

Bacillus thuringiensis: well armed bacteria to attack insects

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INRA A well armed bacteria to attack insects



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Bio-insecticide





Bt-transgenic-plants





Cours AgroParisTech : AIP Santé, thème Science et Société : lutte contre le paludisme Février/Mars 2023

Bacillus thuringiensis is part of the Bacillus cereus group bacteria

Firmicutes, Bacillacae, Gram+, aerobic/ anaerobic, sporeforming, saprophyte, ubiquistic

Crystal / inclusion body = composed of cry toxin-proteins



Light microscopy (Phase contrast)



spore

vip

Electronic transmission microscopy

CľV

Specific Bt insecticidal toxin genes are plasmid-born











Lifecycle of *Bacillus thuringiensis* in the insect larvae (*in vivo*)



Common mode of action steps of three domain Cry toxins



> Protease processing, receptor binding ...



FIOCRUZ

Life cycle of main mosquito species

Adults



Pupae



Larvae

only stage target for Bt crystal toxins

Eggs





B. thuringiensis sorovar.*israelensis* : Bti a bio-pesticide against dipterans





Oral larvicidal toxicity specific target insects = less side effects than chemical insecticides



Simulium

Aedes



Culex



Anopheles





Bti Based larvicides

Adapted for conventional field treatment application

Fermentation -



Easy large scale production

Biomass spores & Crystals

Various formulations

https://publichealth.valentbiosciences.com









eg. Vectobac









Insecticidal toxins of Bti and mode of action

Their potential against culicidae (moquitoes)

Specific targets = low environmental impact





Bti: crystal and insecticidal toxins



Activity spectrum: *Aedes, Simulium, Culex & Anopheles* = mosquistoe species = vectors of disease agents :

Plasmodium and arbo viruses like: Dengue, Chikungunya, Zika





Structure and function of Bti toxins

Cry: 3 domains



D2 and D3: Receptor binding D1: Pore formation



Cyt: 1 domain α - β



Cytolysis Poreforming and /detergent effect

(Vachon et al., ., 2014) (de Maagd et al., 2003)



Mode of action of Bti crystal toxins





Cry11Aa receptor in *Ae. aegypti*

• Immuno toxin detection in the midgut of intoxicated larva





(Fernandez et al., 2006)

Synergy between the diffrent Bti toxins

(Crickmore et al., 1995; Poncet et al., 1997)

Toxicity (ng/ml) against Ae. aegypti						
CL_{50}	Combination	CL_{50}				
10	Cry4A+CytA	75				
1125	Cry11A+CytA	118				
467	Cry4A+Cry11A	173				
224	Cry4A+Cry4B++CytA	77				
1209						
	Toxicity (ng/ml) a CL ₅₀ 10 1125 467 224 1209	Toxicity (ng/ml) against Ae. aegypti CL_{50} 10Cry4A+CytA1125Cry11A+CytA467Cry4A+Cry11A224Cry4A+Cry4B++CytA1209Cry4A+Cry4B++CytA				

CL 50=concentration that kills 50 % of the exposed larvae)



Cry4Ba

Cry4Aa

Cry11Aa

Cyt1Aa



Synergy between Cry11Aa and Cyt 1Aa



Cry11Aa binds to the membrane receptor Cry11Aa binds also to Cyt1Aa as a second receptor



(Pérez et al., 2005; Soberón et al., 2009)



Effects of Bti crystal toxins on intestinal cells of Aedes aegypti







Charles et al., 1988



Bti is very efficient in mousquito control is there a risk for development of resistance in treated populations ?





Link between Mode of action and Resistance mechanisms

Modification of receptor binding sites
 -Risk of selection for resistance/ lower affinity

 Modification of toxin processing -proteolysis of protoxins

• Detoxification enzymes

-Processes often seen for synthetic insecticides

- Mono-oxidases, transferases, esterases





Selection of resistance under laboratory conditions

Resistance levels							
Species	Generatio	n Bti	Cry4A	Cry4B	Cry11A	Cyt1A	
C. pipiens	28	3.2			913		Georghiou & Wirth 1997
	35	4.4			16	2	Wirth et al. 2010
	20	3.0					Mittal et al. 2005
	20	2.8					Saleh et al. 2003
Ae. aegypt	i 15	1.1					Goldman et al. 1986
	15 15	1.1 2.0					Goldman et al. 1986 Goldman et al. 1986
	18	3.4	14	6	30	3	Bonin et al. 2010
	18	3.0	60	4	7	4	Paris et al. 2010
	22		35	11	3		Paris et al. 2011
	54				13		Cadavid-Restrepo et al. 2012

Micalis

In the laboratory very low resistance to the full Bti toxin mix but resistance is possible to the individual toxins



Exemples of Bti field applications





Persistance of Bti in the field

Tablet comercial90-112 daysTablet comercial54-166 daysTablet experimental84 daysCommercial formulation35 days

Mulla et al., 2004 Benjamim et al., 2005 Armengol et al., 2006 Lee & Zairi, 2006

Sun exposure

Recycling (spore –toxin/crystals)

Toxins crystals must be available for larval ingestion

Persistance is larval breedingsite and formulation dependent





Control of *Simulium (Blackfly)* - againt Onchocerciasis (River blindness) with Bti In west Africa (OCP program)

- Actors : OCP/WHO
- ➡ Period: 1982- 2002
- ➡Target: Simulium damnosum complex
- →Region: 11 countries /1.3 Mill Km²/25 .0000 hab.
- ➡ Results:
 - ▼ Control of **Onchocerciasis**
 - **v** Reimplemenation of inhabitants
 - ▼ Strong socio- economic impact







Control of *Ae. vexans* in the Rhine valey, Germany (KABS)

Actor: KABS

- ➡ Period: 1983- still ongoing
- ➡Bti is "gamma irradiated" = no viable spores
- ➡ Target: Aedes vexans
- → Treated surface: 600 km2
- ➡ Results:
 - ▼ Good efficacy, optimized formulations
 - ▼ Ecological survey,
 - **v** Positive impact on tourism





Control of Simulium sp. in Brazil

Actor: Gouvernment (Health secretary)
Period: 1982- still on going
Target: *Simulium pertinax*Region:
Serra Gaúcha S: 43,000 km²/3 milhões hab.
Litoral Norte de SP: 872 km²

- ➡ Results:
 - ▼ Efficacity
 - ▼ positive Impact population and tourism







Use of Bti in France

• Actor: EID (Entente interdepartemental de démoustication)

- Period: 1990- still ongoing
- →Target: *Aedes*, *Culex spp*
- ➡Region:
 - > Rhônes-Alpes : www.eid-rhonealpes.com
 - Mediteranean http://www.eid-med.org
 - > Atlantic coast www.eidatlantique.eu
- ➡ Results:
 - **V** Decrease in nuisance
 - Positive Impact on population and tourism







Integrated control of Aedes in Brazil

Excellent larvicidal activity field efficacy Possibility to be used in integrated control







Life cycle of main mosquito species

Adults





Pupae





Larvae



Eggs





Control of Aedes: integrated control measures are needed

Monitoring





Elimination of egg laying areas

Capture of adults mousquitoes











Use of Bti in Brazil to control Dengue disease





2000-2002: Detecion of resistance against Organophosphate insectides (OP)

Application of Bti againt OP resistant populations





Bti as a possible alternative to Temephos

> Resistance to Temephos is related to detoxication enzymes

> Is there a possibility of cross resistance between Bti and Temephos ?









Bti Susceptibility of *Aedes* **populations (strains)**

with moderate resistance to Temephos

Strain	exposition	Temephos RR	RR Bti
Rock	Non	1	1.0
F. Noronha	Bti	2.4	1.6
Recife	T-Bti	7.1	1.8
Bacabal	T-Bti-IGR	6.6	1.8
Macapa	T-Bti-IGR	11.0	1.5
J.Pessoa	T-Bti-IGR	11.7	1.4

T= Temephos, IGR = inhibition of growth regulator RR= resistancde level ; 1 = no resistance





Resistance ratio (RR): Temephos and Bti



Temephos RR: 2 to 253 fold

Bti RR: 1 to 2 fold

No cross resistance to Bti





Conclusion related to Bti

- > Bti very low risk for resistance development (several toxins and different host receptors, a complex mode of action)
- > No cross resistance between Bti and the Organophosphate insecticide Temephos
- Bti can be used to control of *Aedes* and other mosquitoes like *Anopheles* to decrease malaria





Are Bt strains really without risk for non targets ?

• Recommendet for use in drinking water (WHO, 1999, 2012) !

• <u>https://www.efsa.europa.eu/fr/efsajournal/pub/4524</u> 2016

Risk of Bt based larvicides ?

Issues: Not only Cry toxins in Bt Products What about the spores / bacteria ?

Bt is part of the *B. cereus* group !



Bacillus cereus group

Bacilli, Gram positif, spore forming & ubiquistic

Common gene pool (~5.5 Mb)

Bacillus thuringiensis Specific genes



Entomopathogen Crystal = specific "Insecticide"



Bacillus cereus **Opportunistic pathogen** Human Animal Gastro intestinal diseases Local and systemic infection

Animal & human pathogen 'Antrax disease"

can

tox



Are Bt strains really without risk for non targets ?

Recommended for use in drinking water (OMS, 1999, 2012, but not at present) !

https://www.efsa.europa.eu/fr/efsajournal/pub/4524 2016

risk of Bt based larvicides? **Issues :** Not only - Cry toxins in Bt Product What about the spore/bacteria Member of the *B. cereus* group !

> Need for « OGM » Bt strain ? Without spores = **MosKO project** in the Micalis GME team More than 60 years of use in rivers, forest without problems !







https://satt-paris-saclay.fr/en/technological-projects-portfolio/mosko/

A new safer and more efficient mosquitocidal larvicide

Today



Bt *serovar israelensis* (Bti), the only mosquitocidal larvicide autorized in France Spores are spread into the environment

The Bti toxins are very efficient

But:

Need to increase the bio-availability of the Cry toxin's due to UV degradation, adsorbance to soil

The spores are spead into the environment = water = potential risk in food contamination



MosKO

Objectifs/ project ongoing



- Absence of bacterial spores
- Bio-larvicidal toxins
 - ✓ Several toxins= increased efficacity/more targets
 - ✓ Protection against UV
 - ✓ Increased bio-availability







Encapsulated crystals NO spores



« Muito obrigada » to my freind and colleague

Maria Helena Neves Lobo Silva Filha FIOCRUZ -PE, Brésil

Merci pour votre attention !





