species potentially affected by neonicotinoids taking into account the co-occurrence of other PPPs in the studied environments (Vijver and van den Brink 2014).

Comparing recorded or predicted concentrations of neonicotinoids in the aquatic environment to ecotoxicity thresholds has raised some concerns for the potential effects of these insecticides in freshwater environments. The review by Sánchez-Bayo et al. (2016) reported widespread effects of neonicotinoids on aquatic species in the USA, and the major risk for aquatic invertebrates was reaffirmed in 2017 (Wood and Goulson 2017). More recently, a study based on an agricultural region located in an ecologically important wetland (Nebraska's Rainwater Basin, USA), showed negative correlations between neonicotinoid concentrations and macroinvertebrate biomass (which represents potential resources for various migratory birds) despite concentrations below the acute toxicity risk thresholds proposed by the USEPA (Schepker et al. 2020).

Long-term ecological impact of neonicotinoids is a particularly salient issue for aquatic invertebrates. The chronic risk mainly results from the ability of neonicotinoids to reach aquatic environments (high solubility in water) and to persist there when they are adsorbed on particles (Armbrust and Peeler 2002). However, this risk is poorly assessed because most often based on toxicity tests on Daphnia, an organism more tolerant than insects and other arthropods to neonicotinoids (Beketov and Liess 2008; Wood and Goulson 2017). Neonicotinoids can have chronic effects on abundance and community structure of freshwater arthropods and other macroinvertebrates at doses in the µg/L range and below (Beketov and Liess 2008; Kattwinkel et al. 2016). After cessation of treatments, the onset of delayed effects was also demonstrated in situ (limnocorrals) for much lower concentrations of imidacloprid and clothianidin (< 0.05 μg/L) resulting in a significant advancement of the emergence date of chironomids and zygopteran odonates (Cavallaro et al. 2018; Williams and Sweetman 2019). From a functional point of view, the desynchronization of phenology of these organisms could have important consequences on ecosystems, especially in terms of biomass input to the terrestrial environment (trophic resource for terrestrial predators such as birds). Lethal and sublethal effects of thiacloprid have been demonstrated in various aquatic invertebrates, several days after exposure, for moderate acute toxicity concentrations (Beketov and Liess 2008). Neury-Ormanni et al. (2020b) documented altered feeding behavior in chironomids exposed to environmental doses of imidacloprid. The insecticide induced changes in motility, feeding selectivity, and browsing ability. The reduced abundance and altered emergent aquatic insect assemblages in wetlands exposed to neonicotinoids could explain the reduction in densities of insectivorous birds in such environments (Cavallaro et al. 2019).

Investigating the idea of long-term impact of neonicotinoids beyond the lifespan of exposed individuals, recent works with the model amphipod crustacean, *G. pulex*, suggested the development of tolerance towards clothianidin within populations from watercourses in agricultural landscapes (Becker and Liess 2017; Becker et al. 2020; Shahid et al. 2018b). According to the authors, in these populations, the evolution of resistance by natural

selection could be facilitated by factors acting at the population and/or community levels: distance from non-tolerant populations, which would favor selection locally by limiting gene flow and the influx of non-adapted genes into populations (Hoffmann and Willi 2008), and low community diversity which would intensify intraspecific competition in gammarids. Nevertheless, the shift in sensitivity of this non-target species to the neonicotinoid appeared very moderate (less than three-fold change in LC50 for example) in comparison to the genetic resistance reported for other neurotoxic insecticides (pyrethroids and organophosphates) in the amphipod *Hyalella azteca* (Gamble et al. 2023; Weston et al. 2013). In addition, an inverse pattern with increased sensitivities of long-term exposed *G. pulex* populations towards imidacloprid was found in non-agricultural context presenting complex mixture of organic contaminants (Švara et al. 2021). Overall, these results demonstrate the unsuspected importance of evolutionary adaptative processes underway in natural populations unintentionally exposed to neonicotinoids, and the urgency to develop assessment tools specifically focused on long-term effects (Oziolor et al. 2016). Such processes should be anticipated, at least in insects and probably in other arthropods, from the current knowledge on the selective evolution of resistance to neonicotinoids in pests, based either on target-site mutation or on metabolic resistance (Bass et al. 2015).

Although environmentally less realistic than field approaches, experimental studies performed in mesocosms and in the laboratory (e.g., common garden), offer the statistical power required to test patterns observed in natura (Barmentlo et al. 2021), as well as interactions with other environmental factors susceptible to alleviate or aggravate the effects of neonicotinoids, such as PPP mixtures (Sanchez-Bayo and Goka 2012; Rico et al. 2018; Sol Dourdin et al. 2023), temperature/climate (Mohr et al. 2012; Sumon et al. 2018; Rico et al. 2018), nutrients/fertilizers (Barmentlo et al. 2019; Chara-Serna et al. 2019), vegetation disturbance (Cavallaro et al. 2019), and indirect effects between species representative of different functional groups in the community (e.g., such as predator-prey relationships; Miles et al. 2017). In this regard, Alexander et al. (2013) used artificial streams to examine the impact of mixing three insecticides expected to act additively, i.e., imidacloprid (which acts on the acetylcholine receptor) and two organophosphates which act on the acetylcholine esterase enzyme, chlorpyrifos and dimethoate, and under oligotrophic vs mesotrophic (nitrate input), along a Toxic Unit (TU) gradient established for concentrations consistent with environmental data. The study showed a significant interaction between insecticides and nutrients on macroinvertebrate communities, with notably, under mesotrophic condition and low insecticides pressure, an increase in the total abundance and species richness of ephemeropteran, plecopteran and trichopteran insects. At higher insecticides pressure, the overall density of these groups and the entire community was the most reduced in mesotrophic streams. In contrast, for other species groups such as chironomids, detritus feeders, and the odonate predator Gomphus spp., no significant interaction between insecticides and nitrate was detected. In oligotrophic environments, increasing PPP doses decreased predation intensity, which in turn affected abundance patterns while, in mesotrophic environments, a bottom-up effect of nutrients on the periphyton explained the variation in macroinvertebrates abundance and richness. Such causeand-effect relationships were also analyzed with Structural Equation Modeling (SEM) approaches which describe effect pathways among different variables of interest (Miller et al. 2020; Schmidt et al. 2022). At low doses, the toxicity of PPPs appeared hidden by nutrients because of increased compensatory consumption, expression of adaptive plasticity at the intraspecific level, or differential responsiveness across taxa, processes which are not captured by traditional community study methods (taxonomic determination and records of relative abundances). Interactions between nutrients and PPP can thus result in a redirection of energy within food webs towards nonproductive pathways (Davis et al. 2010) or in a shift in communities towards more tolerant groups (Vinebrooke et al. 2004). This type of interactions was also studied in terms of convergence/divergence of invertebrate community structure in open artificial ditches (naturally assembled communities), by combining NPK elements with thiacloprid (Barmentlo et al. 2019). Following thiacloprid treatments designed to maintain concentrations for one month (two spikes separated by two weeks), no effect of treatments, other than an increase in total abundance after four months due to nutrient input, was found in terms of taxon richness, overall abundance, or within-treatment community divergence/convergence through time (\beta dispersion). However, significant changes were observed in community composition under the effect of thiacloprid, nutrients and combination thereof. This effect persisted several months after the disappearance of thiacloprid from the medium. The main compositional changes were a reduction in the abundance of insects and large predators, and an increase in multivoltine species. Some results, such as the particularly strong increase in Helophorus beetles under nutrients and thiacloprid, may reflect a PPPinduced rippling effect on the community amplified by nutrient supply. This study shows that thiacloprid, in addition to its short-term toxicity, induces indirect longer-term ecological effects.

907

908

909

910

911

912

913

914

915

916

917

918

919

920

921

922

923

924

925

926

927

928

929

930

931

932

933

934

935

936

Overall, the corpus analyzed pointed to a marked impact of neonicotinoids on aquatic arthropods at low doses, as demonstrated once again in a recent study which reports the decline in emerging aquatic insects during a three-month semi-field experiment considering environmentally realistic contamination scenarios of thiacloprid (Barmentlo et al. 2021). However, more studies remain to be performed to determine the relationship between the impacts of neonicotinoids and fitness of organisms, in relation to the ecological functions to which they contribute, as well as on the relationship between the impacts of neonicotinoids on the nervous system and the behavior of aquatic invertebrates.

Aquatic vertebrates

Amphibian larvae and tadpoles

The sensitivity of amphibian species to neonicotinoids through water contamination has been rarely studied. Green frog (*Rana clamitans*) tadpoles were found to be relatively insensitive to imidacloprid with mortality observed after 96h of exposure to high concentrations only (150 mg/L) (Puglis and Boone 2011). This lack of sensitivity is likely due to differences in the vertebrate nicotinic acetylcholine receptor relative to their invertebrate homologs (Li et al. 2016). On the contrary, spotted marsh frog tadpoles (*Limnodynastes tasmaniensis*) suffered high mortality rates (up to 17%) when they were exposed to imidacloprid concentrations as low as 0.50 μg/L (Sievers et al. 2018). This exposure level reduced swimming speed and distance, and escape responses which then made the tadpoles more susceptible to predation, while increasing erratic swimming (Sievers et al. 2018). The toxicity of imidacloprid has also been demonstrated in the tadpoles of *Leptodactylus luctator* and *Physalaemus cuvieri* (Samojeden et al. 2022). The consequences of exposure to environmental concentrations (3-300 μg/L) led to a decrease in size, to morphological malformations (for the two species), and to changes in tadpole swimming activity (only for *L. luctator*).

In the current literature, there is limited evidence of the effects of neonicotinoids on amphibians under chronic exposure to aquatic environmental concentrations. However, neurotoxic responses can be observed. Campbell et al. (2022; 2023) demonstrated the ability of imidacloprid to cross the blood-brain barrier and to concentrate over 300-fold in the brain of juvenile northern leopard frogs (Rana pipiens) with some consequences on foraging behavior (e.g., a decrease in reaction times to a food stimulus by 1.5 to 3.2 times for organisms exposed to concentrations up to 10 µg/L). At concentrations ranging from 0.1 to 10 µg/L and over a 21 day exposure period, bioaccumulation of imidacloprid in frog brains is accompanied by a decreased reactivity in individuals subjected to feeding stimuli. Beyond the active substance, the transformation product imidacloprid-olefin was detected in the brains of amphibians at much lower concentrations, which does not mean that this compound cannot be responsible for any toxic action. Surprisingly, exposure of leopard frogs to imidacloprid led to increased growth primarily affecting body length (Campbell et al. 2022). Recent research has further demonstrated that wood frogs (R. sylvatica or L. sylvaticus) exposed to imidacloprid (10 or 100 µg/L) at the tadpole stage were less likely to escape simulated predator attacks in the laboratory, suggesting that exposure to this insecticide may negatively impact tadpole perception and cognitive function (Lee-Jenkins and Robinson 2018; Sweeney et al. 2021). However, at a lower concentration of 0.1 µg/L, imidacloprid did not induce any modulation of acetylcholinesterase activity in bullfrog (Lithobates catesbeiana) tadpoles after three weeks of exposure (Rios et al. 2017). For other less studied neonicotinoids as chlothianidin, frog tadpoles are among the least sensitive species in case of laboratory exposure at sublethal concentrations (Miles et al. 2017). The tadpoles are tolerant to clothianidin, confirming the low toxicity of neonicotinoids in vertebrates (Miles et al. 2017). As stated in the section focused on the impacts of neonicotinoids on amphibians during their terrestrial life, numerous research remain to be done to characterize their impacts on amphibians in aquatic media.

Fish

In general, neonicotinoids exhibit low acute toxicity to fish. The 96h LC50 of clothianidin ranges from 93.6 mg/L for sheepshead minnow (*Cyprinodon variegatus*) to 117 mg/L for bluegill sunfish (*Lepomis macrochirus*) (Anderson et al. 2015). A similar trend is observed for imidacloprid, with 96h LC50 ranging from 211 mg/L for rainbow trout (*Oncorhynchus mykiss*) to 280 mg/L for common carp (*Cyprinus carpio*) (Anderson et al. 2015). Two formulations of thiamethoxam have 96h LC50 above 100 mg/L (Anderson et al. 2015). These results indicate that fish are insensitive to neonicotinoids, probably because of the properties of the vertebrate nicotinic acetylcholine receptor (Li et al. 2016).

Nevertheless, the available data indicate that exposure of aquatic vertebrates to sublethal concentrations of neonicotinoids results in pro-oxidative responses from which genotoxic perturbations arise. A short 48h exposure of the freshwater cichlid fish (*Australoheros facetus*) to imidacloprid concentrations of 100 and 1000 µg/L affected the integrity of fish erythrocyte DNA (COMET assay and micro-nuclei test) (Iturburu et al. 2018). Under short-term exposure to a much lower concentration of thiamethoxam (3.75 µg/L), the siluriform catfish (*Rhamdia quelen*) showed activity inhibition of two liver enzymes, adenylate kinase and pyruvate kinase, as early as 24h of exposure (Baldissera et al. 2018). These inhibitions were associated with a decrease in ATP levels in the liver. The energetic deregulation appeared to persist after the fish were no longer contaminated (Baldissera et al. 2018). Beyond these non-specific effects, neonicotinoids can act on the nervous function of non-target organisms, given their mode of action (binding to nicotinic acetylcholine receptors at neuromuscular junctions leading to insect paralysis) (Kimura-Kuroda et al. 2012). Imidacloprid was found to be neurotoxic to adult rainbow trout (*O. mykiss*) exposed for 21 days to high concentrations (10 and 20 mg/L) (Topal et al. 2017). This neurotoxicity resulted in inhibition of acetylcholinesterase activity, oxidative stress, and a concomitant increase in DNA damage in the fish brains (Topal et al. 2017).

Neurotoxicity of neonicotinoids may also impact the behavior of fish. A laboratory test developed to investigate two key responses of fish anti-predator behaviors revealed that zebrafish (*Danio rerio*) larvae exposed

for 24 hours to acetamiprid exhibited increased fear reflex and faster habituation compared to unexposed larvae (Faria et al. 2020). The concentrations tested in this study were considered to be realistic (0.04 and 0.40 µg/L) in relation to measured concentrations of acetamiprid in surface water (0.008 to 44 µg/L) (Faria et al. 2020). The modulations of fish larvae anti-predator behavior observed in the laboratory raise questions about the environmental reality of such effects and about their hypothetical consequences in terms of survival capacity in the environment. Könemann et al. (2021) observed that zebrafish larvae were able to avoid imidacloprid contamination, but did not react to other neonicotinoids such as thiacloprid. In addition, the experimental ablation of olfaction abolished aversive responses of individuals, indicating that fish may sense insecticides. In this species, the assessment of neural activity in 289 different brain regions revealed a particular modulation of hypothalamic areas involved in the fish stress response, indicating that the observed behavioral patterns are close to those observed for other stress responses (Könemann et al. 2021). Juvenile medaka (Oryzias latipes), exposed to imidacloprid under rice cultivation field conditions, were consecutively infected by a Trichodina parasite (Sánchez-Bayo and Goka, 2005). Such pathology was linked to the chemical stress induced by imidacloprid. If toxicity of imidacloprid to vertebrates was extensively studied, the toxicity related to imidacloprid transformation products (5-hydroxy-imidacloprid, imidacloprid-urea and 6-chloronicotinic acid) was not taken into account until now, despite their presence in various tissues as observed, for example, in muscle, gonads, brain and gills in Goldfish (Carassius auratus) (Xu et al. 2023).

A few studies deal with the combined effects of neonicotinoids with other PPPs but sometimes with experimental approaches that are more or less relevant in the context of ecological risk assessment. Thus, adult zebrafish exposed by immersion during 24 hours to high concentrations of imidacloprid (13.75 mg/L) associated with the organophosphate insecticide dichlorvos (7.5 mg/L) and the herbicide atrazine (1.5 mg/L) showed high levels of lipid peroxidation, particularly in the liver, compared to fish exposed to the same active substances tested in isolation (Shukla et al. 2017). Although this type of study is useful to test the hypothesis of expected synergistic effects, it does not allow estimation of the actual environmental risk, particularly in view of the contamination of surface waters reported by the authors (in the Ebro River in Spain: minimum concentration of imidacloprid of $0.0016 \,\mu\text{g/L}$ and maximum concentration of $0.015 \,\mu\text{g/L}$) (Shukla et al. 2017). It is therefore important to consider such data with caution when assessing the ecotoxicity of neonicotinoids. Similarly, mixture of the order of mg/L imidacloprid and organophosphate insecticide triazophos used to assess embryotoxicity to zebrafish early larvae (blastula stage: 2h post-fertilization) exposed during 96h revealed a strong synergistic effect in terms of acute toxicity (Wu et al. 2018). Although relevant in terms of mixture toxicity assessment, such high concentrations still

lack environmental relevance. It is worth noting that, though concentrations were still high, synergistic effects were also demonstrated on zebrafish larvae (72h post-hatching) for various combinations of imidacloprid with atrazine, butachlor, chlorpyrifos or lambda-cyhalothrin (mixtures containing from two to five substances) (Wang et al. 2017).

No study has been devoted to the effects of neonicotinoid mixtures on aquatic vertebrates (Anderson et al., 2015). In addition, there is a lack of ecosystem-scale studies (mesocosm approaches and/or field studies) to investigate the effects of these insecticides. Work is also needed on sub-lethal or chronic effects to reflect environmental concentration levels. Finally, most of the studies focus on imidacloprid, with very little attention paid to the effects of other neonicotinoids.

Aquatic birds

Aquatic birds include waterbirds, which live in freshwater environments, and seabirds, which feed on the resources of seas and oceans.

The exposure of seabirds to neonicotinoids (acetamiprid, clothianidin, imidacloprid, thiacloprid, thiamethoxam) was characterized by analyzing residues in feathers sampled from the piscivorous Sandwich tern (*Thalasseus sandvicensis*) and the mixotrophic Mediterranean gull (*Ichthyaetus melanocephalus*) in fledglings from the Lagoon of Venice (Distefano et al. 2022). Neonicotinoids were detected in both species, and imidacloprid and clothianidin were the most often quantified ones (100% in Mediterranean gulls and 58% in Sandwich terns, and 100% in Mediterranean gulls and 61% in Sandwich terns, respectively). The detection of thiacloprid was lower (<20% of samples in both species) (Distefano et al. 2022). On the contrary, no residue of neonicotinoids was found in the liver or blood of white-tailed sea eagles (*Haliaeetus albicilla*) and ospreys (*Pandion haliaetus*) (Badry et al. 2021; Badry et al. 2022).

For waterbirds, data are even more scarce. In some rice-growing regions, aquaponic practices involve ducks for the control of weed and pest in rice fields (Mburia, 2016). In this very particular context, ducks may be contaminated with neonicotinoid residues (Khidkhan et al., 2022).

To date, no result on the direct effects of neonicotinoids on seabirds and waterbirds were available in the literature. Thus, even if the toxicity of neonicotinoids to aquatic vertebrates is presumed to be limited, there are still many areas of knowledge that need to be clarified and completed such as toxicity of transformation products, and levels of impregnation of agricultural wetland-living organisms by native substances and their transformation products (Frank and Tooker, 2020).

Food webs

Neonicotinoids can affect terrestrial and aquatic biodiversity by spreading through food webs, by the propagation of adverse biological effects in food webs and disturbance of trophic interactions (e.g., reduced predation rate, increased mortality of predators), and/or by reducing food resources (Alsafran et al. 2022). However, the number of results which have been published in the literature remains limited.

Terrestrial ecosystems

Focusing on insects, Tooker and Pearsons (2021) reviewed the mechanisms underlying the effects of insecticides on food webs. They highlighted how neonicotinoids influence trophic interactions and food webs, and contribute to insect declines. Neonicotinoids spread across trophic levels, primary and secondary consumers being exposed through several routes (including dietary and trophic routes), and they may also bioaccumulate in some organisms (Tooker and Pearsons 2021). Neonicotinoids distort food webs by significantly decreasing insect abundance and diversity of both preys and consumers, as evidenced in various ecosystems (e.g., croplands, woodlands, watercourses). Depopulated and less diversified insect communities lead to food scarcity for their predators, thereby adversely impacting their local population dynamics. Importantly, food web disruption can occur even when neonicotinoids do not bioaccumulate or biomagnify in food webs, depending on the sensitivity of the taxa constituting the lower trophic levels (i.e., toxic effects on prey inducing adverse effects on higher levels via trophic cascades) and/or the sensitivity of higher trophic levels (i.e., relatively low concentrations but high enough to induce toxic effects on sensitive predators) (Tooker and Pearsons 2021).

In terrestrial invertebrates, thiamethoxam has been reported to have no effect on the predation rates of two predators, *Orius insidious* insidious flower bug and *Hippodamia convergens* ladybug, after consuming aphids reared on thiamethoxam-treated plants (Esquivel et al. 2020). On the contrary, insidious flower bug survival, unlike that of ladybugs, was reduced following aphid consumption. However, the reduction in bug survival was only observed in the first few weeks after thiamethoxam application, and no reduction was noted one month after treatment or beyond. In an urban context (Central Park, New York City, USA) where trees were treated with imidacloprid against an alien beetle (*Anoplophora glabripennis*), unexpected outbreaks of a formerly innocuous herbivore, *Tetranychus schoenei* (Tetranychidae), followed insecticide applications to elms (Szczepaniec et al. 2011). Changes in the structure of arthropod communities sampled in elm canopies after imidacloprid treatments were evidenced, mainly related to an increase in the abundance of *T. schoenei*. Laboratory tests showed that

exposure to imidacloprid through consumption of imidacloprid-treated elm foliage enhanced the fecundity of T. schoenei by 40%: adult T. schoenei fed leaves from treated elms laid more eggs than when fed with leaves from untreated elms (Szczepaniec et al. 2011). However, no effect of imidacloprid on T. schoenei fecundity was detected when mites were directly sprayed with the insecticide. The longevity of mites was also not affected by exposure to imidacloprid via food. Two model predators of spider mites, the Coccinellidae Stethorus punctillum (adult) and the Chrysopidae Chrysoperla rufilabris (larva), showed significant decrease in feeding rates when offered mites from imidacloprid-treated elms as preys. Moreover, the predators exhibited signs of intoxication (partial or complete lack of response to touch, tremors, regurgitation, excessive grooming, and inability to right themselves when placed on their back) and deleterious effects when exposed to imidacloprid by consuming prey from leaves of treated trees such as impaired mobility and reduced longevity (about one-two days when mites fed from treated trees versus 9-13 days when T. schoenei fed from untreated trees) (Szczepaniec et al. 2011). By stimulating reproduction of mites while poisoning insect predators of spider mites which may reduce top-down regulation, imidacloprid tree treatments finally led a non-target innocuous herbivore to reach a pest status (Szczepaniec et al. 2011). This study underlined how neonicotinoids may disrupt ecosystem functioning and impair ecological balance that ultimately can favor pest outbreaks. Studying the effect of thiamethoxam on the spider mite (Tetranychus urticae, considered as a pest in various agricultural systems) and its predator Phytoseiulus persimilis, Pozzebon et al. (2011) showed that the neonicotinoid was toxic to both T. urticae and P. persimilis, but that the impact of thiamethoxam varied according to the routes of exposure. The authors demonstrated that topical exposure led to sublethal effects in predators and prevs while residual and contaminated food exposures led to both lethal and sublethal effects. In addition, toxicity increased when several exposure routes were involved. By limiting exposure to thiamethoxam to ingestion of contaminated food only, the impact of the insecticide was more favorable to P. persimilis than to its prey (Pozzebon et al. 2011).

1086

1087

1088

1089

1090

1091

1092

1093

1094

1095

1096

1097

1098

1099

1100

1101

1102

1103

1104

1105

1106

1107

1108

1109

1110

1111

1112

1113

1114

1115

The propagation of sublethal effects of neonicotinoids via trophic interactions was evidenced in a three-level food chain gathering wild strawberry (*Fragaria vesca*), wood cricket (*Nemobius sylvestris*) and nursery web spider (*Pisaura mirabili*): strawberries were treated with imidacloprid at different doses and crickets were allowed to feed on them (Uhl et al. 2015). In this tritrophic system, feeding, mass gain, thorax growth and mobility of wood crickets was reduced, and herbivory and predation diminished at sublethal imidacloprid doses in the non-target organisms (Uhl et al. 2015). The effects of thiamethoxam, applied as a soybean seed treatment, on interactions between soybeans, non-target herbivorous mollusks (pests), and predatory insects was studied in the laboratory and in the field (Douglas et al. 2015). In the laboratory, the slug *Deroceras reticulatum* was not affected by

thiamethoxam, but predatory ground beetles (*Chlaenius tricolor*) which ate these slugs were affected or died in over 60% of cases. In the field, thiamethoxam seed treatments decreased the activity and density of predatory arthropods, thereby releasing slug predation and reducing soybean densities by 19% and yield by 5%. The analyses of thiamethoxam residues revealed a transfer in food webs: they showed that insecticide concentrations decreased throughout the food chain, but that levels in slugs collected in the field were still high enough to adversely affect predatory insects. According to Douglas et al. (2015), this work on the trophic transfer of thiamethoxam challenges the idea that seed treatments with neonicotinoids specifically target herbivore pests, and underscores the need to consider predatory arthropods and soil organism communities in neonicotinoid risk assessment and management.

1116

1117

1118

1119

1120

1121

1122

1123

1124

1125

1126

1127

1128

1129

1130

1131

1132

1133

1134

1135

1136

1137

1138

1139

1140

1141

1142

1143

1144

If neonicotinoids can affect vertebrates through direct effects, as reviewed above, they can also affect wildlife through a reduction in food resources (Gibbons et al. 2015). Further, the trophic transfer of neonicotinoids has been recently evidenced, especially in birds. The presence of 54 residues of PPPs or transformation products was investigated in the food bolus (insects) provided by the parents of the tree swallow (Tachycineta bicolor) to their chicks, in 40 Canadian farms (Poisson et al. 2021). This multi-residue analysis included seven neonicotinoids (acetamiprid, clothianidin, dinotefuran, imidacloprid, nitenpyram, thiacloprid, thiamethoxam). The results attested to the ubiquitous trophic exposure, with nearly half of the food boluses showing contamination by at least one substance, clothianidin being among the most frequently detected PPPs (9%). Mixtures of 2 to 16 PPPs, among which five (clothianidin, dinotefuran, imidacloprid, thiacloprid, thiamethoxam) of the seven neonicotinoids, were also detected in 21% of the food boluses (and 45% of the contaminated boluses). A study conducted in Switzerland reported that at least one neonicotinoid was detected in 100% of food boluses collected from Alpine swift (Tachymarptis melba) provisioning their nestlings, 75% of the food boluses exhibiting measurable concentrations (Humann-Guilleminot et al. 2021). Both acetamiprid and thiacloprid were found, and thiacloprid showed the highest occurrence (up to 66.7%) and the highest concentrations (up to 0.6 μg/kg). Surveys on birds in the USA and Europe revealed exposure/accumulation of neonicotinoids in all trophic groups such as nectarivores and granivores, insectivores and predators including top-predators (raptors), and piscivores, strongly suggesting the occurrence of trophic transfer in food webs (Badry et al. 2021; Bishop et al. 2020; Bro et al. 2016; Byholm et al. 2018; Distefano et al. 2022; Humann-Guilleminot et al. 2021; Taliansky-Chamudis et al. 2017). In 60 sites over a wide cereal plain in France, the bioaccumulation of several neonicotinoids has been evidenced in both granivorous/omnivorous rodents, and insectivorous shrews as well as in earthworms and carabid beetles, which were their potential preys (Pelosi et al. 2021; Fritsch et al. 2022). Finally, residues in tissues have also been detected in terrestrial invertebrates and vertebrates, including wildlife species other than granivores (which can be exposed directly via ingestion of treated seeds) as detailed in previous sections (e.g., chiropterans).

1145

1146

1147

1148

1149

1150

1151

1152

1153

1154

1155

1156

1157

1158

1159

1160

1161

1162

1163

1164

1165

1166

1167

1168

1169

1170

1171

1172

1173

1174

Some studies highlighted the potential for neonicotinoids to negatively impact terrestrial insectivorous vertebrate abundance and diversity through indirect effects related to the reduction in quantity and quality of food resources. Such indirect effects have rarely been studied on vertebrates but Gibbons et al. (2015) showed that systemic insecticides can induce effects on wildlife via trophic cascades: the reduction in food supply related to the use of imidacloprid led to impairments in fish species.

Long before major publications based on large-scale correlative analyses between PPP use and population, Tennekes and Zillweger (2010) argued that neonicotinoid contamination of surface waters in Europe was one of the factors responsible for the continental-scale decline in insect biomass, which in turn led to many of the widespread declines in birds (golden oriole Oriolus oriolus, northern wheatear Oenanthe oenanthe, starling Stumus vulgaris...). This was studied by Hallmann et al. (2014) who observed that insectivorous bird populations in the Netherlans declined in areas with surface water concentrations of imidacloprid higher than 0.02 µg/L. Spatial differences in land-use changes related to agricultural intensification (urban area, natural area, cropped area, fertilizers) have been considered but they did not alter the significance of the observed effects. In the USA, Li et al. (2020) found that the increase in neonicotinoid use was related to reductions of 4% and 3% in grassland and insectivorous bird biodiversity, respectively, over 2008-2014. Such a trend was also found for non-grassland and non-insectivorous birds, with an average annual rate of reduction of 2%. Recently, Kraus et al. (2021) conducted surveys in wetlands of cropland and grassland landscapes which allowed to characterize cross-ecosystem fluxes of PPPs mediated by aquatic insect emergence, and discussed their implications for terrestrial insectivores. Aquatic insects were estimated to transfer fluxes ranging from 2 to 180 µg of total insecticides per wetland per day to the terrestrial ecosystem. Seven PPPs were detected in newly emerged insects, among which clothianidin and imidacloprid, and biomass of emerging aquatic insects was reduced up to 73% in cropland wetlands. The authors suggested that the availability of emerging adult aquatic insect prey for insectivores was reduced by insecticides, and that accumulated insecticide could be responsible for insectivore exposure to insect-borne PPPs. Along the observed gradient in PPP levels among the different wetlands, a decrease of 43% in insect emergence but an increase of 50% in insect-mediated PPP flux with increasing insecticide concentrations were reported (from 3 to 577 ng of insecticide per gram of insect) (Kraus et al. 2021). In addition, the presence of these neonicotinoids also led to a reduction in insect resources for consumer invertebrates (Kraus et al. 2021). Although bioaccumulation in organisms and transfer in food webs have been demonstrated together with sublethal and lethal effects propagated along food chains, the major process involved in shaping the impact of neonicotinoids in food webs is considered as being food web simplification (Tooker and Pearsons 2021). Such indirect effect of neonicotinoids affects both prey and predator populations through trophic cascade mechanisms and feedbacks. The initial decrease in resources when lower trophic levels are directly impacted by the use of the insecticides affect the dynamics of consumer populations at higher trophic levels through food scarcity (bottom-up control). When consumers are adversely impacted either directly (toxicity) or indirectly (lack of food supply), a subsequent decrease in predation occurs, affecting the dynamics of prey populations (top-down control). Compensatory mechanisms for consumers to overcome the decrease of one or a few food resources, such as switching to other food items, hardly occur when the predator of concern are specialist species, and seemed currently hampered in the case of neonicotinoids because of their widespread use (huge spatial extent worldwide, perennial and frequent use), the ubiquity of their environmental contamination, their broad toxicity to non-target fauna, and time-cumulative toxicity (Tooker and Pearsons 2021).

Aquatic ecosystems

Adverse effects of neonicotinoids can propagate through aquatic food webs via contaminated primary producers (Lima-Fernandes et al. 2019). Lima-Fernandes et al. (2019) used imidacloprid-contaminated and uncontaminated black alder tree (*Alnus glutinosa*) leaves to feed the stonefly shredder *Protonemura* sp., which were later given as prey to *Isoperla* sp. They showed that survival, body length and biomass of the shredders as well as leaf decomposition were 20% to 50% greater in the uncontaminated treatment in comparison to imidacloprid exposure. The biomass and length of predators were 11% and 4.3% higher, respectively, when fed with uncontaminated prey than when fed with imidacloprid exposed prey (Lima-Fernandes et al. 2019).Bioaccumulation of imidacloprid has been evidenced in both *Desmognathus* salamanders (*D. monticola* and *D. fuscus*) and benthic macroinvertebrates sampled from water streams adjacent to treated hemlock stands in the USA (Crayton et al. 2020), which represents a potential source of exposure for consumers at higher trophic levels. If exposure via the trophic route was likely for salamanders, several non-exclusive routes of exposure might be involved in the subsequent bioaccumulation, including dermal and dietary uptake (Crayton et al. 2020).

Hayasaka et al. (2012) showed that successive applications of imidacloprid and the phenylpyrazole insecticide fipronil (also a systemic insecticide) in experimental rice fields resulted in reduced growth of medaka fish, *Oryzias latipes*, adults and fry, most likely through reduced medaka prey abundance. Indeed, the concentrations (approximately 1 to 50 μ g/L) were too low to have a direct effect on fish. As indicated above, the

decline of emerging insects from aquatic ecosystems towards riparian and surrounded terrestrial landscapes strongly decrease the prey availability for numerous consumers, and overall minor energy transfer across ecosystems (Kraus et al. 2021).

In a Japanese lacustrine ecosystem, Yamamuro et al. (2019) demonstrated the existing relationship between decline in fishery yields and neonicotinoids. The use of neonicotinoids on watersheds since 1993 coincided with an 83% decrease in average zooplankton biomass in spring, causing the smelt (*H. nipponensis*) harvest to collapse from 240 to 22 tons. Young smelts consume zooplankton crustaceans, and their decreased abundance was linked to the reduction of zooplankton biomass caused by the introduction of neonicotinoids. This study demonstrates the indirect effects of neonicotinoids along an aquatic food web through cascading effects.

Waterbirds living and feeding in lakes and ponds (ducks, waders, cormorants...) may depend on aquatic invertebrates as their food source. Consequently, the depletion of this food source must necessarily affect them (Sánchez-Bayo et al. 2016). Duckling abundance is thus related to aquatic macroinvertebrate abundance, which is consistent with other studies, and collectively suggests that neonicotinoids contamination could influence duckling abundance indirectly by impacting aquatic macroinvertebrate communities (Tyler 2022). The available data indicate that the effects of neonicotinoids on aquatic bird life are indirect, as for other bird families, and are associated with the direct toxic impacts of these contaminants on invertebrates (Sánchez-Bayo et al. 2016).

Conclusion

Neonicotinoids, in particular imidacloprid, and to a lesser extent thiamethoxam and clothianidin, are very frequently detected in soils and freshwaters, even several years after their use. In addition, the presence of acetamiprid, imidacloprid, thiacloprid and thiamethoxam was observed in the air. Neonicotinoids have only been recently monitored in coastal and marine environments (since 2010s), but many studies report the presence of imidacloprid and thiamethoxam in different transitional ecosystems such as Mediterranean lagoons.

This contamination of the environment leads to the exposure of non-target organisms and impacts biodiversity. The ecotoxicological effects of neonicotinoids depend on the studied organisms, but this review showed that these substances have particularly high direct and indirect impacts on terrestrial invertebrates and vertebrates, and on aquatic invertebrates. The impacts on aquatic vertebrates are less documented.

The effects of neonicotinoids on terrestrial heterotrophic microorganisms vary according to the conditions: in field studies, these substances have little or no effect, while in the laboratory, impacts on the structure and on different microbial activities were observed (however, the tested concentrations are sometimes unrealistic).

Laboratory studies are not always environmentally relevant, but they are complementary to field approaches as they can help to understand the effects at lower levels of biological organization (sub-individual, individual) that have consequences on higher levels (populations, community) observed in the field. Although contradictory results have been noted in the literature, neonicotinoids have negative effects (mortality, mobility disturbance) at the individual level on pollinators (honeybees in particular). In addition, exposure to neonicotinoids increases the susceptibility of honeybees to diseases and pests. Despite the importance of wild pollinators and their crucial role in pollination, the number of studies focused on the impacts of neonicotinoids on this highly diverse group of organisms is very limited. Furthermore, neonicotinoids have been shown to have effects on other terrestrial invertebrates such as natural enemies, earthworms or nematodes. Neonicotinoids are also largely involved in the decline of birds. Consumption of treated seeds is mainly responsible for neonicotinoid direct poisoning, but birds could be exposed to these insecticides especially by trophic route after consumption of contaminated insects. Neonicotinoids have negative effects on bats, amphibians, and on reptiles (though available data are still scarce for this group). For aquatic invertebrates and vertebrates, the data on the effects of neonicotinoids remain limited. The available results indicate correlations between neonicotinoid concentrations and declines in arthropod taxa. Neonicotinoids seem to be not very toxic to aquatic vertebrates such as fish, but recent studies provide worrying results for amphibians. However, the number of studies remains low and few studies focused on marine organisms. In addition to their toxicity to directly exposed organisms, neonicotinoid-induced indirect effects via trophic cascades have been demonstrated to affect some species (terrestrial and aquatic invertebrates) but data are still too few to get a clear picture.

This critical review highlighted numerous knowledge gaps. First, there was a lack of data regarding the effects of neonicotinoids on primary producers (although the mode of action of neonicotinoids is unlikely to result in effects; Anderson et al. 2015), aquatic heterotrophic microorganisms, wild pollinators, raptors, mammals, reptiles, amphibians, aquatic vertebrates, and on organisms in the marine environment in general. In addition: (1) the majority of studies focused on only one neonicotinoid making generalization difficult; (2) while imidacloprid is the most commonly studied neonicotinoid, data are limited for the other substances; (3) most laboratory studies do not reflect realistic and representative uses under in field application conditions; (4) very few studies consider transformation products and mixtures with other PPPs; (5) the number of studies considering the impact of neonicotinoids on high levels of biological organization (i.e., beyond individual and population) is low; (6) the effects of neonicotinoids on maintenance of pest regulation and soil functions are hardly reported; (7) there is a lack of time series to survey mid- or long-term effects as well as post-exposure effects; (8) there is a lack of data

regarding the effects of neonicotinoids on ecosystem functioning and services, yet the few existing studies suggest that they might significantly alter important provision and regulation ecosystem services (Pesce et al. 2023). More research remains to be done to better characterize the impacts of neonicotinoids to protect biodiversity.

Acknowledgements This review was done under the framework of the French collective scientific assessment (CSA) about "the impacts of plant protection products on biodiversity and ecosystem services" coordinated by INRAE and Ifremer for the French Ministries of Ecology, for Research, and for Agriculture. The authors would like to acknowledge Dr Thierry Caquet, the INRAE scientific Director of Environment, and the general directorate of the Ifremer; Dr Guy Richard, head of the INRAE Directorate for Collective Scientific Assessment, Foresight and Advanced Studies (DEPE); and Lucile Wargniez for illustrations.

Declarations

- Ethics approval and consent to participate: Not applicable
- 1278 Consent to publish: Not applicable

Authors contributions Laure Mamy, Stéphane Pesce, Wilfried Sanchez, Stéphanie Aviron, Carole Bedos, Philippe Berny, Colette Bertrand, Stéphane Betoulle, Sandrine Charles, Arnaud Chaumot, Michael Coeurdassier, Marie-Agnès Coutellec, Olivier Crouzet, Juliette Faburé, Clémentine Fritsch, Patrice Gonzalez, Mickael Hedde, Christophe Leboulanger, Christelle Margoum, Christian Mougin, Dominique Munaron, Sylvie Nélieu, Céline Pelosi, Magali Rault, Eliott Sucré, Marielle Thomas, Julien Tournebize, and Sophie Leenhardt performed the review of the literature. Anne-Laure Achard, Morgane Le Gall and Sophie Le Perchec conducted the literature search, the paper collection, and the bibliometric analysis. Laure Mamy drafted the first version of the manuscript based on this original contribution. Laure Mamy, Stéphane Pesce, Wilfried Sanchez, Stéphanie Aviron, Carole Bedos, Philippe Berny, Colette Bertrand, Stéphane Betoulle, Sandrine Charles, Arnaud Chaumot, Michael Coeurdassier, Marie-Agnès Coutellec, Olivier Crouzet, Juliette Faburé, Clémentine Fritsch, Patrice Gonzalez, Mickael Hedde, Christophe Leboulanger, Christelle Margoum, Christian Mougin, Dominique Munaron, Sylvie Nélieu, Céline Pelosi, Magali Rault, Eliott Sucré, Marielle Thomas, Julien Tournebize, and Sophie Leenhardt contributed to subsequent revisions to the manuscript. All authors approved the final submitted version.

- **Funding** This work was funded by the French Office for Biodiversity (OFB) through the national Ecophyto plan.
- **Competing interests:** The authors declare that they have no competing interests
- 1294 Availability of data and materials: Not applicable

1295 References

1296	Adriaanse P, Arce A, Focks A, Ingels B, Daniela J, Del Aguila M, Ercolano V, Ferilli F (2023) Guidance on the
1297	risk assessment of plant protection products on bees (Apis mellifera, Bombus spp. and solitary bees). EFSA
1298	J 21:7989. https://doi.org/10.2903/j.efsa.2023.7989
1299	Ahemad M, Khan MS (2011) Effects of insecticides on plant-growth-promoting activities of phosphate
1300	solubilizing rhizobacterium Klebsiella sp strain PS19. Pestic Biochem Physiol 100(1):51-56.
1301	http://dx.doi.org/10.1016/j.pestbp.2011.02.004
1302	Alexander AC, Luis AT, Culp JM, Baird DJ, Cessna AJ (2013) Can nutrients mask community responses to
1303	insecticide mixtures? Ecotoxicology 22(7):1085-1100. http://dx.doi.org/10.1007/s10646-013-1096-3
1304	Alford AM, Krupke CH (2017) Translocation of the neonicotinoid seed treatment clothianidin in maize. Plos One
1305	12:e0173836. http://dx.doi.org/10.1371/journal.pone.0173836
1306	Alford AM, Krupke CH (2019) Movement of the neonicotinoid seed treatment clothianidin into groundwater,
1307	aquatic plants, and insect herbivores. Environ Sci Technol 53(24):14368-14376.
1308	http://dx.doi.org/10.1021/acs.est.9b05025.
1309	Alkassab AT, Kunz N, Bischoff G, Pistorius J (2020) Comparing response of buff-tailed bumblebees and red
1310	mason bees to application of a thiacloprid-prochloraz mixture under semi-field conditions. Ecotoxicology
1311	29(7):846-855. http://dx.doi.org/10.1007/s10646-020-02223-2
1312	Alsafran M, Rizwan M, Usman K, Saleem MH, Al Jabri H (2022) Neonicotinoid insecticides in the environment:
1313	A critical review of their distribution, transport, fate, and toxic effects. J Environ Chem Engineer 10:108485.
1314	https://doi.org/10.1016/j.jece.2022.108485
1315	Anderson JC, Dubetz C, Palace VP (2015) Neonicotinoids in the Canadian aquatic environment: A literature
1316	review on current use products with a focus on fate, exposure, and biological effects. Sci Total Environ
1317	505:409-422. http://dx.doi.org/10.1016/j.scitotenv.2014.09.090
1318	Anderson MJ, Valdiviezo A, Conway MH, Farrell C, Andringa RK, Janik A, Chiu WA, Rusyn I, Hamer SA (2023)
1319	Imidacloprid exposure is detectable in over one third of wild bird samples from diverse Texas ecoregions.
1320	Sci Total Environ 876:162723. https://doi.org/10.1016/j.scitotenv.2023.162723
1321	Arena M, Sgolastra F (2014) A meta-analysis comparing the sensitivity of bees to pesticides. Ecotoxicology
1322	23(3):324-334. http://dx.doi.org/10.1007/s10646-014-1190-1
1323	Armbrust KL, Peeler HB (2002) Effects of formulation on the run-off of imidacloprid from turf. Pest Manage Sci
1324	58(7):702-706. http://dx.doi.org/10.1002/ps.518

1325	Auby I, Bost CA, Budzinski H (2011) Regression des nerbiers de zosteres dans le Bassin d'Arcachon : état des
1326	lieux et recherche des causes. Rapport Ifremer RST/LER/AR/11.007, 195.
1327	Avery ML, Decker DG, Fischer DL (1994) Cage and flight pen evaluation of avian repellency and hazard
1328	associated with imidacloprid-treated rice seed. Crop Protec 13(7):535-540. http://dx.doi.org/10.1016/0261-
1329	2194(94)90107-4
1330	Badry A, Schenke D, Treu G, Krone O (2021) Linking landscape composition and biological factors with exposure
1331	levels of rodenticides and agrochemicals in avian apex predators from Germany. Environ Res 193:110602.
1332	http://dx.doi.org/10.1016/j.envres.2020.110602
1333	Badry A, Schenke D, Brücher H, Chakarov N, Grünkorn T, Illner H, Krüger O, Marczak T, Müskens G, Nachtigall
1334	W, Zollinger R, Treu G, Krone O (2022) Spatial variation of rodenticides and emerging contaminants in
1335	blood of raptor nestlings from Germany. Environ Sci Pollut Res 29:60908-60921.
1336	https://doi.org/10.1007/s11356-022-20089-1
1337	Baldissera MD, Souza CF, Golombieski JI, Seben D, Sippert LR, Salbego J, Zanella R, Baldisserotto B (2018)
1338	Thiamethoxam induced hepatic energy changes in silver catfish via impairment of the phosphoryl transfer
1339	network pathway: Toxicological effects on energetics homeostasis. Environ Tox Pharm 60:1-4.
1340	https://doi.org/10.1016/j.etap.2018.04.002
1341	Barmentlo SH, Schrama M, van Bodegom PM, de Snoo GR, Musters CJM, Vijver MG (2019) Neonicotinoids and
1342	fertilizers jointly structure naturally assembled freshwater macroinvertebrate communities. Sci Total Environ
1343	691:36-44. http://dx.doi.org/10.1016/j.scitotenv.2019.07.110
1344	Barmentlo SH, Schrama M, de Snoo GR, van Bodegom PM, van Nieuwenhuijzen A, Vijver MG (2021)
1345	Experimental evidence for neonicotinoid driven decline in aquatic emerging insects. Proc Natl Acad Sci USA
1346	118:e2105692118. http://dx.doi.org/10.1073/pnas.2105692118
1347	Bass C, Denholm I, Williamson MS, Nauen R (2015) The global status of insect resistance to neonicotinoid
1348	insecticides. Pestic Biochem Physiol 121:78-87. https://doi.org/10.1016/j.pestbp.2015.04.004
1349	Bayat S, Geiser F, Kristiansen P, Wilson SC (2014) Organic contaminants in bats: Trends and new issues. Environ
1350	Int 63:40-52. http://dx.doi.org/10.1016/j.envint.2013.10.009
1351	Becher MA, Grimm V, Thorbek P, Horn J, Kennedy PJ, Osborne JL (2014) BEEHAVE: a systems model of
1352	honeybee colony dynamics and foraging to explore multifactorial causes of colony failure. J Appl Ecol
1353	51·470-482 https://doi.org/https://doi.org/10.1111/1365-2664.12222

1354	Becher MA, Twiston-Davies G, Penny TD, Goulson D, Rotheray EL, Osborne JL (2018) Bumble-BEEHAVE: A
1355	systems model for exploring multifactorial causes of bumblebee decline at individual, colony, population and
1356	community level. J Appl Ecol 55:2790-2801. https://doi.org/https://doi.org/10.1111/1365-2664.13165
1357	Becker JM, Liess M (2017) Species diversity hinders adaptation to toxicants. Environ Sci Technol 51(17):10195-
1358	10202. http://dx.doi.org/10.1021/acs.est.7b02440
1359	Becker JM, Russo R, Shahid N, Liess M (2020) Drivers of pesticide resistance in freshwater amphipods. Sci Total
1360	Environ 735:139624. http://dx.doi.org/10.1016/j.scitotenv.2020.139264
1361	Beers EH, Brunner JF, Dunley JE, Doerr M, Granger K (2005) Role of neonicotinyl insecticides in Washington
1362	apple integrated pest management. Part II. Nontarget effects on integrated mite control. J Insect Sci 5:16.
1363	http://dx.doi.org/10.1093/jis/5.1.16
1364	Beketov MA, Liess M (2008) Acute and delayed effects of the neonicotinoid insecticide thiacloprid on seven
1365	freshwater arthropods. Environ Tox Chem 27(2):461-470. http://dx.doi.org/10.1897/07-322r.1
1366	Belsky J, Joshi NK (2020) Effects of fungicide and herbicide chemical exposure on Apis and non-Apis bees in
1367	agricultural landscape. Front Environ Sci 8:81. http://dx.doi.org/10.3389/fenvs.2020.00081
1368	Berny PJ, Buronfosse R, Videmann B, Buronfosse T (1999) Evaluation of the toxicity of imidacloprid in wild
1369	birds. A new high performance thin layer chromatography (HPTLC) method for the analysis of liver and crop
1370	samples in suspected poisoning cases. J Liq Chrom Rel Technol 22(10):1547-1559.
1371	http://dx.doi.org/10.1081/jlc-100101750
1372	Bishop CA, Moran AJ, Toshack MC, Elle E, Maisonneuve F, Elliott JE (2018) Hummingbirds and bumble bees
1373	exposed to neonicotinoid and organophosphate insecticides in the Fraser Valley, British Columbia, Canada.
1374	Environ Tox Chem 37(8):2143-2152. http://dx.doi.org/10.1002/etc.4174
1375	Bishop CA, Woundneh MB, Maisonneuve F, Common J, Elliott JE, Moran AJ (2020) Determination of
1376	neonicotinoids and butenolide residues in avian and insect pollinators and their ambient environment in
1377	Western Canada (2017, 2018). Sci Total Environ 737:139386.
1378	http://dx.doi.org/10.1016/j.scitotenv.2020.139386
1379	Bonmatin JM, Moineau I, Charvet R, Colin ME, Fleche C, Bengsch ER (2005a) Behaviour of imidacloprid in
1380	fields. Toxicity for honey bees. In: Lichtfouse E, Schwarzbauer J, Robert D (eds). Environmental chemistry.
1381	Springer, Berlin. pp. 483–494. http://link.springer.com/chapter/10.1007%2F3-540-26531-7 44.

1382 Bonmatin JM, Marchand PA, Charvet R, Moineau I, Bengsch ER, Colin ME (2005b) Quantification of 1383 Food imidacloprid J Agric Chem 53(13):5336-5341. uptake in maize crops. 1384 https://doi.org/10.1021/jf0479362 1385 Bonmatin JM, Giorio C, Girolami V, Goulson D, Kreutzweiser DP, Krupke C, Liess M, Long E, Marzaro M, 1386 Mitchell EAD, Noome DA, Simon-Delso N, Tapparo A (2015) Environmental fate and exposure, 1387 neonicotinoids and fipronil. Environ Sci Pollut Res 22(1):35-67. https://doi.org/10.1007/s11356-014-3332-7 1388 Botias C, David A, Horwood J, Abdul-Sada A, Nicholls E, Hill E, Goulson D (2015) Neonicotinoid residues in 1389 wildflowers, a potential route of chronic exposure for bees. Environ Sci Technol 49(21):12731-12740. 1390 https://doi.org/10.1021/acs.est.5b03459 1391 Botias C, David A, Hill EM, Goulson D (2016) Contamination of wild plants near neonicotinoid seed-treated 1392 crops, and implications for non-target insects. Sci Total Environ 566:269-278. 1393 http://dx.doi.org/10.1016/j.scitotenv.2016.05.065 1394 Botina LL, Bernardes RC, Barbosa WF, Lima MAP, Guedes RNC, Martins GF (2020) Toxicological assessments 1395 of agrochemical effects on stingless bees (Apidae, Meliponini). MethodsX 1396 http://dx.doi.org/10.1016/j.mex.2020.100906 1397 Bradford BR, Whidden E, Gervasio ED, Checchi PM, Raley-Susman KM (2020) Neonicotinoid-containing 1398 insecticide disruption of growth, locomotion, and fertility in Caenorhabditis elegans. Plos One 15:e0238637. 1399 https://doi.org/10.1371/journal.pone.0238637 1400 Bredeson MM, Lundgren JG (2018) Thiamethoxam seed treatments reduce foliar predator and pollinator 1401 populations in sunflowers (Helianthus annuus), and extra-floral nectaries as a route of exposure for seed 1402 treatments to affect the predator, Coleomegilla maculata (Coleoptera: Coccinellidae). Crop Protec 106:86-1403 92. http://dx.doi.org/10.1016/j.cropro.2017.12.019 1404 Bredeson MM, Reese RN, Lundgren JG (2015) The effects of insecticide dose and herbivore density on tri-trophic 1405 effects of thiamethoxam in a system involving wheat, aphids, and ladybeetles. Crop Protec 69:70-76. 1406 http://dx.doi.org/10.1016/j.cropro.2014.12.010 1407 Bro E, Decors A, Millot F, Soyez D, Moinet M, Berny P, Mastain O (2010) Intoxications des perdrix grises en 1408 nature. Nouveau bilan de la surveillance SAGIR. Faune Sauvage 289:26-32. 1409 Bro E, Devillers J, Millot F, Decors A (2016) Residues of plant protection products in grey partridge eggs in 1410 French cereal ecosystems. Environ Sci Pollut Res 23(10):9559-9573. http://dx.doi.org/10.1007/s11356-016-

1411

6093-7

1412	Brodeur JC, Damonte MJ, Rojas DE, Cristos D, Vargas C, Poliserpi MB, Andriulo AE (2022) Concentration of
1413	current-use pesticides in frogs from the Pampa region and correlation of a mixture toxicity index with
1414	biological effects. Environ Res 204:112354. https://doi.org/10.1016/j.envres.2021.112354
1415	Buchweitz JP, Viner TC, Lehner AF (2019) Qualitative identification of imidacloprid in postmortem animal tissue
1416	by gas chromatography-tandem mass spectrometry. Toxicol Mech Meth 29(7):511-517.
1417	http://dx.doi.org/10.1080/15376516.2019.1616344
1418	Byholm P, Makelainen S, Santangeli A, Goulson D (2018) First evidence of neonicotinoid residues in a long-
1419	distance migratory raptor, the European honey buzzard (Pernis apivorus). Sci Total Environ 639:929-933.
1420	http://dx.doi.org/10.1016/j.scitotenv.2018.05.185
1421	Camp AA, Batres MA, Williams WC, Koethe RW, Stoner KA, Lehmann DM (2020) Effects of the neonicotinoid
1422	acetamiprid in pollen on Bombus impatiens microcolony development. Environ Tox Chem 39(12):2560-
1423	2569. http://dx.doi.org/10.1002/etc.4886
1424	Camp AA, Lehmann DM (2021) Impacts of neonicotinoids on the bumble bees Bombus terrestris and Bombus
1425	impatiens examined through the lens of an Adverse Outcome Pathway framework. Environ Tox Chem
1426	40(2):309-322. http://dx.doi.org/10.1002/etc.4939
1427	Campbell KS, Keller PG, Heinzel LM, Golovko SA, Seeger DR, Golovko MY, Kerby JL (2022) Detection of
1428	imidacloprid and metabolites in Northern Leopard frog (Rana pipiens) brains. Sci Total Environ 813:152424.
1429	http://dx.doi.org/10.1016/j.scitotenv.2021.152424
1430	Campbell KS, Keller P, Golovko SA, Seeger D, Golovko MY, Kerby JL (2023) Connecting the pipes: Agricultural
1431	tile drains and elevated imidacloprid brain concentrations in juvenile Northern Leopard frogs (Rana pipiens).
1432	Environ Sci Technol 57:2758-2767. https://doi.org/10.1021/acs.est.2c06527
1433	Cavallaro MC, Liber K, Headley JV, Peru KM, Morrissey CA (2018) Community-level and phenological
1434	responses of emerging aquatic insects exposed to 3 neonicotinoid insecticides: An in situ wetland limnocorral
1435	approach. Environ Tox Chem 37(9):2401-2412. http://dx.doi.org/10.1002/etc.4187
1436	Cavallaro MC, Main AR, Liber K, Phillips LD, Headley JV, Peru KM, Morrissey CA (2019) Neonicotinoids and
1437	other agricultural stressors collectively modify aquatic insect communities. Chemosphere 226:945-955.
1438	http://dx.doi.org/10.1016/j.chemosphere.2019.03.176
1439	Chiaia-Hernandez AC, Keller A, Wächter D, Steinlin C, Camenzuli L, Hollender J, Krauss M (2017) Long-term
1440	persistence of pesticides and TPs in archived agricultural soil samples and comparison with pesticide
1441	application Environ Sci Technol 51:10642-10651, http://dx.doi.org/10.1021/acs.est.7b02529

1442	Chan DSW, Raine NE (2021) Population decline in a ground-nesting solitary squash bee (Eucera pruinosa)
1443	following exposure to a neonicotinoid insecticide treated crop (Cucurbita pepo). Scientif Rep 11:4241.
1444	http://dx.doi.org/10.1038/s41598-021-83341-7
1445	Chara-Serna AM, Epele LB, Morrissey CA, Richardson JS (2019) Nutrients and sediment modify the impacts of
1446	a neonicotinoid insecticide on freshwater community structure and ecosystem functioning. Sci Total Environ
1447	692:1291-1303. http://dx.doi.org/10.1016/j.scitotenv.2019.06.301
1448	Chauzat MP, Faucon JP, Martel AC, Lachaize J, Cougoule N, Aubert M (2006) A survey of pesticide residues in
1449	pollen loads collected by honey bees in France. J Econ Entomol 99(2):253-262.
1450	Chauzat MP, Martel AC, Cougoule N, Porta P, Lachaize J, Zeggane S, Aubert M, Carpentier P, Faucon JP (2011)
1451	An assessment of honeybee colony matrices, apis mellifera (hymenoptera apidae) to monitor pesticide
1452	presence in continental France. Environ Tox Chem 30(1):103-111. https://doi.org/10.1002/etc.361
1453	Chen Y, Zheng X, Liu J, Wei H, Chen YD, Su XX, Zhang J (2016) Appraisal of the impact of three insecticides
1454	on the principal rice pests and their predators in China. Florida Entomol 99(2):210-220.
1455	http://dx.doi.org/10.1653/024.099.0209
1456	Chen Y, Zhang L, Hu H, Wu R, Ling J, Yue S, Yang D, Yu W, Du W, Shen G, Zhao M (2022) Neonicotinoid
1457	pollution in marine sediments of the East China Sea. Sci Total Environ 842:156658.
1458	https://doi.org/10.1016/j.scitotenv.2022.156658
1459	Coscollà C, Yusà V (2016) Chapter 17 - Pesticides and agricultural air quality. In: de la Guardia, M., Armenta, S.,
1460	eds. Comprehensive Analytical Chemistry, Elsevier, 423-490.
1461	https://www.sciencedirect.com/science/article/pii/S0166526X16300654
1462	Crayton SM, Wood PB, Brown DJ, Millikin AR, McManus TJ, Simpson TJ, Ku KM, Park YL (2020)
1463	Bioaccumulation of the pesticide imidacloprid in stream organisms and sublethal effects on salamanders.
1464	Glob Ecol Conserv 24:e01292. https://doi.org/10.1016/j.gecco.2020.e01292
1465	Criquet J, Dumoulin D, Howsam M, Mondamert L, Goossens JF, Prygiel J, Billon G (2017) Comparison of POCIS
1466	passive samplers vs. composite water sampling: A case study. Sci Total Environ 609:982-991.
1467	https://doi.org/10.1016/j.scitotenv.2017.07.227
1468	Cycoń M, Piotrowska-Seget Z, (2015a) Biochemical and microbial soil functioning after application of the
1469	insecticide imidacloprid. J Environ Sci 27:147-158. http://dx.doi.org/10.1016/j.jes.2014.05.034

1470	Cycoń M, Piotrowska-Seget Z (2015b) Community structure of ammonia-oxidizing archaea and ammonia-
1471	oxidizing bacteria in soil treated with the insecticide imidacloprid. Biomed Res Int 2015:582938.
1472	http://dx.doi.org/10.1155/2015/582938
1473	Davis JM, Rosemond AD, Eggert SL, Cross WF, Wallace JB (2010) Long-term nutrient enrichment decouples
1474	predator and prey production. Proc Natio Acad Sciences USA, 107(1):121-126.
1475	http://dx.doi.org/10.1073/pnas.0908497107
1476	de Araújo EP, Dutra Caldas E, Oliveira-Filho EC (2022) Pesticides in surface freshwater: a critical review. Environ
1477	Monit Assess 194:452. http://dx.doi.org/10.1007/s10661-022-10005-y
1478	de Snoo GR, Scheidegger NMI, de Jong FMW (1999) Vertebrate wildlife incidents with pesticides: a European
1479	survey. Pestic Sci 55(1):47-54. https://doi.org/10.1002/(sici)1096-9063(199901)55:1<47::aid-
1480	ps859>3.3.co,2-r
1481	Demortain D (2021) The science behind the ban: the outstanding impact of ecotoxicological research on the
1482	regulation of neonicotinoids. Curr Op Insect Sci 46:78-82. http://dx.doi.org/10.1016/j.cois.2021.02.017
1483	Désert M, Ravier S, Gille G, Quinapallo A, Armengaud A, Pochet G, Savelli JL, Wortham H, Quivet E (2018)
1484	Spatial and temporal distribution of current-use pesticides in ambient air of Provence-Alpes-Côte-d'Azur
1485	Region and Corsica, France. Atm Environ 192:241-256. https://doi.org/10.1016/j.atmosenv.2018.08.054
1486	Disque HH, Hamby KA, Dubey A, Taylor C, Dively GP (2019) Effects of clothianidin-treated seed on the
1487	arthropod community in a mid-Atlantic no-till corn agroecosystem. Pest Manage Sci 75 (4):969-978.
1488	http://dx.doi.org/10.1002/ps.5201
1489	Distefano GG, Zangrando R, Basso M, Panzarin L, Gambaro A, Volpi Ghirardini A, Picone M (2022) The ubiquity
1490	of neonicotinoid contamination: Residues in seabirds with different trophic habits. Environ Res 206:112637.
1491	https://doi.org/10.1016/j.envres.2021.112637
1492	Dittbrenner N, Triebskorn R, Moser I, Capowiez Y (2010) Physiological and behavioural effects of imidacloprid
1493	on two ecologically relevant earthworm species (Lumbricus terrestris and Aporrectodea caliginosa).
1494	Ecotoxicology 19:1567-1573.
1495	Dittbrenner N, Moser I, Triebskorn R, Capowiez Y (2011a) Assessment of short and long-term effects of
1496	imidacloprid on the burrowing behaviour of two earthworm species (Aporrectodea caliginosa and Lumbricus
1497	terrestris) by using 2D and 3D post-exposure techniques. Chemosphere 84:1349-1355.

1498	Dittorenner N, Schmitt H, Capowiez Y, Triebskorn R (2011b) Sensitivity of Eisenia fetida in comparison to
1499	Aporrectodea caliginosa and Lumbricus terrestris after imidacloprid exposure. Body mass change and
1500	histopathology. J Soils Sed 11:1000-1010.
1501	Douglas MR, Rohr JR, Tooker JF (2015) Neonicotinoid insecticide travels through a soil food chain, disrupting
1502	biological control of non-target pests and decreasing soya bean yield. J Appl Ecol 52(1):250-260.
1503	http://dx.doi.org/10.1111/1365-2664.12372
1504	Douglas MR, Tooker JF (2016) Meta-analysis reveals that seed-applied neonicotinoids and pyrethroids have
1505	similar negative effects on abundance of arthropod natural enemies. Peerj 4:e2776.
1506	http://dx.doi.org/10.7717/peerj.2776
1507	Dubey A, Lewis MT, Dively GP, Hamby KA (2020) Ecological impacts of pesticide seed treatments on arthropod
1508	communities in a grain crop rotation. J Appl Ecol 57(5):936-951. http://dx.doi.org/10.1111/1365-2664.13595
1509	Duso C, Ahmad S, Tirello P, Pozzebon A, Klaric V, Baldessari M, Malagnini V, Angeli G (2014) The impact of
1510	insecticides applied in apple orchards on the predatory mite Kampimodromus aberrans (Acari: Phytoseiidae).
1511	Exp Appl Acarol 62(3):391-414. http://dx.doi.org/10.1007/s10493-013-9741-3
1512	EFSA (2018) https://www.efsa.europa.eu/en/press/news/180228. Accessed 20 September 2023
1513	EFSA PPR Panel (2015) Statement on the suitability of the BEEHAVE model for its potential use in a regulatory
1514	context and for the risk assessment of multiple stressors in honeybees at the landscape level. EFSA J 13:4125.
1515	https://doi.org/10.2903/j.efsa.2015.4125
1516	EFSA Scientific Committee, More S, Bampidis V, Benford D, Bragard C, Halldorsson T, Hernández-Jerez A,
1517	Bennekou SH, Koutsoumanis K, Machera K, Naegeli H, Nielsen SS, Schlatter J, Schrenk D, Silano V, Turck
1518	D, Younes M, Arnold G, Dorne J-L, Maggiore A, Pagani S, Szentes C, Terry S, Tosi S, Vrbos D, Zamariola
1519	G and Rortais A (2021) Scientific Opinion on a systems-based approach to the environmental risk assessment
1520	of multiple stressors in honey bees. EFSA J 19(5):6607. https://doi.org/10.2903/j.efsa.2021.6607
1521	Eng ML, Stutchbury BJM, Morrissey CA (2017) Imidacloprid and chlorpyrifos insecticides impair migratory
1522	ability in a seed-eating songbird. Scient Rep 7:15176. http://dx.doi.org/10.1038/s41598-017-15446-x
1523	Eng ML, Stutchbury BJM, Morrissey CA (2019) A neonicotinoid insecticide reduces fueling and delays migration
1524	in songbirds. Science 365(6458):1177-1180. http://dx.doi.org/10.1126/science.aaw9419
1525	English SG, Sandoval-Herrera NI, Bishop CA, Cartwright M, Maisonneuve F, Elliott JE, Welch KC (2021)
1526	Neonicotinoid pesticides exert metabolic effects on avian pollinators. Scient Rep 11:2914.
1527	http://dx.doi.org/10.1038/s41598-021-82470-3

1528 Ertl HM, Mora MA, Brightsmith DJ, Navarro-Alberto JA (2018) Potential impact of neonicotinoid use on Northern 1529 bobwhite (Colinus virginianus) in Texas: A Plos One 13:0191100. historical analysis. 1530 https://doi.org/10.1371/journal.pone.0191100 1531 Esquivel CJ, Martinez EJ, Baxter R, Trabanino R, Ranger CM, Michel A, Canas LA (2020) Thiamethoxam 1532 differentially impacts the survival of the generalist predators, Orius insidiosus (Hemiptera: Anthocoridae) 1533 and Hippodamia convergens (Coleoptera: Coccinellidae), when exposed via the food chain. J Insect Sci 1534 20(4):13, 1-10. http://dx.doi.org/10.1093/jisesa/ieaa070 EU 1535 Pesticides https://ec.europa.eu/food/plant/pesticides/eu-pesticidesdatabase (2023)1536 database/start/screen/active-substances. Accessed 20 September 2023 1537 European Commission (2023) https://food.ec.europa.eu/plants/pesticides/approval-active-substances/renewal-1538 approval/neonicotinoids en. Accessed 20 September 2023 1539 Evelsizer V, Skopec M (2018) Pesticides, including neonicotinoids, in drained wetlands of Iowa's prairie pothole 1540 region. Wetlands 38(2):221-232. http://dx.doi.org/10.1007/s13157-016-0796-x 1541 Faria M, Wu X, Luja-Mondragón M, Prats E, Gómez-Oliván LM, Piña B, Raldúa D (2020) Screening anti-predator 1542 behaviour in fish larvae exposed to environmental pollutants. Sci Total Environ 714:136759. 1543 https://doi.org/10.1016/j.scitotenv.2020.136759 1544 Fauser A, Sandrock C, Neumann P, Sadd B (2017) Neonicotinoids override a parasite exposure impact on 1545 hibernation success of a key bumblebee pollinator. **Ecol** Entomol 42(3):306-314. 1546 http://dx.doi.org/10.1111/een.12385 1547 Filimon MN, Voia SO, Popescu R, Dumitrescu G, Ciochina LP, Mituletu M, Vlad DC (2015) The effect of some 1548 insecticides on soil microorganisms based on enzymatic and bacteriological analyses. Rom Biotechnol Lett 1549 20(3):10439-10447. 1550 Fountain MT, Harris AL (2015) Non-target consequences of insecticides used in apple and pear orchards on 1551 Forficula auricularia L. (Dermaptera: Forficulidae). Biol Control 91:27-33. 1552 http://dx.doi.org/10.1016/j.biocontrol.2015.07.007 Fountain MT, Medd N (2015) Integrating pesticides and predatory mites in soft fruit crops. Phytoparasitica 1553 1554 43(5):657-667. http://dx.doi.org/10.1007/s12600-015-0485-y 1555 Frame ST, Pearsons KA, Elkin KR, Saporito LS, Preisendanz HE, Karsten HD, Tooker JF (2021) Assessing 1556 surface and subsurface transport of neonicotinoid insecticides from no-till crop fields. J Environ Qual

50(2):476-484. http://dx.doi.org/10.1002/jeq2.20185

- 1558 Frank SD, Tooker JF (2020) Neonicotinoids pose undocumented threats to food webs. Proc Natio Acad Sci USA, 1559 117:22609-22613. https://doi.org/10.1073/pnas.2017221117 1560 Fritsch C, Appenzeller B, Burkart L, Coeurdassier M, Scheifler R, Raoul F, Driget V, Powolny T, Gagnaison C, 1561 Rieffel D, Afonso E, Goydadin AC, Hardy EM, Palazzi P, Schaeffer C, Gaba S, Bretagnolle V, Bertrand C, 1562 Pelosi C (2022) Pervasive exposure of wild small mammals to legacy and currently used pesticide mixtures 1563 in arable landscapes. Sci Rep 12:15904. https://doi.org/10.1038/s41598-022-19959-y Froger C, Jolivet C, Budzinski H, Pierdet M, Caria G, Saby NPA, Arrouays D, Bispo A (2023). Pesticide residues 1564 1565 in French soils: Occurrence, risks, and persistence. Environ Sci Technol 57:7818-7827. 1566 https://doi.org/10.1021/acs.est.2c09591 1567 Fuentes E, Gaffard A, Rodrigues A, Millet M, Bretagnolle V, Moreau J, Monceau K (2023) Neonicotinoids: Still 1568 present in farmland birds despite their ban. Chemosphere 321:138091. 1569 https://doi.org/10.1016/j.chemosphere.2023.138091 1570 Gamble NE, Huff Hartz KE, Figuero AE, Poynton HC, Lydy MJ (2023) Development of insecticide resistance in 1571 Hyalella azteca. Environ Pollut 322:121165. https://doi.org/10.1016/j.envpol.2023.121165 1572 Gibbons D, Morrissey CA, Mineau P (2015) A review of the direct and indirect effects of neonicotinoids and 1573 fipronil on vertebrate wildlife. Environ Sci Pollut Res 22(1):103-118. http://dx.doi.org/10.1007/s11356-014-1574 3180-5 1575 Gilburn AS, Bunnefeld N, McVean Wilson J, Botham MS, Brereton TM, Fox R, Goulson D (2015) Are 1576 neonicotinoid insecticides driving declines of widespread butterflies? PeerJ 3:e1402. 1577 https://doi.org/10.7717/peerj.1402 1578 Gill RJ, Raine NE (2014) Chronic impairment of bumblebee natural foraging behaviour induced by sublethal 1579 pesticide exposure. Funct Ecol 28(6):1459-1471. http://dx.doi.org/10.1111/1365-2435.12292 N°1. GIP 1580 Loire Bretagne (2013)Micropolluants Les Pesticides. Cahiers Indicateurs 1581 https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&ved=2ahUKEwikgNPv1d3-1582 AhUESaQEHb-QAEgQFnoECBkQAQ&url=https%3A%2F%2Fwww.loireestuaire.org%2Fupload%2Fiedit%2F1%2Fpj%2F51858 2874 L2A6a micropolluants pesticides.pdf&usg 1583 1584 =AOvVaw3x7V9PEzwdEFrnfyUDo GT. Accessed 20 September 2023
- 1587 https://doi.org/10.1002/minf.201800137

data:

predictive

toxicological

1585

1586

Gökçe S, Saçan MT (2019) Assessments of algal toxicity and PBT behaviour of pesticides with no eco-

of

ability

QSA/(T)R

Molec

Inf

models.

38:1800137.

1588	Goulson D (2013) An overview of the environmental risks posed by neonicotinoid insecticides. J Appl Ecol
1589	50:977-987. https://doi.org/ 10.1111/1365-2664.12111
1590	Grassl J, Holt S, Cremen N, Peso M, Hahne D, Baer B (2018) Synergistic effects of pathogen and pesticide
1591	exposure on honey bee (Apis mellifera) survival and immunity. J Invert Pathol 159:78-86.
1592	http://dx.doi.org/10.1016/j.jip.2018.10.005
1593	Graves EE, Meese RJ, Holyoak M (2022) Neonicotinoid exposure in Tricolored Blackbirds (Agelaius tricolor).
1594	Environ Sci Pollut Res 30:15392-15399. https://doi.org/10.1007/s11356-022-23290-4
1595	Gunstone T, Cornelisse T, Klein K, Dubey A, Donley N (2021) Pesticides and soil invertebrates: A hazard
1596	assessment. Front Environ Sci 9:643847. https://doi.org/10.3389/fenvs.2021.643847
1597	Hallmann CA, Foppen RPB, van Turnhout CAM, de Kroon H, Jongejans E (2014) Declines in insectivorous birds
1598	are associated with high neonicotinoid concentrations. Nature 511(7509):341-343+.
1599	http://dx.doi.org/10.1038/nature13531
1600	Hayasaka D, Korenaga T, Suzuki K, Saito F, Sánchez-Bayo F, Goka K (2012) Cumulative ecological impacts of
1601	two successive annual treatments of imidacloprid and fipronil on aquatic communities of paddy mesocosms.
1602	Ecotox Environ Safe 80:355-362. http://dx.doi.org/10.1016/j.ecoenv.2012.04.004
1603	Hayes TB, Falso, P, Gallipeau S, Stice M (2010) The cause of global amphibian declines: a developmental
1604	endocrinologist's perspective. J Exp Biol 213:921-933. http://dx.doi.org/10.1242/jeb.040865
1605	Henry M, Beguin M, Requier F, Rollin O, Odoux JF, Aupinel P, Aptel J, Tchamitchian S, Decourtye A (2012) A
1606	common pesticide decreases foraging success and survival in honey bees. Science 336(6079):348-350.
1607	http://dx.doi.org/10.1126/science.1215039
1608	Henry M, Bertrand C, Le Feon V, Requier F, Odoux JF, Aupinel P, Bretagnolle V, Decourtye A (2014) Pesticide
1609	risk assessment in free-ranging bees is weather and landscape dependent. Nature Comm 5(1):1-8.
1610	http://dx.doi.org/10.1038/ncomms5359
1611	Henry M, Cerrutti N, Aupinel P, Decourtye A, Gayrard M, Odoux JF, Pissard A, Ruger C, Bretagnolle V (2015)
1612	Reconciling laboratory and field assessments of neonicotinoid toxicity to honeybees. Proc Royal Soc B-Biol
1613	Sci 282:20152110. http://dx.doi.org/10.1098/rspb.2015.2110
1614	Henry M, Becher MA, Osborne JL, Kennedy PJ, Aupinel P, Bretagnolle V, Brun F, Grimm V, Horn J, Requier F
1615	(2017) Predictive systems models can help elucidate bee declines driven by multiple combined stressors.
1616	Apidologie 48(3):328-339. http://dx.doi.org/10.1007/s13592-016-0476-0

1617	Hladik ML, Vandever M, Smalling KL (2016) Exposure of native bees foraging in an agricultural landscape to
1618	current-use pesticides. Sci Total Environ 542:469-477. http://dx.doi.org/10.1016/j.scitotenv.2015.10.077
1619	Hoffmann AA, Willi Y (2008) Detecting genetic responses to environmental change. Nature Rev Gen 9(6):421-
1620	432. http://dx.doi.org/10.1038/nrg2339
1621	Hooper SE, Amelon SK, Lin CH (2022) Development of an LC-MS/MS method for non-invasive biomonitoring
1622	of neonicotinoid and systemic herbicide pesticide residues in bat hair. Toxics 10:73.
1623	https://doi.org/10.3390/toxics10020073
1624	Hsiao CJ, Lin CL, Lin TY, Wang SE, Wu CH (2016) Imidacloprid toxicity impairs spatial memory of echolocation
1625	bats through neural apoptosis in hippocampal CA1 and medial entorhinal cortex areas. Neuroreport
1626	27(6):462-468. http://dx.doi.org/10.1097/wnr.000000000000562
1627	Humann-Guilleminot S, Binkowski ŁJ, Jenni L, Hilke G, Glauser G, Helfenstein F (2019a) A nation-wide survey
1628	of neonicotinoid insecticides in agricultural land with implications for agri-environment schemes. J Appl
1629	Ecol 56:1502-1514. https://doi.org/10.1111/1365-2664.13392
1630	Humann-Guilleminot S, Clement S, Desprat J, Binkowski LJ, Glauser G, Helfenstein F (2019b) A large-scale
1631	survey of house sparrows feathers reveals ubiquitous presence of neonicotinoids in farmlands. Sci Total
1632	Environ 660:1091-1097. http://dx.doi.org/10.1016/j.scitotenv.2019.01.068
1633	Humann-Guilleminot S, de Montaigu CT, Sire J, Grunig S, Gning O, Glauser G, Vallat A, Helfenstein F (2019c)
1634	A sublethal dose of the neonicotinoid insecticide acetamiprid reduces sperm density in a songbird. Environ
1635	Res 177:108589. http://dx.doi.org/10.1016/j.envres.2019.108589
1636	Humann-Guilleminot S, Laurent S, Bize P, Roulin A, Glauser G, Helfenstein F (2021) Contamination by
1637	neonicotinoid insecticides in barn owls (Tyto alba) and Alpine swifts (Tachymarptis melba). Sci Total
1638	Environ 785:147403. http://dx.doi.org/10.1016/j.scitotenv.2021.147403
1639	Iturburu FG, Simoniello MF, Medici S, Panzeri AM, Menone ML (2018) Imidacloprid Causes DNA damage in
1640	fish: Clastogenesis as a mechanism of genotoxicity. Bull Environ Cont Toxicol 100:760-764.
1641	https://doi.org/10.1007/s00128-018-2338-0
1642	James DG (2019) A neonicotinoid insecticide at a rate found in nectar reduces longevity but not oogenesis in
1643	monarch butterflies, Danaus plexippus (L.). (Lepidoptera: Nymphalidae). Insects 10:276.
1644	http://dx.doi.org/10.3390/insects10090276
1645	Jones A, Harrington P, Turnbull G (2014) Neonicotinoid concentrations in arable soils after seed treatment
1646	applications in preceding years. Pest Manage Sci 70:1780-1784, https://doi.org/10.1002/ps.3836

1647	JORF (2021) Arrêté du 5 février 2021 autorisant provisoirement l'emploi de semences de betteraves sucrières
1648	traitées avec des produits phytopharmaceutiques contenant les substances actives imidaclopride ou
1649	thiamethoxam. Journal Officiel de la République Française, Texte 25 sur 113, 6 février 2021.
1650	https://www.legifrance.gouv.fr/download/pdf?id=hbs9mp5-
1651	XF68jgbjPxIB6NVeMJTi2C06ic6UTBmB0po=. Accessed 20 September 2023
1652	JORF (2022) Arrêté du 31 janvier 2022 autorisant provisoirement l'emploi de semences de betteraves sucrières
1653	traitées avec des produits phytopharmaceutiques contenant les substances actives imidaclopride ou
1654	thiamethoxam et précisant les cultures qui peuvent être semées, plantées ou replantées au titre des campagnes
1655	suivantes. Journal Officiel de la République Française, Texte 44 sur 145, 1er février 2022.
1656	https://www.legifrance.gouv.fr/download/pdf?id=6WRjAfdMSHV4TefP9-
1657	NdUXcMTd9GOIfKK9zcag5Ik0E=. Accessed 20 September 2023
1658	Kaczynski P, Lozowicka B, Perkowski M, Zon W, Hrynko I, Rutkowska E, Skibko Z (2021) Impact of broad-
1659	spectrum pesticides used in the agricultural and forestry sector on the pesticide profile in wild boar, roe deer
1660	and deer and risk assessment for venison consumers. Sci Total Environ 784:147215.
1661	http://dx.doi.org/10.1016/j.scitotenv.2021.147215
1662	Kattwinkel M, Reichert P, Ruegg J, Liess M, Schuwirth N (2016) Modeling macroinvertebrate community
1663	dynamics in stream mesocosms contaminated with a pesticide. Environ Sci Technol 50(6):3165-3173.
1664	http://dx.doi.org/10.1021/acs.est.5b04068
1665	Kenna D, Cooley H, Pretelli I, Rodrigues AR, Gill SD, Gill RJ (2019) Pesticide exposure affects flight dynamics
1666	and reduces flight endurance in bumblebees. Ecol Evol 9(10):5637-5650.
1667	http://dx.doi.org/10.1002/ece3.5143
1668	Khans FR, Alhewairini SS (2019) Effects of insecticides on natural population of hymenopterous parasitoids in
1669	Alfalfa (Medicago sativa L.) agro-ecosystem. Pakistan J Agric Sci 56(4):1087-1093.
1670	Khidkhan K, Poapolathep S (2022) Residues of neonicotinoids and fipronil in paddy fields and
1671	duck eggs: Do ducks transform and accumulate these substances to egg. J Toxicol Sci P-22E.
1672	https://doi.org/10.14869/toxpt.49.1.0_P-22E
1673	Kimura-Kuroda J, Komuta Y, Kuroda Y, Hayashi M, Kawano H (2012) Nicotine-like effects of the neonicotinoid
1674	insecticides acetamiprid and imidacloprid on cerebellar neurons from neonatal rats. Plos One 7:e32432.
1675	https://doi.org/10.1371/journal.pone.0032432

16/6	Klaus F, Ischarntke I, Bischoff G, Grass I (2021) Floral resource diversification promotes solitary bee
1677	reproduction and may offset insecticide effects - evidence from a semi-field experiment. Ecol Lett 24(4):668-
1678	675. http://dx.doi.org/10.1111/ele.13683
1679	Klingelhöfer D, Braun M, Brüggman D, Groneberg DA (2022) Neonicotinoids: A critical assessment of the global
1680	research landscape of the most extensively used insecticide. Environ Res 213:113727.
1681	https://doi.org/10.1016/j.envres.2022.113727
1682	Kobashi K, Harada T, Adachi Y, Mori M, Ihara M, Hayasaka D (2017) Comparative ecotoxicity of imidacloprid
1683	and dinotefuran to aquatic insects in rice mesocosms. Ecotox Environ Safe 138:122-129.
1684	https://doi.org/10.1016/j.ecoenv.2016.12.025
1685	Könemann S, Meyer S, Betz A, Županič A, vom Berg C (2021) Sub-Lethal Peak Exposure to Insecticides Triggers
1686	Olfaction-Mediated Avoidance in Zebrafish Larvae. Environ Sci Technol 55:11835-11847.
1687	https://doi.org/10.1021/acs.est.1c01792
1688	Koppenhöfer AM, Cowles RS, Cowles EA, Fuzy EM, Kaya HK (2003) Effect of neonicotinoid synergists on
1689	entomopathogenic nematode fitness. Entomol Exp Applic 106(1):7-18. http://dx.doi.org/10.1046/j.1570-
1690	7458.2003.00008.x
1691	Kraus JM, Kuivila KM, Hladik ML, Shook N, Mushet DM, Dowdy K, Harrington R (2021) Cross-ecosystem
1692	fluxes of pesticides from prairie wetlands mediated by aquatic insect emergence: Implications for terrestrial
1693	insectivores. Environ Tox Chem 40(8):2282-2296. http://dx.doi.org/10.1002/etc.5111
1694	Kreutzweiser D, Good K, Chartrand D, Scarr T, Thompson D (2007) Non-target effects on aquatic decomposer
1695	organisms of imidacloprid as a systemic insecticide to control emerald ash borer in riparian trees. Ecotox
1696	Environ Safe 68(3):315-325. http://dx.doi.org/10.1016/j.ecoenv.2007.04.011
1697	Kreutzweiser DP, Good KP, Chartrand DT, Scarr TA, Thompson DG (2008) Toxicity of the systemic insecticide,
1698	imidacloprid, to forest stream insects and microbial communities. Bull Environ Contam Toxicol 80(3):211-
1699	214. http://dx.doi.org/10.1007/s00128-007-9347-8
1700	Krupke CH, Hunt GJ, Eitzer BD, Andino G, Given K (2012) Multiple routes of pesticide exposure for honey bees
1701	living near agricultural fields. Plos One 7(1):e29268. http://dx.doi.org/10.1371/journal.pone.0029268
1702	Krupke CH, Long EY (2015) Intersections between neonicotinoid seed treatments and honey bees. Curr Op Insect
1703	Sci 10:8-13 http://dx.doi.org/10.1016/j.cois.2015.04.005

1704 Kudelska MM, Holden-Dye L, O'Connor V, Doyle DA (2017) Concentration-dependent effects of acute and 1705 chronic neonicotinoid exposure on the behaviour and development of the nematode Caenorhabditis elegans. 1706 Pest Manage Sci 73:1345-1351. http://dx.doi.org/10.1002/ps.4564 1707 Kuechle KJ, Webb EB, Mengel D, Main AR (2019) Factors influencing neonicotinoid insecticide concentrations 1708 in floodplain wetland sediments across Missouri. Environ Sci Technol 53(18):10591-10600. 1709 http://dx.doi.org/10.1021/acs.est.9b01799 1710 Kuzukiran O, Simsek I, Yorulmaz T, Yurdakok-Dikmen B, Ozkan O, Filazi A (2021) Multiresidues of 1711 environmental Turkey. 282:131022. contaminants in bats from Chemosphere 1712 https://doi.org/10.1016/j.chemosphere.2021.131022 1713 LaLone CA, Villeneuve DL, Wu-Smart J, Milsk RY, Sappington K, Garber KV, Housenger J, Ankley GT (2017) 1714 Weight of evidence evaluation of a network of adverse outcome pathways linking activation of the nicotinic 1715 acetylcholine receptor in honey bees to colony death. Sci Total Environ 584:751-775. 1716 http://dx.doi.org/10.1016/j.scitotenv.2017.01.113 Laycock I, Lenthall KM, Barratt AT, Cresswell JE (2012) Effects of imidacloprid, a neonicotinoid pesticide, on 1717 1718 reproduction in worker bumble bees (Bombus terrestris). Ecotoxicology 21(7):1937-1945. 1719 http://dx.doi.org/10.1007/s10646-012-0927-y 1720 Lee-Jenkins SSY, Robinson SA (2018) Effects of neonicotinoids on putative escape behavior of juvenile wood 1721 frogs (Lithobates sylvaticus) chronically exposed as tadpoles. Environ Tox Chem 37(12):3115-3123. 1722 https://doi.org/10.1002/etc.4284 1723 Lennon RJ, Isaac NJB, Shore RF, Peach WJ, Dunn JC, Pereira MG, Arnold KE, Garthwaite D, Brown CD (2019) 1724 Using long-term datasets to assess the impacts of dietary exposure to neonicotinoids on farmland bird 1725 populations in England. Plos One 14(10):1-21. http://dx.doi.org/10.1371/journal.pone.0223093 1726 Lennon RJ, Shore RF, Pereira MG, Peach WJ, Dunn JC, Arnold KE, Brown CD (2020a) High prevalence of the 1727 neonicotinoid clothianidin in liver and plasma samples collected from gamebirds during autumn sowing. Sci 1728 Total Environ 742:140493. http://dx.doi.org/10.1016/j.scitotenv.2020.140493 1729 Lennon RJ, Peach WJ, Dunn JC, Shore RF, Pereira MG, Sleep D, Dodd S, Wheatley CJ, Arnold KE, Brown CD 1730 (2020b) From seeds to plasma: Confirmed exposure of multiple farmland bird species to clothianidin during 1731 sowing of winter cereals. Sci Total Environ 723:138056. http://dx.doi.org/10.1016/j.scitotenv.2020.138056

1732 Levesque B, Cachot J, Boet P, Lepage M, Mazella N, Martin C, Gourves PY, Legeay A (2018) Seasonal variations 1733 of contamination and exoskeletal malformations in the white shrimps Palaemon longirostris in the Gironde 1734 estuary, France. Environ Sci Pollut Res 25(23):22689-22701. https://doi.org/10.1007/s11356-018-2241-6 1735 Li L, Chen X, Zhang D, Pan X (2010) Effects of insecticide acetamiprid on Photosystem II (PSII) activity of 1736 Synechocystis sp. (FACHB-898). Pestic Biochem Physiol 98:300-304. 1737 https://doi.org/10.1016/j.pestbp.2010.06.022 1738 Li MD, Yang Z, Guo H, Dash B (2016) Evolutionary relationship of nicotinic acetylcholine receptor subunits in 1739 both vertebrate and invertebrate species. Neuromethods 117:227-254. https://doi.org/10.1007/978-1-4939-1740 3768-4 12 1741 Li YF, An JJ, Dang ZH, Lv HY, Pan WL, Gao ZL (2018) Treating wheat seeds with neonicotinoid insecticides 1742 does not harm the rhizosphere microbial community. Plos One 13(12):1-12. 1743 http://dx.doi.org/10.1371/journal.pone.0205200 1744 Li YJ, Miao RQ, Khanna M (2020) Neonicotinoids and decline in bird biodiversity in the United States. Nature 1745 Sustain 3:1027-1035. http://dx.doi.org/10.1038/s41893-020-0582-x 1746 Lima-Fernandes E, Bundschuh M, Bakanov N, Englert D, Schulz R, Schafer RB (2019) Effects of a systemic 1747 pesticide along an aquatic tri-trophic food chain. Bull Environ Contam Toxicol 103:507-514. 1748 http://dx.doi.org/10.1007/s00128-019-02696-w 1749 Lin CH, Sponsler DB, Richardson RT, Watters HD, Glinski DA, Henderson WM, Minucci JM, Lee EH, Purucker 1750 ST, Johnson RM (2021) Honey bees and neonicotinoid-treated corn seed: Contamination, exposure, and 1751 effects. Environ Tox Chem 40(4):1212-1221. http://dx.doi.org/10.1002/etc.4957 1752 Liu H, Tang X, Xu X, Dai Y, Zhang X, Yang Y (2021) Potential for phytoremediation of neonicotinoids by nine 1753 wetland plants. Chemosphere 283:131083. https://doi.org/10.1016/j.chemosphere.2021.131083 1754 Lopez-Antia A, Ortiz-Santaliestra ME, Mougeot F, Mateo R (2013) Experimental exposure of red-legged 1755 partridges (Alectoris rufa) to seeds coated with imidacloprid, thiram and difenoconazole. Ecotoxicology 1756 22(1):125-138. http://dx.doi.org/10.1007/s10646-012-1009-x Lopez-Antia A. Ortiz-Santaliestra ME, Mateo R (2014) Experimental approaches to test pesticide-treated seed 1757 1758 avoidance by birds under a simulated diversification of food sources. Sci Total Environ, 496: 179-187. 1759 http://dx.doi.org/10.1016/j.scitotenv.2014.07.031

1760 Lopez-Antia A, Ortiz-Santaliestra ME, Mougeot F, Mateo R (2015) Imidacloprid-treated seed ingestion has lethal 1761 effect on adult partridges and reduces both breeding investment and offspring immunity. Environ Res 136:97-1762 107. http://dx.doi.org/10.1016/j.envres.2014.10.023 1763 Lopez-Antia A, Feliu J, Camarero PR, Ortiz Santaliestra ME, Mateo R (2016) Risk assessment of pesticide seed 1764 treatment for farmland birds using refined field data. J Appl Ecol 53(5):1373-1381. 1765 https://doi.org/10.1111/1365-2664.12668 1766 Lundin O, Rundlöf M, Smith HG, Fries I, Bommarco R (2015) Neonicotinoid insecticides and their impacts on 1767 bees: A systematic review of research approaches and identification of knowledge gaps. Plos One 1768 10(8):e0136928. http://dx.doi.org/10.1371/journal.pone.0136928 1769 MacDonald AM, Jardine CM, Thomas PJ, Nemeth NM (2018) Neonicotinoid detection in wild turkeys (Meleagris 1770 gallopavo silvestris) in Ontario, Canada. Environ Sci Pollut Res 25(16):16254-16260. 1771 http://dx.doi.org/10.1007/s11356-018-2093-0 1772 Main AR, Fehr J, Liber K, Headley JV, Peru KM, Morrissey CA (2017) Reduction of neonicotinoid insecticide 1773 residues in Prairie wetlands by common wetland plants. Sci Total Environ 579:1193-1202. 1774 http://dx.doi.org/10.1016/j.scitotenv.2016.11.102 1775 Main AR, Webb EB, Govne KW, Mengel D (2020) Reduced species richness of native bees in field margins 1776 associated with neonicotinoid concentrations in non-target soils. Agric Ecosys Environ 287:106693. 1777 http://dx.doi.org/10.1016/j.agee.2019.106693 1778 Malagnoux L, Capowiez Y, Rault M (2015) Impact of insecticide exposure on the predation activity of the 1779 Forficula auricularia. Environ Sci Pollut Res 22:14116-14126. European earwig 1780 http://dx.doi.org/10.1007/s11356-015-4520-9 Malev O, Sauerborn Klobučar R, Fabbretti E, Trebse P (2012) Comparative toxicity of imidacloprid and its 1781 1782 transformation product 6-chloronicotinic acid to non-target organisms: microalgae Desmodesmus 1783 subspicatus and amphipod Gammarus fossarum. Pestic Biochem Physiol 104:178-186. 1784 http://dx.doi.org/10.1016/j.pestbp.2012.07.008 1785 Mansoor MM, Shad SA (2020) Genetics, cross-resistance and realized heritability of resistance to acetamiprid in 1786 generalist predator, Chrysoperla carnea (Steph.) (Neuroptera: Chrysopidae). Egyptian J Biol Pest Cont 30:23.

http://dx.doi.org/10.1186/s41938-020-0213-x

1/88	Mouria R (2016) The use of ducks in rice fields in the control of weeds and pests. The Permaculture Research
1789	Institute. https://www.permaculturenews.org/2016/09/15/use-ducks-rice-fields-control-weed-pest/ Accessed
1790	20 September 2023
1791	McGee S, Whitfield-Aslund M, Duca D, Kopysh N, Dan T, Knopper L, Brewer L (2018) Field evaluation of the
1792	potential for avian exposure to clothianidin following the planting of clothianidin-treated corn seed. Peerj
1793	6:e5880. http://dx.doi.org/10.7717/peerj.5880
1794	Menet-Nedelec F, Gonzalez JL, Halm-Lemeille MP, Repecaud M, Facq JV, Maheux F, Pierre-Duplessix O, Simon
1795	B (2018) Etude d'outils d'évaluation de la contamination chimique dans les eaux de la Manche – ECUME
1796	Rapport Ifremer ODE/UL/LERN/18-02, 53. http://archimer.ifremer.fr/doc/00434/54582/55966.pdf
1797	Miles JC, Hua J, Sepulveda MS, Krupke CH, Hoverman JT (2017) Effects of clothianidin on aquatic communities:
1798	Evaluating the impacts of lethal and sublethal exposure to neonicotinoids. PLoS One 12:1-24.
1799	https://doi.org/10.1371/journal.pone.0174171
1800	Miller JL, Schmidt TS, van Metre PC, Mahler BJ, Sandstrom MW, Nowell LH, Carlisle DM, Moran PW (2020)
1801	Common insecticide disrupts aquatic communities: A mesocosm-to-field ecological risk assessment of
1802	fipronil and its degradates in U.S. streams. Science Adv 6:1-12. https://doi.org/10.1126/sciadv.abc1299
1803	Millot F, Decors A, Mastain O, Quintaine T, Berny P, Vey D, Lasseur R, Bro E (2017) Field evidence of bird
1804	poisonings by imidacloprid-treated seeds: a review of incidents reported by the French SAGIR network from
1805	1995 to 2014. Environ Sci Pollut Res 24(6):5469-5485. http://dx.doi.org/10.1007/s11356-016-8272-y
1806	Mineau P (2017) Organophosphorous and carbamate insecticides: Impacts on birds. Encycl Anthropo 111-117.
1807	http://dx.doi.org/10.1016/B978-0-12-409548-9.09887-0
1808	Mineau P, Callaghan C (2018) Neonicotinoid insecticides and bats: an assessment of the direct and indirect risks.
1809	Ontario, Canada: Canadian Wildlife Federation, 83 p.
1810	Mineau P, Kern H (2023). Neonicotinoid insecticides: Failing to come to grips with a predictable environmental
1811	disaster. American Bird Conservancy, June 2023, 121 p.
1812	Mineau P, Palmer C (2013) The impact of the nation's most widely used insecticides on birds. American Bird
1813	Conservancy, USA, 2013: American Bird Conservancy, Neonicotinoid Insecticides and Birds, 96 p.
1814	Mitchell EAD, Mulhauser B, Mulot M, Mutabazi A, Glauser G, Aebi A (2017) A worldwide survey of
1815	neonicotinoids in honey. Science 358(6359):109-111. https://doi.org/10.1126/science.aan3684

1816 Mohr S, Berghahn R, Schmiediche R, Hübner V, Loth S, Feibicke M, Mailahn W, Wogram J (2012) 1817 Macroinvertebrate community response to repeated short-term pulses of the insecticide imidacloprid. Aquat 1818 Toxicol 110:25-36. https://doi.org/10.1016/j.aquatox.2011.11.016 1819 Monchanin C, Henry M, Decourtye A, Dalmon A, Fortini D, Boeuf E, Dubuisson L, Aupinel P, Chevallereau C, 1820 Petit J, Fourrier J (2019) Hazard of a neonicotinoid insecticide on the homing flight of the honeybee depends 1821 climatic conditions infestation. Chemosphere 224:360-368. on and Varroa 1822 http://dx.doi.org/10.1016/j.chemosphere.2019.02.129 1823 Moreau J, Rabdeau J, Badenhausser I, Giraudeau M, Sepp T, Crépin M, Gaffard A, Bretagnolle V, Monceau K 1824 (2022) Pesticide impacts on avian species with special reference to farmland birds; a review. Environ Monit 1825 Assess 194:790. https://doi.org/10.1007/s10661-022-10394-0 1826 Morrissey CA, Mineau P, Devries JH, Sánchez-Bayo F, Liess M, Cavallaro MC, Liber K (2015) Neonicotinoid 1827 contamination of global surface waters and associated risk to aquatic invertebrates: A review. Environ Int 1828 74:291-303. https://doi.org/10.1016/j.envint.2014.10.024 1829 Müller C (2018) Impacts of sublethal insecticide exposure on insects - Facts and knowledge gaps. Basic App Ecol 1830 30:1-10. http://dx.doi.org/10.1016/j.baae.2018.05.001 1831 Munaron D, Derolez V, Foucault E, Cimiterra N, Tapie N, Budzinski H, Giraud A (2020) OBSLAG - Volet 1832 Pesticides. Bilan 2017-2019 du suivi des lagunes méditerranéennes. Rapport Ifremer n° ODE/UL/LER-1833 LR/20.09. https://archimer.ifremer.fr/doc/00656/76769/ Accessed 20 September 2023 1834 Munaron D, Gianaroli C, Cimiterra N, Derolez V, Ouisse V, Giraud A (2022) OBSLAG - Pesticides, Bilan 2020-1835 2021 du suivi des lagunes méditerranéennes. Rapport de la Convention AERMC-Ifremer 2020, n° 1836 ODE/UL/LER-LR/22.12, août 2022. 90p. https://archimer.ifremer.fr/doc/00795/90672/. Accessed 20 1837 September 2023 1838 Munaron D, Mérigot B, Derolez V, Tapie N, Budzinski H, Fiandrino A (2023) Evaluating pesticide mixture risks 1839 in French Mediterranean coastal lagoons waters. Sci Total Environ 867:161303. 1840 https://doi.org/10.1016/j.scitotenv.2022.161303 Nafaji G, Razi M, Hoshyar A, Shahmohamadloo S, Feyzi S (2010) The effect of chronic exposure with 1841 1842 imidacloprid insecticide on fertility in mature male rats. Int J Fertil Steril 4:9-16. 1843 Naranjo SE (2001) Conservation and evaluation of natural enemies in IPM systems for Bemisia tabaci. Crop Protec 1844 20(9):835-852. http://dx.doi.org/10.1016/s0261-2194(01)00115-6

1845 Naumann T, Bento CPM, Wittmann A, Gandrass J, Tang J, Zhen X, Liu L, Ebinghaus R (2022) Occurrence and 1846 ecological risk assessment of neonicotinoids and related insecticides in the Bohai Sea and its surrounding 1847 rivers, China. Water Res 209:117912. https://doi.org/10.1016/j.watres.2021.117912 1848 Nélieu S, Lamy I, Karolak S, Delarue G, Crouzet O, Barraud C, Bimbot M, Allaoui F, Hanot C, Delorme A, Levi 1849 Y, Hulot FD, Baudry E (2021) Impact of peri-urban landscape on the organic and mineral contamination of 1850 waters and related risk assessment. Environ Sci Pollut Res 28:59256-59267. 1851 https://doi.org/10.1007/s11356-020-10355-5 1852 Neury-Ormanni J, Doose C, Majdi N, Vedrenne J, Morin S, Höss S, Traunspurger W (2019) Tolerance of free-1853 nematode species to imidacloprid and diuron. Invertebr Biol 138:e12272. 1854 https://doi.org/10.1111/ivb.12272 1855 Neury-Ormanni J, Vedrenne J, Morin S (2020a) Benthic diatom growth kinetics under combined pressures of microalgal competition, predation and chemical stressors. Sci Total Environ 734:139484. 1856 1857 https://doi.org/10.1016/j.scitotenv.2020.139484 1858 Neury-Ormanni J, Doose C, Majdi N, Vedrenne J, Traunspurger W, Morin S (2020b) Selective grazing behaviour 1859 of chironomids on microalgae under pesticide pressure. Sci Total Environ 730:138673. 1860 http://dx.doi.org/10.1016/j.scitotenv.2020.138673 1861 Oliveira JM, Destro ALF, Freitas MB, Oliveira LL (2021) How do pesticides affect bats? - A brief review of recent 1862 publications. Braz J Biol 81(2):499-507. http://dx.doi.org/10.1590/1519-6984.225330 1863 Oziolor EM, De Schamphelaere K, Matson CW (2016) Evolutionary toxicology: Meta-analysis of evolutionary 1864 Ecotoxicology 25(10):1858-1866. events in response to chemical stressors. 1865 http://dx.doi.org/10.1007/s10646-016-1735-6 1866 Pearsons KA, Tooker JF (2021) Preventive insecticide use affects arthropod decomposers and decomposition in 1867 field crops. App Soil Ecol 157:103757. http://dx.doi.org/10.1016/j.apsoil.2020.103757 1868 Pekar S (2012) Spiders (Araneae) in the pesticide world: an ecotoxicological review. Pest Manage Sci 1869 68(11):1438-1446. http://dx.doi.org/10.1002/ps.3397 1870 Pekar S, Kocourek F (2004) Spiders (Araneae) in the biological and integrated pest management of apple in the 1871 Czech Republic, J App Entomol 128(8):561-566. http://dx.doi.org/10.1111/j.1439-0418.2004.00884.x 1872 Pelosi C, Bertrand C, Daniele G, Coeurdassier M, Benoit P, Nelieu S, Lafay F, Bretagnolle V, Gaba S, Vulliet E, 1873 Fritsch C (2021) Residues of currently used pesticides in soils and earthworms: A silent threat? Agric Ecosyst

Environ 305:107167. http://dx.doi.org/10.1016/j.agee.2020.107167

18/5	Penn HJ, Dale AM (2017) Imidacioprid seed treatments affect individual ant benavior and community structure
1876	but not egg predation, pest abundance or soybean yield. Pest Manage Sci 73(8):1625-1632.
1877	http://dx.doi.org/10.1002/ps.4499
1878	Pepin JF, Benabdelmouna A, Degremont L, Guesdon S, Le Moine O, Morga B, Bierne N, Travers MA, Robert S,
1879	Soletchnik P (2017) Mortalités de moules bleues dans les secteurs mytilicoles charentais et vendéens :
1880	description et facteurs liés – MORBLEU. Rapport Ifremer RBE/SG2M-LGPMM, 93p.
1881	Pesce S, Mamy L, Achard AL, Le Gall M, Le Perchec S, Réchauchère O, Tibi A, Leenhardt S, Sanchez W (2021)
1882	Collective scientific assessment as a relevant tool to inform public debate and policymaking: an illustration
1883	about the effects of plant protection products on biodiversity and ecosystem services. Environ Sci Pollut Res
1884	28:38448-38454. https://doi.org/10.1007/s11356-021-14863-w
1885	Pesce S, Mamy L, Sanchez W, Amichot M, Artigas J, Aviron S, Barthélémy C, Beaudouin R, Bedos C, Bérard A,
1886	Berny P, Bertrand C, Bertrand C, Betoulle S, Bureau-Point E, Charles S, Chaumot A, Chauvel B,
1887	Coeurdassier M, Corio-Costet MF, Coutellec MA, Crouzet O, Doussan I, Faburé J, Fritsch C, Gallai N,
1888	Gonzalez P, Gouy V, Hedde M, Langlais A, Le Bellec F, Leboulanger C, Margoum C, Martin-Laurent F,
1889	Mongruel R, Morin S, Mougin C, Munaron D, Nélieu S, Pelosi C, Rault M, Sabater S, Stachowski-Haberkorn
1890	S, Sucré E, Thomas M, Tournebize J, Leenhardt S (2023) Main conclusions and perspectives from the
1891	collective scientific assessment on the effects of plant protection products on biodiversity and ecosystem
1892	services along the land-sea continuum in France and French overseas territories. Environ Sci Pollut Res (in
1893	press). https://doi.org/10.1007/s11356-023-26952-z
1894	Pettis JS, Lichtenberg EM, Andree M, Stitzinger J, Rose R, Vanengelsdorp D (2013) Crop pollination exposes
1895	honey bees to pesticides which alters their susceptibility to the gut pathogen Nosema ceranae. Plos One
1896	8(7):9. http://dx.doi.org/10.1371/journal.pone.0070182
1897	Phytatmo database (2023) https://www.atmo-france.org/article/phytatmo. Accessed 20 September 2023
1898	Picone M, Distefano GG, Zangrando R, Gambaro A, Volpi Ghirardini A (2023). Neonicotinoids and
1899	pharmaceuticals in hair of the Red fox (Vulpes vulpes) from the Cavallino-Treporti peninsula, Italy. Environ
1900	Res 228:115837. https://doi.org/10.1016/j.envres.2023.115837
1901	Pietrzak D, Kania J, Malina G, Kmiecik E, Wator K (2019) Pesticides from the EU first and second watch lists in
1902	the water environment. Clean-Soil Air Water 47(7):1800376. https://doi.org/10.1002/clen.201800376

1903	Pilling E, Campbell P, Coulson M, Ruddle N, Tornier I (2013) A four-year field program investigating long-rerm
1904	effects of repeated exposure of honey bee colonies to flowering crops treated with thiamethoxam. Plos One
1905	8(10):14. https://doi.org/10.1371/journal.pone.0077193
1906	Pisa LW, Amaral-Rogers V, Belzunces LP, Bonmatin JM, Downs CA, Goulson D, Kreutzweiser DP, Krupke C,
1907	Liess M, McField M, Morrissey CA, Noome DA, Settele J, Simon-Delso N, Stark JD, Van der Sluijs JP, Van
1908	Dyck H, Wiemers M (2015) Effects of neonicotinoids and fipronil on non-target invertebrates. Environ Sci
1909	Pollut Res 22(1):68-102. http://dx.doi.org/10.1007/s11356-014-3471-x
1910	Poisson MC, Garrett DR, Sigouin A, Belisle M, Garant D, Haroune L, Bellenger JP, Pelletier F (2021) Assessing
1911	pesticides exposure effects on the reproductive performance of a declining aerial insectivore. Ecol Applic
1912	31(7):e02415. http://dx.doi.org/10.1002/eap.2415
1913	Pozzebon A, Duso C, Tirello P, Ortiz PB (2011) Toxicity of thiamethoxam to Tetranychus urticae Koch and
1914	Phytoseiulus persimilis Athias-Henriot (Acari Tetranychidae, Phytoseiidae) through different routes of
1915	exposure. Pest Manage Sci 67:352-359. https://doi.org/10.1002/ps.2072
1916	PPDB (2023) Pesticide Properties DataBase. http://sitem.herts.ac.uk/aeru/ppdb/. Accessed 20 September 2023
1917	Preuss TG, Agatz A, Goussen B, Roeben V, Rumkee J, Zakharova L, Thorbek P (2022) The BEEHAVE_ecotox
1918	model - Integrating a mechanistic effect module into the honeybee colony model. Environ Toxicol Chem
1919	41:2870-2882. https://doi.org/10.1002/etc.5467
1920	Prosser P, Hart ADM (2005) Assessing potential exposure of birds to pesticide-treated seed. Ecotoxicology
1921	14:679-691. https://doi.org/10.1007/s10646-005-0018-4
1922	Prouteau L (2021) Caractérisation de la contamination en pesticides azoles et néonicotinoïdes chez les espèces
1923	d'intérêt localisées en région Nouvelle-Aquitaine : développement de méthodes analytiques et applications
1924	(PhD Thesis). La Rochelle Université, La Rochelle, France.
1925	Puglis HJ, Boone MD (2011) Effects of technical-grade active ingredient vs. commercial formulation of seven
1926	pesticides in the presence or absence of UV radiation on survival of green frog tadpoles. Arch Environ
1927	Contam Toxicol 60:145-155. https://doi.org/10.1007/s00244-010-9528-z
1928	Qi S, Wang D, Zhu L, Teng M, Wang C, Xue X, Wu L (2018) Effects of a novel neonicotinoid insecticide
1929	cycloxaprid on earthworm, Eisenia fetida. Environ Sci Pollut Res 25:14138-14147.
1020	https://doi.org/10.1007/s11356.018.1624.7

1931	Raina-Fulton R (2015) Determination of neonicotinoid insecticides and strobilurin fungicides in particle phase
1932	atmospheric samples by liquid chromatography-tandem mass spectrometry. J Agric Food Chem 63(21):5152-
1933	5162. https://doi.org/10.1021/acs.jafc.5b01347
1934	Resende-Silva GA, Turchen LM, Guedes RNC, Cutler GC (2019) Imidacloprid soil drenches affect weight and
1935	functional response of spined soldier bug (Hemiptera: Pentatomidae). J Eco Entomol 112(2):558-564.
1936	http://dx.doi.org/10.1093/jee/toy401
1937	Rico A, Arenas-Sánchez A, Pasqualini J, García-Astillero A, Cherta L, Nozal L, Vighi M (2018) Effects of
1938	imidacloprid and a neonicotinoid mixture on aquatic invertebrate communities under Mediterranean
1939	conditions. Aquat Toxicol 204:130-143. http://dx.doi.org/10.1016/j.aquatox.2018.09.004
1940	Riedo J, Wettestein FE, Rösch A, Herzog C, Banerjee S, Büchi L, Charles R, Wächter D, Martin-Laurent F,
1941	Bucheli TD, Walder T, van der Heijden MGA (2021) Widespread occurrence of pesticides in organically
1942	managed agricultural soils - the ghost of a conventional agricultural past? Environ Sci Technol 55:2919-
1943	2928. https://dx.doi.org/10.1021/acs.est.0c06405
1944	Rios FM, Wilcoxen TE, Zimmerman LM (2017) Effects of imidacloprid on Rana catesbeiana immune and nervous
1945	system. Chemosphere 188:465-469. https://doi.org/10.1016/j.chemosphere.2017.08.155
1946	Rogers MA, Krischik VA, Martin LA (2007) Effect of soil application of imidacloprid on survival of adult green
1947	lacewing, Chrysoperla carnea (Neuroptera : Chrysopidae), used for biological control in greenhouse. Biol
1948	Control 42(2):172-177. http://dx.doi.org/10.1016/j.biocontrol.2007.05.006
1949	Rogers KH, McMillin S, Olstad KJ, Poppenga RH (2019) Imidacloprid poisoning of songbirds following a drench
1950	application of trees in a residential neighborhood in California, USA. Environ Tox Chem 38(8):1724-1727.
1951	http://dx.doi.org/10.1002/etc.4473
1952	Rolke D, Fuchs S, Grunewald B, Gao ZL, Blenau W (2016) Large-scale monitoring of effects of clothianidin-
1953	dressed oilseed rape seeds on pollinating insects in Northern Germany: effects on honey bees (Apis
1954	mellifera). Ecotoxicology 25(9):1648-1665. http://dx.doi.org/10.1007/s10646-016-1725-8
1955	Rosas-Ramos N, Banos-Picon L, Tormos J, Asis JD (2020) Natural enemies and pollinators in traditional cherry
1956	orchards: Functionally important taxa respond differently to farming system. Agric Ecosyst Environ
1957	295:106920. http://dx.doi.org/10.1016/j.agee.2020.106920
1958	Rouchaud J, Gustin F, Wauters A (1994) Soil biodegradation and leaf transfer of insecticide imidacloprid applied
1959	in seed dressing in sugar beet crops. Bull Environ Contam Toxicol 53(3):344-350.
1960	https://doi.org/10.1007/BF00197224

1961	Roy CL, Coy PL, Chen D, Ponder J, Jankowski M (2019) Multi-scale availability of neonicotinoid-treated seed
1962	for wildlife in an agricultural landscape during spring planting. Sci Total Environ 682:271-281.
1963	https://doi.org/10.1016/j.scitotenv.2019.05.010
1964	Roy CL, Jankowski MD, Ponder J, Chen D (2020) Sublethal and lethal methods to detect recent imidacloprid
1965	exposure in birds with application to field studies. Environ Tox Chem 39(7):1355-1366.
1966	http://dx.doi.org/10.1002/etc.4721
1967	Roy CL, Chen D (2023) High population prevalence of neonicotinoids in sharp-tailed grouse and greater prairie-
1968	chickens across an agricultural gradient during spring and fall. Sci Total Environ 856:159120.
1969	https://doi.org/10.1016/j.scitotenv.2022.159120
1970	Rundlöf M, Andersson GKS, Bommarco R, Fries I, Hederstrom V, Herbertsson L, Jonsson O, Klatt BK, Pedersen
1971	TR, Yourstone J, Smith HG (2015) Seed coating with a neonicotinoid insecticide negatively affects wild
1972	bees. Nature 521(7550):77-U162. http://dx.doi.org/10.1038/nature14420
1973	Sabin LB, Mora MA (2022) Ecological risk assessment of the effects of neonicotinoid insecticides on northern
1974	bobwhites (Colinus virginianus) in the South Texas Plains Ecoregion. Integr Environ Assess Manage 18:488-
1975	499. https://doi.org/10.1002/ieam.4479
1976	Saito T, Doi M, Katayama H, Kaneko S, Tagami Y, Sugiyama K (2008) Seasonal abundance of hymenopteran
1977	parasitoids of the leafminer Chromatomyia horticola (Diptera: Agromyzidae) and the impact of insecticide
1978	applications on parasitoids in garden pea field. Appl Entomol Zool 43(4):617-624.
1979	http://dx.doi.org/10.1303/aez.2008.617
1980	Samson-Robert O, Labrie G, Chagnon M, Fournier V (2017) Planting of neonicotinoid-coated corn raises honey
1981	bee mortality and sets back colony development. Peerj 5:e3670. http://dx.doi.org/10.7717/peerj.3670
1982	Samojeden CG, Pavan FA, Rutkoski CF, Folador A, da Fré SP, Müller C, Hartmann PA, Hartman M (2022)
1983	Toxicity and genotoxicity of imidacloprid in the tadpoles of Leptodactylus luctator and Physalaemus cuvieri
1984	(Anura: Leptodactylidae). Scient Rep 12:11926. https://doi.org/10.1038/s41598-022-16039-z
1985	Sánchez-Bayo F, Goka K (2005) Unexpected effects of zinc pyrithione and imidacloprid on Japanese medaka fish
1986	(Oryzias latipes). Aquatic Toxicol 74:285-293. https://doi.org/10.1016/j.aquatox.2005.06.003
1987	Sánchez-Bayo F, Goka K, Hayasaka D (2016) Contamination of the aquatic environment with neonicotinoids and
1988	its implication for ecosystems. Front Environ Sci 4:71. http://dx.doi.org/10.3389/fenvs.2016.00071

1989 Sandrock C, Tanadini LG, Pettis JS, Biesmeijer JC, Potts SG, Neumann P (2014) Sublethal neonicotinoid 1990 insecticide exposure reduces solitary bee reproductive success. Agric For Entomol 16:119-128. 1991 https://doi.org/10.1111/afe.12041 1992 Sappington JD (2018) Imidacloprid alters ant sociobehavioral traits at environmentally relevant concentrations. 1993 Ecotoxicology 27:1179-1187. https://doi.org/10.1007/s10646-018-1976-7 1994 Sarnaik SS, Kanekar PP, Raut VM, Taware SP, Chavan KS, Bhadbhade BJ (2006) Effect of application of different 1995 pesticides to soybean on the soil microflora. J Environ Biol 27(2):423-426. 1996 Schaafsma AW, Limay-Rios V, Baute TS, Smith JL (2019) Neonicotinoid insecticide residues in subsurface 1997 drainage and open ditch water around maize fields in southwestern Ontario. Plose One 14(4): e021478. 1998 https://doi.org/10.1371/journal.pone.0214787 1999 Schepker TJ, Webb EB, Tillitt D, LaGrange T (2020) Neonicotinoid insecticide concentrations in agricultural 2000 wetlands and associations with aquatic invertebrate communities. Agric Ecosyst Environ 287:106678. 2001 http://dx.doi.org/10.1016/j.agee.2019.106678 2002 Schmidt TS, Miller JL, Mahler BJ, Van Metre PC, Nowell LH, Sandstrom MW, Carlisle DM, Moran PW, Bradley 2003 PM (2022) Ecological consequences of neonicotinoid mixtures in streams. Science Adv 8:1-12. 2004 https://doi.org/10.1126/sciadv.abj8182 2005 Schott M, Sandmann M, Cresswell JE, Becher MA, Eichner G, Brandt DT, Halitschke R, Krueger S, Morlock G, 2006 During RA, Vilcinskas A, Meixner MD, Buchler R, Brandt A (2021) Honeybee colonies compensate for 2007 pesticide-induced effects on royal jelly composition and brood survival with increased brood production. 2008 Scient Rep 11:62. http://dx.doi.org/10.1038/s41598-020-79660-w 2009 Sgolastra F, Medrzycki P, Bortolotti L, Renzi MT, Tosi S, Bogo G, Teper D, Porrini C, Molowny-Horas R, Bosch 2010 J (2017) Synergistic mortality between a neonicotinoid insecticide and an ergosterol-biosynthesis-inhibiting 2011 fungicide in three bee species. Pest Manage Sci 73(6):1236-1243. http://dx.doi.org/10.1002/ps.4449 Shahid N, Becker JM, Krauss M, Brack W, Liess M (2018a) Adaptation of Gammarus pulex to agricultural 2012 2013 insecticide contamination Sci Total Environ in streams. 621:479-485. 2014 http://dx.doi.org/10.1016/j.scitotenv.2017.11.220 2015 Shahid N, Becker JM, Krauss M, Brack W, Liess M (2018b) Pesticide body burden of the grustacean Gammarus 2016 pulex as a measure of toxic pressure in agricultural streams. Environ Sci Technol 52(14):7823-7832.

2017

http://dx.doi.org/10.1021/acs.est.8b01751

2018 Shaw PW, Wallis DR (2010) Susceptibility of the European earwig, Forficula auricularia, to insecticide residues 2019 on apple leaves. N Z Plant Prot 63:55-59. 2020 Shinya S, Sashika M, Minamikawa M, Itoh T, Yohannes YB, Nakayama SMM, Ishizuka M, Nimako C, Ikenaka 2021 Y (2022) Estimation of the effects of neonicotinoid insecticides on wild raccoon, procyon lotor, in Hokkaido, 2022 Japan: Urinary concentrations and hepatic metabolic capability of neonicotinoids. Environ Toxicol Chem 2023 41:1865-1874. https://doi.org/10.1002/etc.5349 2024 Shukla S, Jhamtani RC, Dahiya MS, Agarwal R (2017) Oxidative injury caused by individual and combined 2025 exposure of neonicotinoid, organophosphate and herbicide in zebrafish. Toxicol Rep 4:240-244. 2026 https://doi.org/10.1016/j.toxrep.2017.05.002 2027 Sievers M, Hale R, Swearer SE, Parris KM (2018) Contaminant mixtures interact to impair predator-avoidance 2028 behaviours and survival in a larval amphibian. **Ecotox** Environ Safe 161:482-488. 2029 https://doi.org/10.1016/j.ecoenv.2018.06.028 2030 Silva V, Mol HGJ, Zomer P, Tienstra M, Ritsema CJ, Geissen V (2019) Pesticide residues in European agricultural 2031 soils hidden reality unfolded. Sci Total Environ 653:1532-1545. 2032 https://doi.org/10.1016/j.scitotenv.2018.10.441. 2033 Simon-Delso N, Amaral-Rogers V, Belzunces LP, Bonmatin JM, Chagnon M, Downs C, Furlan L, Gibbons DW, 2034 Giorio C, Girolami V, Goulson D, Kreutzwiser DP, Krupke CH, Liess M, Long E, McField M, Mineau P, 2035 Mitchell EAD, Morrissey CA, Noome DA, Pisa L, Settele J, Stark JD, Tapparo A, Van Dyck H, Van Praagh 2036 J, Van der Sluijs JP, Whitehorn PR, Wiemers M (2015) Systemic insecticides (neonicotinoids and fipronil): 2037 metabolites. Environ Sci Pollut 22:5-34. trends. uses. mode of action and Res 2038 http://dx.doi.org/10.1007/s11356-014-3470-y 2039 Sol Dourdin T, Rivière G, Cormier A, Di Poi C, Guyomard K, Rabiller M, Akcha F, Bah Sadialiou T, Le Monier 2040 P, Sussarellu R (2023) Molecular and phenotypic effects of early exposure to an environmentally relevant 2041 pesticide mixture in the Pacific oyster, Crassostrea gigas. Environ Pollut 326:121472. 2042 https://doi.org/10.1016/j.envpol.2023.121472 2043 Stahlschmidt P, Brühl CA (2012) Bats at risk? Bat activity and insecticide residue analysis of food items in an 2044 apple orchard. Environ Tox Chem 31(7):1556-1563. http://dx.doi.org/10.1002/etc.1834 2045 Stahlschmidt P, Hahn M, Brühl CA (2017) Nocturnal risks-high bat activity in the agricultural landscape indicates

potential pesticide exposure. Front Environ Sci 5:62. http://dx.doi.org/10.3389/fenvs.2017.00062

2047 Stehle S, Bub S, Schulz R (2018) Compilation and analysis of global surface water concentrations for individual 2048 insecticide compounds. Sci Total Environ 639:516-525. https://doi.org/10.1016/j.scitotenv.2018.05.158 2049 Strobl V, Albrecht M, Villamar-Bouza L, Tosi S, Neumann P, Straub L (2021a) The neonicotinoid thiamethoxam 2050 impairs fertility in solitary bees, Osmia cornuta. Environ Pollut 284:117106. 2051 http://dx.doi.org/10.1016/j.envpol.2021.117106 2052 Strobl V, Bruckner S, Radford S, Wolf S, Albrecht M, Villamar-Bouza L, Maitip J, Kolari E, Chantawannakul P, 2053 Glauser G, Williams GR, Neumann P, Straub L (2021b) No impact of neonicotinoids on male solitary bees 2054 Osmia semi-field conditions. Physiological Entomology cornuta under 46(1):105-109. 2055 http://dx.doi.org/10.1111/phen.12349 2056 Stuligross C, Williams NM (2020) Pesticide and resource stressors additively impair wild bee reproduction. Proc 2057 Royal Soc B 287:20201390. https://doi.org/10.1098/rspb.2020.1390 2058 Stuligross C, Williams NM (2021) Past insecticide exposure reduces bee reproduction and population growth rate. 2059 Proc Natio Acad Sci USA 118:e2109909118. https://doi.org/10.1073/pnas.2109909118 2060 Sumon KA, Ritika AK, Peeters ET, Rashid H, Bosma RH, Rahman MS, Fatema MK, Van den Brink PJ (2018) 2061 Effects of imidacloprid on the ecology of sub-tropical freshwater microcosms. Environ Pollut 236:432-441. 2062 http://dx.doi.org/10.1016/j.envpol.2018.01.102 2063 Sur R, Stork A (2003). Uptake, translocation and metabolism of imidacloprid in plants. Bull Insectol 56:35-40. 2064 Suryanarayanan S (2013) Balancing control and complexity in field studies of neonicotinoids and honey bee 2065 health. Insects 4(1):153-167. https://www.mdpi.com/2075-4450/4/1/153 2066 Švara V, Krauss M, Michalski SG, Altenburger R, Brack W, Luckenbach T (2021) Chemical pollution levels in a 2067 river explain site-specific sensitivities to micropollutants within a genetically homogeneous population of 2068 freshwater amphipods. Environ Sci Technol 55(9):6087-6096. http://dx.doi.org/10.1021/acs.est.0c07839 2069 Sweeney MR, Thompson CM, Popescu VD (2021) Sublethal, behavioral, and developmental effects of the 2070 neonicotinoid pesticide imidacloprid on larval wood frogs (Rana sylvatica). Environ Tox Chem 40(7):1840-2071 1849. https://doi.org/10.1002/etc.5047 2072 Szczepaniec A, Creary SF, Laskowski KL, Nyrop JP, Raupp MJ (2011) Neonicotinoid insecticide imidacloprid 2073 causes outbreaks of spider mites on elm trees in urban landscapes. Plos One 6:e20018. 2074 https://doi.org/10.1371/journal.pone.0020018 2075 Taliansky-Chamudis A, Gomez-Ramirez P, Leon-Ortega M, Garcia-Fernandez AJ (2017) Validation of a 2076 QuECheRS method for analysis of neonicotinoids in small volumes of blood and assessment of exposure in

2077 Eurasian Sci Total 595:93-100. eagle owl (Bubo bubo) nestlings. Environ 2078 http://dx.doi.org/10.1016/j.scitotenv.2017.03.246 2079 Tapie N, Budzinski H (2018) Quantification de la présence dans les eaux bilan de 2010 à 2016 Rapport REPAR 2080 (Reseau Pesticides du Bassin d'Arcachon), 25. 2081 Tapparo A, Giorio C, Marzaro M, Marton D, Solda L, Girolami V (2011) Rapid analysis of neonicotinoid 2082 insecticides in guttation drops of corn seedlings obtained from coated seeds. J Environ Monitor 13(6):1564-2083 1568. https://doi.org/10.1039/c1em10085h 2084 Tennekes H, Zillweger AB (2010) The systemic insecticides: a disaster in the making. Swiss Society of 2085 Toxicology, Annual Meeting, 22 November 2012. ETS Nederland BV Zutphen, 57 p. 2086 https://www.boerenlandvogels.nl/sites/default/files/Tennekes_Presentation_Annual%20Meeting_Swiss%20 2087 Toxicology%20Society %2022112012.pdf. Accessed 20 September 2023 2088 Tetsatsi ACM, Nkeng-Effouet PA, Alumeti DM, Bonsou GRF, Kamanyi A, Watcho P (2019) Colibri (R) 2089 insecticide induces male reproductive toxicity: alleviating effects of Lannea acida (Anacardiaceae) in rats. 2090 Basic Clini Androl 29:16. http://dx.doi.org/10.1186/s12610-019-0096-4 2091 Thiel S, Kohler HR (2016) A sublethal imidacloprid concentration alters foraging and competition behaviour of 2092 ants. Ecotoxicology 25:814-823. https://doi.org/10.1007/s10646-016-1638-6 2093 Thompson DA, Lehmler HJ, Kolpin DW, Hladik ML, Vargo JD, Schilling KE, LeFevre GH, Peeples TL, Poch 2094 MC, LaDuca LE, Cwiertny DM, Field RW (2020) A critical review on the potential impacts of neonicotinoid 2095 insecticide use: current knowledge of environmental fate, toxicity, and implications for human health. 2096 Environ Sci: Processes Impacts 22:1315-1346. https://doi.org/10.1039/C9EM00586B 2097 Thompson CM, Sweeney MR, Popescu VD (2022) Carryover effects of pesticide exposure and pond drying on 2098 performance, behavior, and sex ratios in a pool breeding amphibian. J Zool 317:229-240. 2099 https://doi.org/10.1111/jzo.12975 2100 Tison L, Roessner A, Gerschewski S, Menzel R (2019) The neonicotinoid clothianidin impairs memory processing 2101 in honey bees. Ecotox Environ Safe 180:139-145. http://dx.doi.org/10.1016/j.ecoenv.2019.05.007 2102 Tomizawa M (2004) Neonicotinoids and derivatives: Effects in mammalian cells and mice. J Pestic Sci 29:177-2103 183. https://doi.org/10.1584/jpestics.29.177 2104 Tooker JF, Pearsons KA (2021) Newer characters, same story: neonicotinoid insecticides disrupt food webs 2105 through direct and indirect effects. Curr Op Insect Sci 46:50-56. https://doi.org/10.1016/j.cois.2021.02.013

2106	Topal A, Alak G, Ozkaraca M, Yeltekin AC, Comaklı S, Acıl G, Kokturk M, Atamanalp (2017) Neurotoxic						
2107	responses in brain tissues of rainbow trout exposed to imidacloprid pesticide: Assessment of 8-hydroxy-2-						
2108	deoxyguanosine activity, oxidative stress and acetylcholinesterase activity. Chemosphere 175:186-191.						
2109	https://doi.org/10.1016/j.chemosphere.2017.02.047						
2110	Torres JB, Silva-Torres CSA, Barros R (2003) Relative effects of the insecticide thiamethoxam on the predator						
2111	Podisus nigrispinus and the tobacco whitefly Bemisia tabaci in nectaried and nectariless cotton. Pest Manage						
2112	Sci 59(3):315-323. http://dx.doi.org/10.1002/ps.640						
2113	Tu C, Wang Y, Duan W, Hertl P, Tradway L, Brandenburg R, Lee D, Snell M, Hu S (2011) Effects of fungicides						
2114	and insecticides on feeding behavior and community dynamics of earthworms: Implications for casting						
2115	control in turfgrass systems. Appl Soil Ecol 47:31-36. http://dx.doi.org/10.1016/j.apsoil.2010.11.002						
2116	Tyler B (2022) Assessment of duckling abundance as a biological indicator of wetland health in the Prairie Pothole						
2117	Region. Master of Science Thesis. https://harvest.usask.ca/handle/10388/14291 Accessed 20 September						
2118	2023						
2119	Uhl P, Bucher R, Schäfer RB, Entling MH (2015) Sublethal effects of imidacloprid on interactions in a tritrophic						
2120	system of non-target species. Chemosphere 132:152-158.						
2121	https://doi.org/10.1016/j.chemosphere.2015.03.027						
2122	Uhl P, Brühl CA (2019) The impact of pesticides on flower-visiting insects: A review with regard to European						
2123	risk assessment. Environ Tox Chem 38(11):2355-2370. http://dx.doi.org/10.1002/etc.4572						
2124	van der Sluijs JP, Simon-Delso N, Goulson D, Maxim L, Bonmatin JM, Belzunces LP (2013) Neonicotinoids, bee						
2125	disorders and the sustainability of pollinator services. Curr Op Environ Sustain 5(3-4):293-305.						
2126	https://doi.org/10.1016/j.cosust.2013.05.007						
2127	van Dijk TC, Van Staalduinen MA, Van der Sluijs JP (2013) Macro-invertebrate decline in surface water polluted						
2128	with imidacloprid. Plos One 8(5):10. http://dx.doi.org/10.1371/journal.pone.0062374						
2129	van Meter RJ, Glinski DA, Henderson WM, Garrison AW, Cyterski M, Purucker ST (2015) Pesticide uptake						
2130	across the amphibian Dermis through soil and overspray exposures. Arch Environ Contam Toxico						
2131	69(4):545-556. http://dx.doi.org/10.1007/s00244-015-0183-2						
2422							
2132	Vijver MG, van den Brink PJ (2014) Macro-invertebrate decline in surface water polluted with imidacloprid: A						

2134 Vinebrooke RD, Cottingham KL, Norberg J, Scheffer M, Dodson SI, Maberly SC, Sommer U (2004) Impacts of 2135 multiple stressors on biodiversity and ecosystem functioning: the role of species co-tolerance. Oikos 2136 104(3):451-457. 2137 Vyas NB (1999) Factors influencing estimation of pesticide-related wildlife mortality. Toxicol Indust Health 15(1-2138 2):187-192. http://dx.doi.org/10.1177/074823379901500116 2139 Walters KFA (2016) Neonicotinoids, bees and opportunity costs for conservation. Insect Conserv Div 9(5):375-2140 383. http://dx.doi.org/10.1111/icad.12177 2141 Wang K, Pang S, Mu XY, Qi SZ, Li DZ, Cui F, Wang CJ (2015) Biological response of earthworm, Eisenia fetida, 2142 five neonicotinoid insecticides. Chemosphere 132:120-126. 2143 http://dx.doi.org/10.1016/j.chemosphere.2015.03.002 Wang Y, Yang G, Dai D, Xu Z, Cai L, Wang Q, Yu Y (2017) Individual and mixture effects of five agricultural 2144 pesticides on zebrafish (Danio rerio) larvae. Environ Sci Pollut Res 24(5) 4528-4536. 2145 2146 https://doi.org/10.1007/s11356-016-8205-9 Wang P, Zhou LL, Yang F, Liu XM, Wang Y, Lei CL, Si SY (2018a) Lethal and behavioral sublethal side effects 2147 2148 of thiamethoxam on the predator Harmonia axyridis. Entomol Exp Applic 166(8):703-712. 2149 http://dx.doi.org/10.1111/eea.12702 2150 Wang YH, Zhang Y, Xu P, Guo BY, Li W (2018b) Metabolism distribution and effect of thiamethoxam after oral 2151 exposure in Mongolian Racerunner (Eremias argus). J Agric Food Chem 66(28):7376-7383. 2152 http://dx.doi.org/10.1021/acs.jafc.8b02102 2153 Wang YH, Zhang Y, Li W, Yang L, Guo BY (2019) Distribution, metabolism and hepatotoxicity of neonicotinoids 2154 in small farmland lizard and their effects on GH/IGF axis. Sci Total Environ 662:834-841. 2155 http://dx.doi.org/10.1016/j.scitotenv.2019.01.277 2156 Wang ZK, Tian ZN, Chen L, Zhang WJ, Zhang LY, Li Y, Diao JL, Zhou ZQ (2020) Stereoselective metabolism 2157 and potential adverse effects of chiral fungicide triadimenol on Eremias argus. Environ Sci Pollut Res 2158 27(8):7823-7834. http://dx.doi.org/10.1007/s11356-019-07205-4

74

Wang JQ, Yin R, Liu YL, Wang BJ, Wang NN, Xiao PF, Xiao TF, Hirai H (2023) Meta-analysis of neonicotinoid

waters.

Environ

Sci

Pollut

Res

30:1039-1047.

surface

2159

2160

2161

insecticides

in

global

http://dx.doi.org/10.1007/s11356-022-22270-y

2162	Warne MSJ, Turner RDR, Davis AM, Smith R, Huang A (2022) Temporal variation of imidacloprid concentration
2163	and risk in waterways discharging to the Great Barrier Reef and potential causes. Sci Total Environ
2164	823:153556. https://doi.org/10.1016/j.scitotenv.2022.153556
2165	Weston DP, Poynton HC, Wellborn GA, Lydy MJ, Blalock BJ, Sepulveda MS, Colbourne JK (2013) Multiple
2166	origins of pyrethroid insecticide resistance across the species complex of a nontarget aquatic crustacean,
2167	Hyalella azteca. Proc Natio Acad Sci USA, 110(41):16532-16537. https://doi.org/10.1073/pnas.1302023110
2168	Wettstein FE, Kasteel R, Garcia Delgado MF, Hanke I, Huntscha S, Balmer ME, Poiger T, Bucheli TD (2016)
2169	Leaching of the neonicotinoids thiamethoxam and imidacloprid from sugar beet seed dressings to subsurface
2170	tile drains. J Agric Food Chem 64(33):6407-6415. http://dx.doi.org/10.1021/acs.jafc.6b02619
2171	Whitehorn PR, O'Connor S, Wackers FL, Goulson D (2012) Neonicotinoid pesticide reduces bumble bee colony
2172	growth and queen production. Science 336(6079):351-352. http://dx.doi.org/10.1126/science.1215025
2173	Wiest L, Bulete A, Giroud B, Fratta C, Amic S, Lambert O, Pouliquen H, Arnaudguilhem C (2011) Multi-residue
2174	analysis of 80 environmental contaminants in honeys, honeybees and pollens by one extraction procedure
2175	followed by liquid and gas chromatography coupled with mass spectrometric detection. J Chrom A
2176	1218(34):5743-5756. https://doi.org/10.1016/j.chroma.2011.06.079
2177	Wilcox AAE, Newman AEM, Raine NE, Mitchell GW, Norris DR (2021) Effects of early-life exposure to
2178	sublethal levels of a common neonicotinoid insecticide on the orientation and migration of monarch
2179	butterflies (Danaus plexippus). J Exp Biol 224:jeb230870. http://dx.doi.org/10.1242/jeb.230870
2180	Willemsen RE, Hailey A (2001) Effects of spraying the herbicides 2,4-D and 2,4,5-T on a population of the tortoise
2181	Testudo hermanni in southern Greece. Environ Pollut 113(1):71-78. http://dx.doi.org/10.1016/s0269-
2182	7491(00)00160-3
2183	Williams N, Sweetman J (2019) Effects of neonicotinoids on the emergence and composition of chironomids in
2184	the Prairie Pothole Region. Environ Sci Pollut Res 26:3862-3868. http://dx.doi.org/10.1007/s11356-018-
2185	3683-6
2186	Wood TJ, Goulson D (2017) The environmental risks of neonicotinoid pesticides: a review of the evidence post
2187	2013. Environ Sci Pollut Res 24(21):17285-17325. http://dx.doi.org/10.1007/s11356-017-9240-x
2188	Woodcock BA, Isaac NJB, Bullock JM, Roy DB, Garthwaite DG, Crowe A, Pywell RF (2016) Impacts of
2189	neonicotinoid use on long-term population changes in wild bees in England. Nature Com 7:12459.
2190	https://doi.org/10.1038/ncomms12459

2191	Woodcock BA, Bullock JM, Shore RF, Heard MS, Pereira MG, Redhead J, Ridding L, Dean H, Sleep D, Henry						
2192	P, Peyton J, Hulmes S, Hulmes L, Sarospataki M, Saure C, Edwards M, Genersch E, Knabe S, Pywell R						
2193	(2017) Country-specific effects of neonicotinoid pesticides on honey bees and wild bees. Science						
2194	356(6345):1393-1395. http://dx.doi.org/10.1126/science.aaa1190						
2195	Wu S, Li X, Liu X, Yang G, An X, Wang Q, Wang Y (2018) Joint toxic effects of triazophos and imidacloprid of						
2196	zebrafish (Danio rerio). Environ Pollut 235:470-481. https://doi.org/10.1016/j.envpol.2017.12.120						
2197	Wu CH, Lin CL, Wang SE, Lu CW (2020) Effects of imidacloprid, a neonicotinoid insecticide, on the echolocation						
2198	system of insectivorous bats. Pestic Biochem Physiol 163:94-101.						
2199	https://doi.org/10.1016/j.pestbp.2019.10.010						
2200	Wu-Smart J, Spivak M (2018) Effects of neonicotinoid imidacloprid exposure on bumble bee (Hymenoptera:						
2201	Apidae) queen survival and nest initiation. Environ Entomol 47(1):55-62.						
2202	http://dx.doi.org/10.1093/ee/nvx175						
2203	Xu W, Zhang L, Hou J, Du X, Chen L (2023) Absorption and distribution of imidacloprid and its metabolites in						
2204	Goldfish (Carassius auratus Linnaeus). Toxics 11:619. https://doi.org/10.3390/toxics11070619						
2205	Yamamuro M, Komuro T, Kamiya H, Kato T, Hasegawa H, Kameda Y (2019) Neonicotinoids disrupt aquation						
2206	food webs and decrease fishery yields. Science 366(6465):620-623. https://doi.org/10.1126/science.aax344						
2207	Yang L, Shen Q, Zeng T, Li J, Li W, Wang Y (2020) Enrichment of imidacloprid and its metabolites in lizard						
2208	and its toxic effects on gonads. Environ Pollut 258:113748. https://doi.org/10.1016/j.envpol.2019.113748						
2209	Yu B, Chen ZY, Lu XX, Huang YT, Zhou Y, Zhang Q, Wang D, Li JY (2020) Effects on soil microbial community						
2210	after exposure to neonicotinoid insecticides thiamethoxam and dinotefuran. Sci Total Environ 725:138328						
2211	http://dx.doi.org/10.1016/j.scitotenv.2020.138328						
2212	Zhang M, Liang YC, Son A, Yu B, Zeng XB, Chen MS, Yin HQ, Zhang XX, Sun BL, Fan FL (2017) Loss of soi						
2213	microbial diversity may increase insecticide uptake by crop. Agric Ecosyst Environ 240:84-91						
2214	http://dx.doi.org/10.1016/j.agee.2017.02.010						
2215	Zhang C, Wang X, Kaur P, Gan J (2023) A critical review on the accumulation of neonicotinoid insecticides in						
2216	pollen and nectar: Influencing factors and implications for pollinator exposure. Sci Total Environ						
2217	899:165670. https://doi.org/10.1016/j.scitotenv.2023.165670						
2218							
2219							

2221 Figures

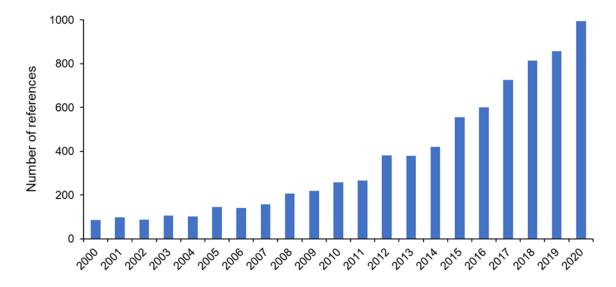


Fig. 1 Time course of references focused on the impacts of neonicotinoids on biodiversity.

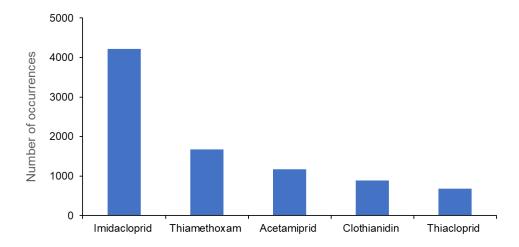


Fig. 2 Occurrences of imidacloprid, thiamethoxam, acetamiprid, clothianidin and thiacloprid in title and abstract of the references constituting the bibliographic corpus on the impacts of neonicotinoids on biodiversity, from 2000 to 2020.

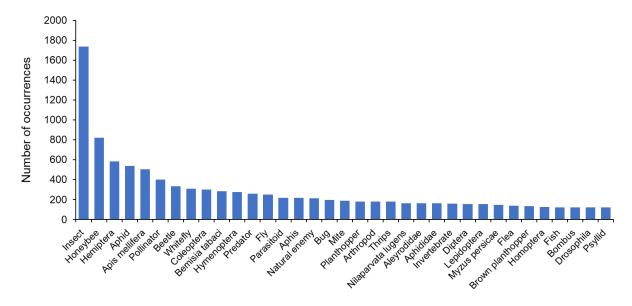


Fig. 3 Occurrences of the first 35 organisms studied in the bibliographic corpus on the impacts of neonicotinoids on biodiversity, from 2000 to 2020. Occurrences are counted from titles and abstracts. When occurring, alternative spellings were gathered into one category, for example "honeybee", "honeybees" and "honey bees".

2252 Table

Table 1 Maximum concentration levels of acetamiprid, clothianidin, imidacloprid, thiacloprid and thiamethoxam in soil, air and water observed in France, Europe and in the world. nd: not determined, *particulate phase.

Neonicotinoid	Geographic zone	graphic Soil		Air		Water	
		Concentration (µg/kg)	Reference	Concentration (ng/m³)	Reference	Concentration (µg/L)	Reference
Acetamiprid	France	0.48	Froger et al. (2023)	0.26	Phytatmo database (2023)	nd	nd
	Europe	nd	nd	0.031 (Spain)	Coscollà and Yusà (2016)	4.00 (Spain, freshwater)	de Araújo et al. (2022)
	World	nd	nd	0.036* (Canada)	Raina-Fulton (2015)	2.86 (Turkey, freshwater)	de Araújo et al. (2022)
Clothianidin	France	2.7	Froger et al. (2023)	nd	nd	nd	nd
	Europe	57 (Switzerland)	Riedo et al. (2021)	nd	nd	nd	nd
	World	nd	nd	0.09* (Canada)	Raina-Fulton (2015)	3.50 (USA, drained wetlands) 0.132 (USA, freshwater)	Evelsizer and Skopec (2018) de Araújo et al. (2022)
Imidacloprid	France	160	Pelosi et al. (2021)	2.3	Phytatmo database (2023)	2.22 (peri-urban ponds)0.905 (agricultural/urban rivers)0.14 (marine waters)	Nélieu et al. (2021) Criquet et al. (2017) Auby et al. (2011)
	Europe	138 (Switzerland)	Chiaia-Hernandez et al. (2017)	0.014 (Spain)	Coscollà and Yusà (2016)	0.342 (Spain, freshwater)	de Araújo et al. 2022
	World	nd	nd	0.36* (Canada)	Raina-Fulton (2015)	9.14 (USA, freshwater)	Wang et al. (2023)
Thiacloprid	France	1.4	Pelosi et al. (2021)	0.47	Phytatmo database (2023)	nd	nd
•	Europe	14 (Switzerland)	Riedo et al. (2021)	nd	nd	0.159 (Portugal, freshwater)	de Araújo et al. (2022)
	World	nd	nd	nd	nd	1.37 (Australia, lagoon)	Wang et al. (2023)
Thiamethoxam	France	2.0	Pelosi et al. (2021)	0.06	Phytatmo database (2023)	0.0039 (bay)	Tapie and Budzinski (2018)
	Europe	24 (Switzerland)	Riedo et al. (2021)	nd	nd	0.215 (Portugal, freshwater)	de Araújo et al. (2022)
	World	nd	nd	nd	nd	6.90 (USA, drained wetlands) 3.82 (Canada, freshwater)	Evelsizer and Skopec (2018) Wang et al. (2023)