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## The anchoring of the polysaccharide II is essential for *Clostridioides difficile* survival

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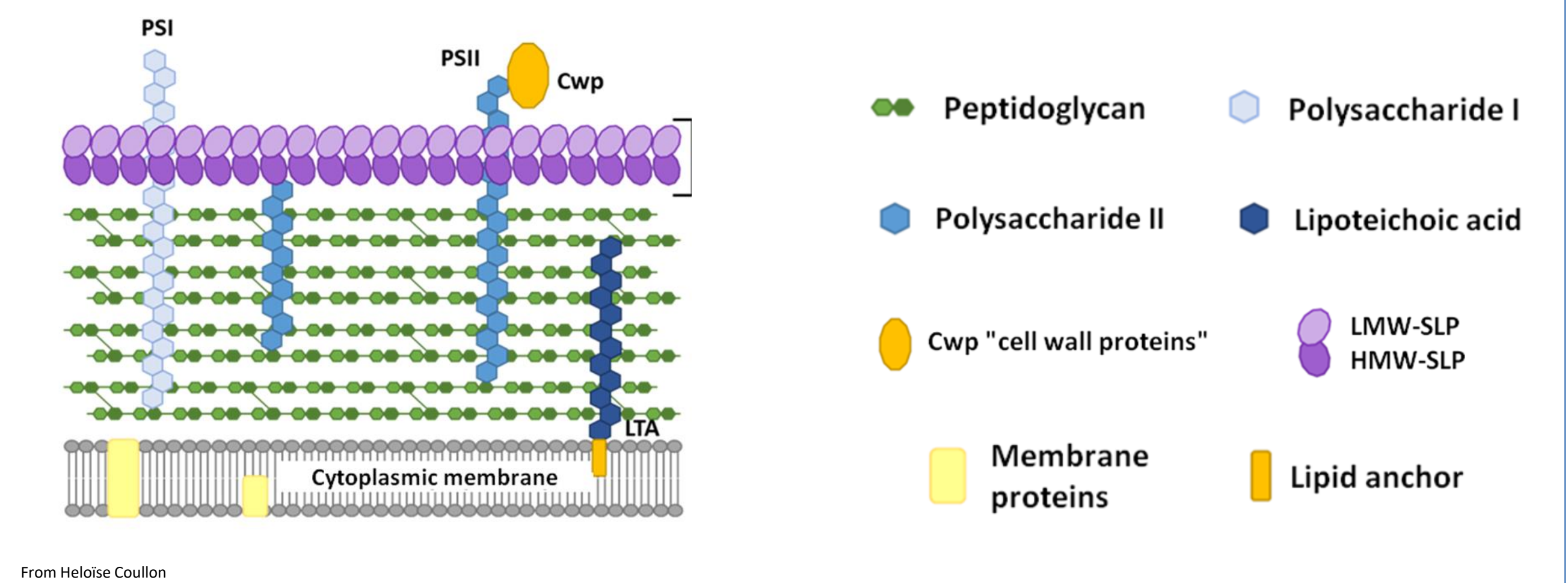
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# The anchoring of the polysaccharide II is essential for *Clostridioides difficile* survival

## INTRODUCTION

*C. difficile* is an anaerobic, motile and spore-forming bacterium, responsible for 15 to 25% of post-antibiotic diarrhea and 95% of pseudomembranous colitis. While its toxins are described to be the major virulence factors in *C. difficile* infections, there is an increasing interest in the role of non-toxin factors in pathogenesis and virulence. In many other pathogens, cell wall glycopolymers influence the virulence. In *C. difficile*, three major carbohydrates are described: the polysaccharide I (PSI), the polysaccharide II (PSII) and the lipoteichoic acid (LTA). In the synthesis model suggested before<sup>1</sup>, the polysaccharide II is synthesized in the cytoplasm, then exported to the surface by a flippase and finally anchored into the peptidoglycan by two proteins of the LytR family: LcpA and LcpB. The two *lcp* genes are supposed to be non-redundant and were previously disrupted by insertional inactivation<sup>1</sup>.



## AIMS OF THE STUDY

The aims of the study were first to investigate the **role of the polysaccharide II and its correct anchoring** for growth, surface set-up and survival of *C. difficile* and also to develop an innovative technique to **study essential genes** in *C. difficile*.

## RESULTS

### CONDITIONAL-LETHAL MUTANT TECHNIQUE TO STUDY ESSENTIAL GENES

#### Single and double mutation of the *lcp* genes

➤  $\Delta lcpA$  and  $\Delta lcpB$ : constructed by allele-coupled exchange technique

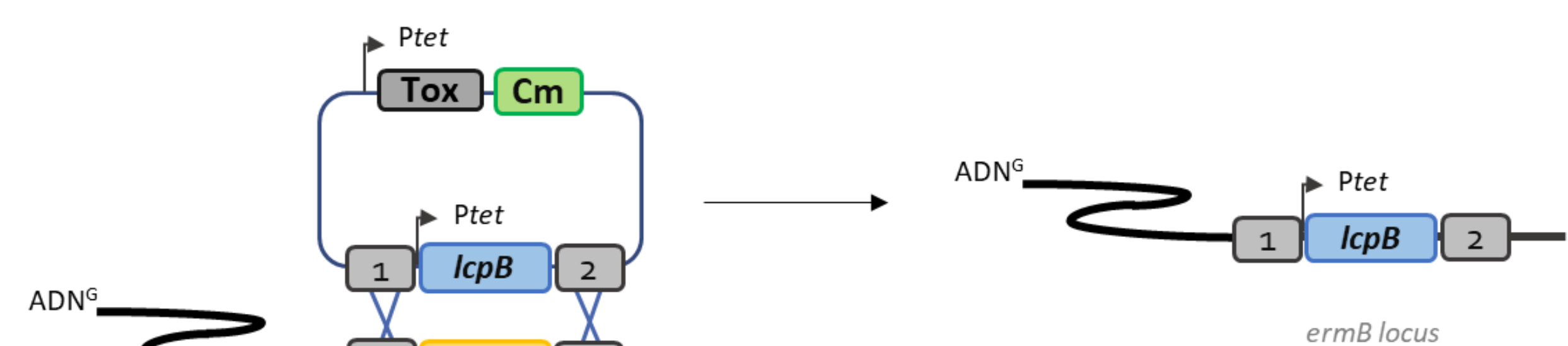
	$\Delta lcpA$	$\Delta lcpB$
Microscopic aspect		
Nb mutant / clones screened	21 / 25	3 / 187

➤  $\Delta lcpA \Delta lcpB$ : unsuccessful to obtain double mutant  
Only WT revertants found in PCR over 457 clones screened

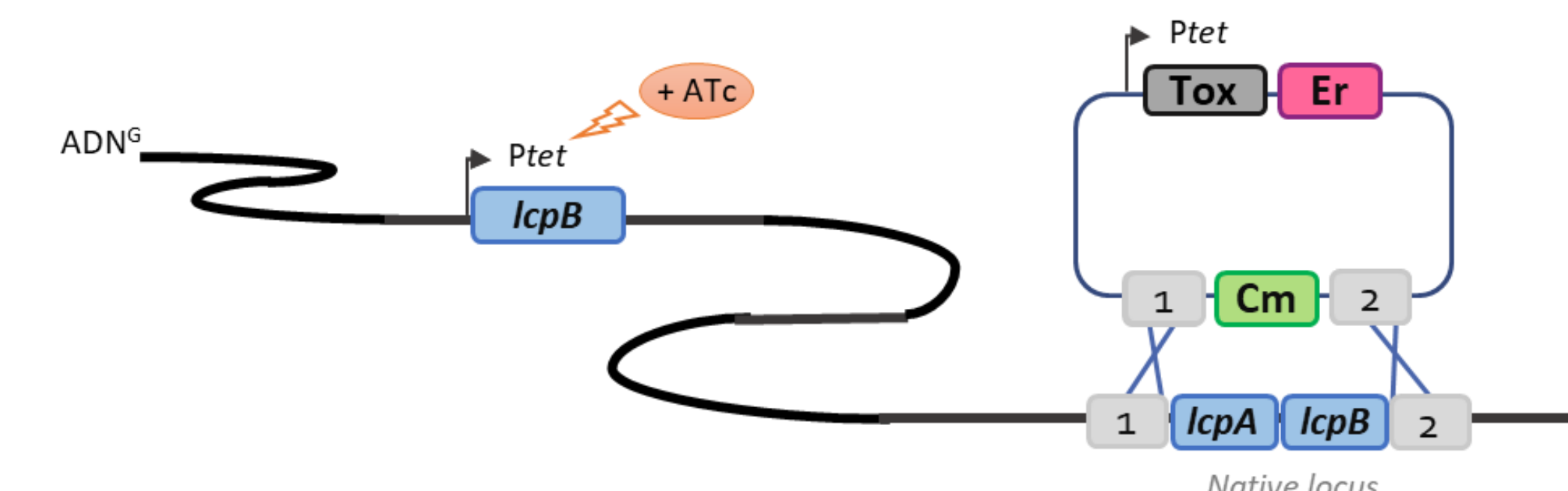
➔ It suggests that LcpA and LcpB have redundant functions and the activity of at least one Lcp seems to be essential for survival

#### Obtention of a conditional-lethal mutant

**Step 1**: insert the supplementary copy into the *ermB* region of the chromosome by ACE and verify by PCR



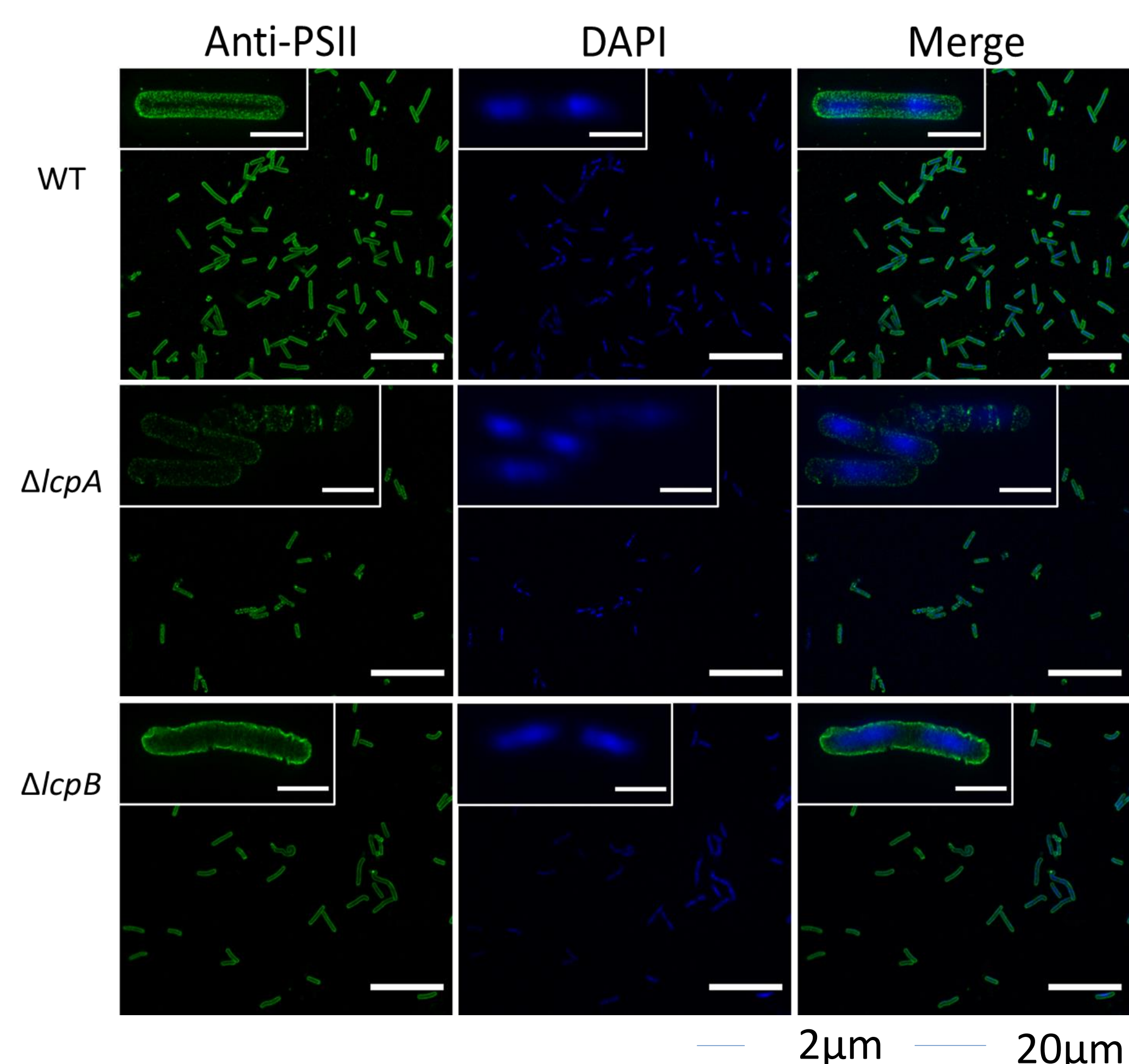
**Step 2**: induce the additional copy with ATc and delete the gene in the native locus



➔ Modulation of the gene expression to allow survival and observe phenotype  
➔ Double *lcp* mutant is not able to grow without ATc ➔ LcpA and LcpB are redundant  
➔ PSII anchoring is essential

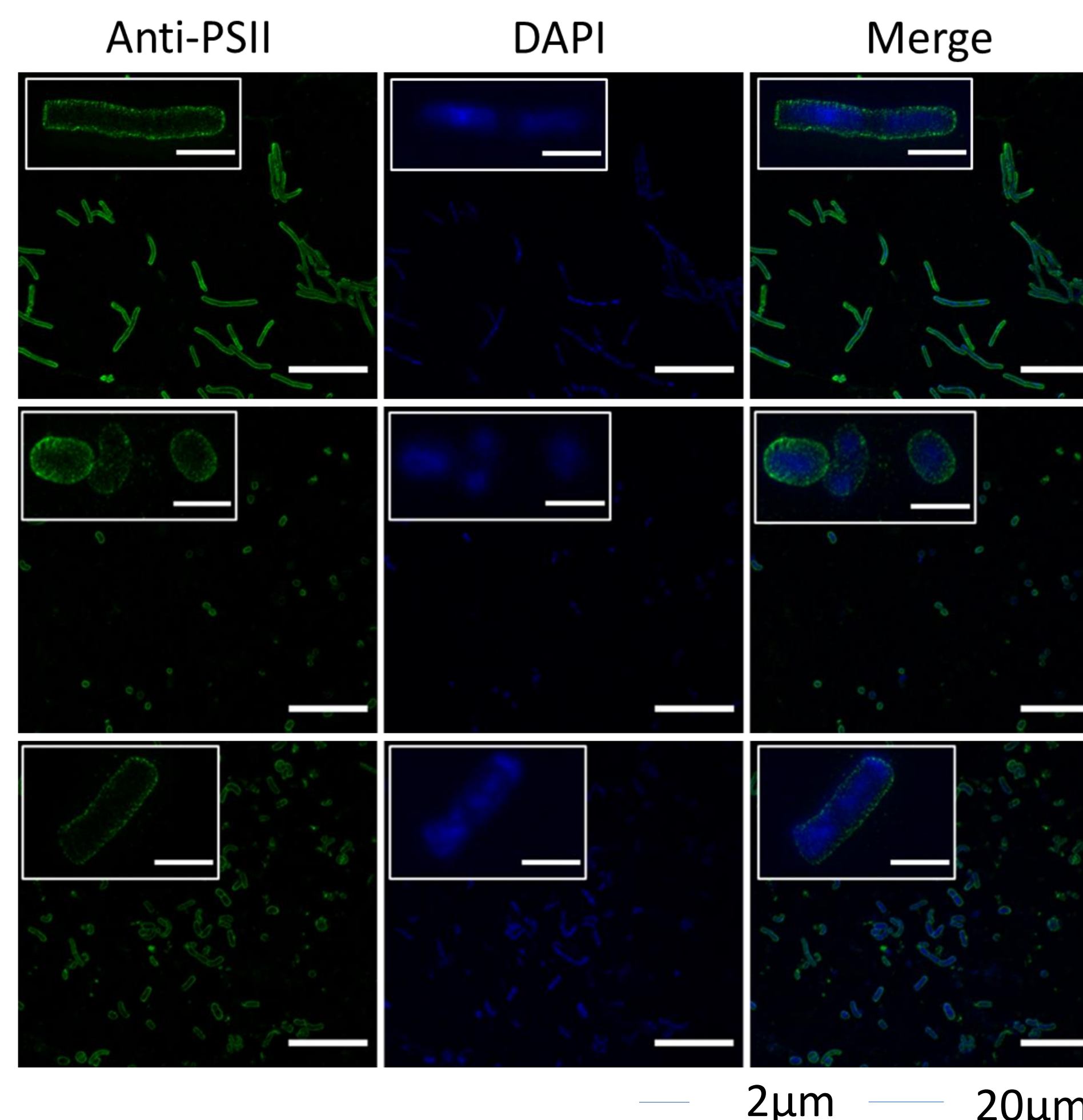
### MUTANTS ANALYSIS BY IMMUNOFLUORESCENCE MICROSCOPY AND WESTERN BLOT

#### Single mutants PSII layer



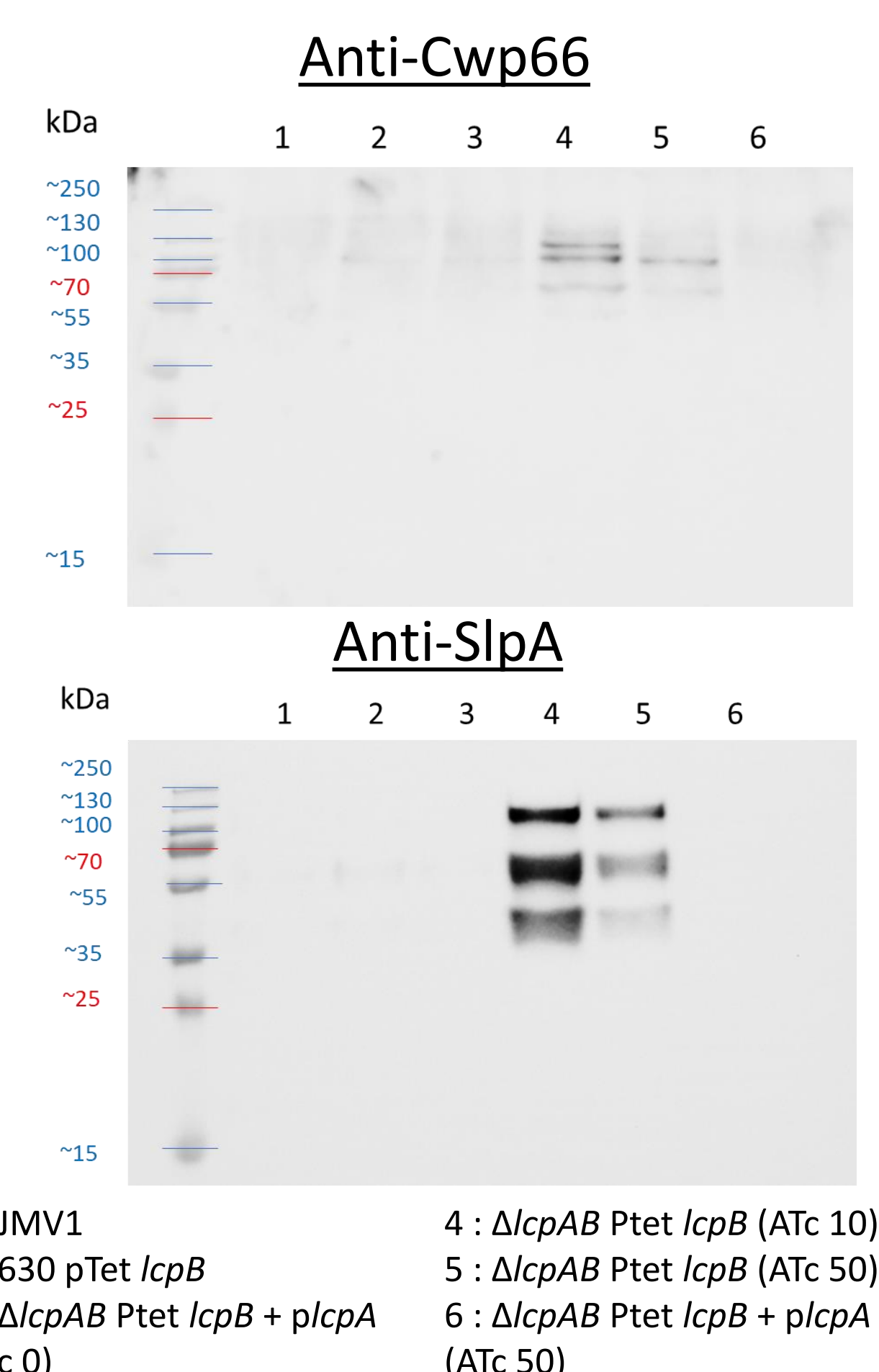
$\Delta lcpA$  mutant strain has a holed layer of PSII and  $\Delta lcpB$  mutant strain has a smooth layer and deformed cells

#### Double conditional-lethal mutant PSII layer



Low induction of *lcpB* gene leads to ellipsoid cells, unable to elongate correctly ➔ PSII anchoring is important for a correct bacterial elongation

#### Protein analysis of the supernatant by WB



1 : JMV1  
2 : 630 pTet *lcpB*  
3 :  $\Delta lcpAB$  Ptet *lcpB* + *plcpA* (ATc 0)  
4 :  $\Delta lcpAB$  Ptet *lcpB* (ATc 10)  
5 :  $\Delta lcpAB$  Ptet *lcpB* (ATc 50)  
6 :  $\Delta lcpAB$  Ptet *lcpB* + *plcpA* (ATc 50)

SlpA and Cwp66 proteins are found in the culture supernatant of the mutant

## CONCLUSION - PERSPECTIVES

Thanks to the construction of the single *lcp* mutants by allelic exchange, we confirmed the phenotypes previously observed<sup>1</sup>. The impossibility to isolate a double *lcp* mutant clone tends to confirm the essentiality of the PSII anchoring into the peptidoglycan. This was confirmed using the conditional-lethal mutant of both *lcp*, which is completely unable to grow without induction of the supplementary copy of *lcpB* added in the chromosome. When *lcpB* expression is very low, we observe ellipsoid cells, unable to elongate. Our results show the critical role of polysaccharide II anchoring in growth, elongation, and correct surface set-up of *C. difficile*. Our technique provides new opportunities to study essential genes in *C. difficile*.