



HAL
open science

Genome sequences of four colistin-resistant **ESKAPE** bacterial strains isolated from patients within the same hospital

Merve Nur Tunç, Virgile Guéneau, Valentin Loux, Rosa del Campo, Rut Carballido Lopez, Romain Briandet

► **To cite this version:**

Merve Nur Tunç, Virgile Guéneau, Valentin Loux, Rosa del Campo, Rut Carballido Lopez, et al.. Genome sequences of four colistin-resistant ESKAPE bacterial strains isolated from patients within the same hospital. *Microbiology Resource Announcements*, 2023, 10.1128/mra.00874-23 . hal-04353850

HAL Id: hal-04353850

<https://hal.inrae.fr/hal-04353850>

Submitted on 19 Dec 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NoDerivatives 4.0 International License

Genome sequences of four colistin-resistant ESKAPE bacterial strains isolated from patients within the same hospital

Merve Nur Tunç,¹ Virgile Guéneau,^{1,2} Valentin Loux,^{3,4} Rosa del Campo,⁵ Rut Carballido Lopez,¹ Romain Briandet¹

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT The genomes of four clinical Gram-negative ESKAPE bacterial strains highly resistant to the last-resort antibiotic colistin were sequenced and analyzed. The strains were found to carry multidrug-resistant genes besides colistin-resistant genes.

KEYWORDS *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, colistin-resistance, multi-drug resistant, ESKAPE

Colistin (polymyxin E) is a last-resort antibiotic used against multidrug-resistant (MDR) Gram-negative bacteria. However, its efficacy is increasingly compromised by the emergence of colistin-resistant strains. Here, we sequenced the genomes of four colistin-resistant strains isolated from unrelated patients suspected of infection at the Microbiology Department of the Ramón y Cajal University Hospital of Madrid, Spain, with the approval from The Drug Research Ethics Committee of the hospital (CEIm, identification number 2017-163/17). The strains belong to two different species: two are of *Pseudomonas aeruginosa* and two of *Klebsiella pneumoniae*. The *P. aeruginosa* strains were isolated from the sputum of cystic fibrosis patients and *K. pneumoniae* strains from a patient with a urinary tract infection. Bacterial samples were grown on McConkey growth medium (Difco) at 37°C and identified by MALDI-TOF MS (Burker).

The minimum inhibitory concentration (MIC) for colistin was determined according to the EUCAST procedure, using the microdilution method with the cationic-adjusted Mueller–Hinton broth medium (Difco). The MIC values of all strains were well above the species breakpoint (2 mg/L for *K. pneumoniae* and 4 mg/L for *P. aeruginosa*), classifying them as colistin-resistant (Table 1).

For genome sequencing, strains were retrieved from –70°C glycerol stocks and cultured overnight in Lysogenic Broth medium at 37°C with agitation. Genomic DNA was extracted using the GenElute Bacterial Genomic DNA Kits (Sigma-Aldrich), quantified using the Qubit dsDNA Kit (Thermo Fisher Scientific) and sent to Eurofins Genomics Europe Sequencing (Germany) for sequencing. DNA-seq libraries were prepared according to Eurofins Illumina's protocol and sequenced on NovaSeq 6000 with 2 × 150 bp paired-end read mode and output of approximately 5 million paired-end reads per sample. Reads quality was checked by FastQC (version:0.11.9) (1).

The reads were analyzed using the Galaxy software (<https://galaxy.migale.inrae.fr/>) (2). For each tool, default parameters were used except where otherwise stated. *De novo* assembly was performed using Unicycler (Galaxy version:0.4.8.0) (3) and quality control using Quast (Galaxy version:5.0.2+galaxy4) (4–7). Genome annotation was completed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP version:6.5) (8–10). Total genome size, percent genome coverage, number of contigs, raw read pairs, CDS, rRNAs, tRNAs, GC content, N50 value, and the identified plasmids are listed in Table 1.

In addition to antimicrobial resistant genes annotated by Staramr (Galaxy version:0.8.0+galaxy0) (11), genes known to be associated with colistin tolerance/resistance mechanisms in the annotated genome sequences are listed in Table 1. Colistin is a

Editor Catherine Putonti, Loyola University Chicago, Chicago, Illinois, USA

Address correspondence to Rut Carballido Lopez, rut.carballido-lopez@inrae.fr, or Romain Briandet, romain.briandet@inrae.fr.

The authors declare no conflict of interest.

See the funding table on p. 3.

Received 15 September 2023

Accepted 18 November 2023

Published 19 December 2023

Copyright © 2023 Tunç et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Genome assembly details and statistics of *P. aeruginosa* and *K. pneumoniae* isolated from patients within the same hospital

	PAMNT027	PAMNT030	PAMNT028	PAMNT034
Species	<i>P. aeruginosa</i>	<i>P. aeruginosa</i>	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
Genome size (bp)	6,237,612	6,227,686	5,320,190	5,599,993
The total read coverage (x)	141.0	197.7	140.9	129.2
No. of contigs	55	55	46	84
GC content (%)	66.54	66.53	57.51	57.17
No. of raw read pairs	5.823.045	8.152.234	4.966.094	4.790.919
<i>N</i> ₅₀ value (bp)	703,869	693,914	444,847	222,209
No. of CDS	5,746	5,735	5,095	5,500
No. of rRNAs	3	3	4	4
No. of tRNAs	57	57	74	77
Antimicrobial-resistant genes annotated by Staramr	<i>aph3^{IIb}</i> , <i>blaOXA-486</i> , <i>blaPAO</i> , <i>catB7</i> , <i>fosA</i>	<i>aph3^{IIb}</i> , <i>blaOXA-486</i> , <i>blaPAO</i> , <i>catB7</i> , <i>fosA</i>	<i>aac3-IId</i> , <i>aac6^{IIb-cr}</i> , <i>aadA16</i> , <i>ARR-3</i> , <i>blaSHV-2</i> , <i>dfrA27</i> , <i>fosA16</i> , <i>OqxA</i> , <i>OqxB</i> , <i>qacE</i> , <i>qnrB6</i> , <i>sul1</i> , <i>tetD</i>	<i>aac3-IIa</i> , <i>aac6^{IIb-cr}</i> , <i>aadA2</i> , <i>blaCTX-M-15</i> , <i>blaOXA-1</i> , <i>blaOXA-48</i> , <i>blaSHV-182</i> , <i>catA1</i> , <i>catB3</i> , <i>dfrA12</i> , <i>fosA</i> , <i>OqxA</i> , <i>OqxB</i> , <i>qacE</i> , <i>qnrB1</i> , <i>sul1</i>
Colistin resistance/tolerance related genes	<i>mexA</i> , <i>mexB</i> , <i>mexR</i> , <i>oprM</i> , <i>emrA</i> , <i>emrB</i> , <i>arnBCADTEF</i> , <i>kdsD</i> , <i>araG</i> , <i>eptA</i> , <i>eptC</i> , <i>phoQ</i> , <i>phoP</i>	<i>mexA</i> , <i>mexB</i> , <i>mexR</i> , <i>oprM</i> , <i>emrA</i> , <i>emrB</i> , <i>arnBCADTEF</i> , <i>kdsD</i> , <i>araG</i> , <i>eptA</i> , <i>eptC</i> , <i>phoQ</i> , <i>phoP</i>	<i>mexA</i> , <i>mexB</i> , <i>oprM</i> , <i>emrA</i> , <i>emrB</i> , <i>arnBCADTEF</i> , <i>kdsD</i> , <i>araA</i> , <i>araC</i> , <i>araE</i> , <i>araF</i> , <i>araG</i> , <i>araH</i> , <i>gutQ</i> , <i>eptA</i> , <i>eptB</i> , <i>opgE</i> , <i>phoQ</i> , <i>phoP</i>	<i>mexA</i> , <i>mexB</i> , <i>oprM</i> , <i>emrA</i> , <i>emrB</i> , <i>arnBCADTEF</i> , <i>kdsD</i> , <i>araA</i> , <i>araC</i> , <i>araE</i> , <i>araF</i> , <i>araG</i> , <i>araH</i> , <i>gutQ</i> , <i>eptA</i> , <i>eptB</i> , <i>opgE</i> , <i>phoQ</i> , <i>phoP</i>
MIC for colistin	32	16	256	256
Plasmid finder	–	–	IncR	IncFIB(K), IncR
Accession no.	JASERN000000000	JASERO000000000	JASERQ000000000	JASERR000000000

cationic polypeptide that interacts with phosphate groups of the lipopolysaccharide (LPS) in the outer membrane. The mechanisms of resistance mainly involve modifications of the LPS target (12) or export of the antibiotic by multidrug efflux (Mex) systems (13). The colistin resistance-related genes *arnBCADTEF*, *kdsD*, *eptA*, *phoPQ* which are involved in the addition of either 4-amino-4-deoxy-L-arabinose (L-Ara4N) or phosphoethanolamine (pEtN) to the LPS to reduce its negative charge and thus its affinity for colistin (12, 14, 15) are present in all four genomes. The regulatory genes *mexAB*, *emrAB*, encoding components of the MDR efflux pumps Mex and MFS (13, 16, 17) are also present in all four genomes.

Two *Pseudomonas aeruginosa* and two *Klebsiella pneumoniae* colistin-resistant clinical strains were isolated from unrelated patients under suspicion of infection at the Microbiology Department of the Ramón y Cajal University Hospital of Madrid, Spain. *P. aeruginosa* strains were isolated from sputum of cystic fibrosis patients and *K. pneumoniae* strains were obtained from a patient with urinary tract infection. The four isolates were sequenced by INVIEW Illumina sequencing with 2 × 150 bp paired-end read mode and output of approximately 5 million read pairs. DNAseq libraries were prepared according to Illumina's protocol.

ACKNOWLEDGMENTS

This study was funded by INRAE. The Ph.D. of M.N. Tunç is funded by the Région Ile-de-France (DIM 1HEALTH, project No. 2021-13001574-6382 to R.C.-L. and R.B.). We are grateful to the INRAE MIGALE bioinformatics facility (MIGALE, INRAE, 2020. Migale Bioinformatics Facility, doi:10.15454/1.5572390655343293E12) for providing technical support. The authors express their gratitude to the French Antibioideal network of the Promise PPR antibioresistance ANR program for fostering valuable scientific exchanges.

AUTHOR AFFILIATIONS

¹Université Paris-Saclay, INRAE, AgroParisTech, Micalis Institute, Jouy-en-Josas, France

²Lallemand SAS, Blagnac, France

³Université Paris-Saclay, INRAE, MaAGE, Jouy-en-Josas, France

⁴INRAE, BioinfOmics, MIGALE bioinformatics facility, Université Paris-Saclay, Jouy-en-Josas, France

⁵Servicio de Microbiología, Hospital Universitario Ramón y Cajal and Instituto Ramón y Cajal de Investigación Sanitaria, Madrid, Spain

AUTHOR ORCIDS

Merve Nur Tunç  <http://orcid.org/0009-0000-4115-1137>

Rosa del Campo  <http://orcid.org/0000-0003-1147-7923>

Romain Briandet  <http://orcid.org/0000-0002-8123-3492>

FUNDING

Funder	Grant(s)	Author(s)
Région Ile-de-France, DIM 1HEALTH	2021-13001574-6382	Merve Nur Tunç
Lallemand SAS		Virgile Guéneau

AUTHOR CONTRIBUTIONS

Merve Nur Tunç, Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Software, Writing – original draft | Virgile Guéneau, Methodology, Software, Writing – review and editing | Valentin Loux, Supervision, Validation, Writing – review and editing | Rosa del Campo, Conceptualization, Resources, Writing – review and editing | Rut Carballido Lopez, Conceptualization, Funding acquisition, Project administration, Supervision, Writing – review and editing | Romain Briandet, Conceptualization, Funding acquisition, Project administration, Resources, Supervision, Writing – review and editing

DATA AVAILABILITY

This Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under the accession numbers [JASERN000000000](#), [JASERQ000000000](#), [JASERO000000000](#), [JASERR000000000](#) for strains PAMNT027, PAMNT028, PAMNT030, and PAMNT034 respectively and the draft genome assembly and annotation can be found in NCBI under BioProject number [PRJNA966919](#) and SRA numbers, [SRR25684271](#), [SRR25684269](#), [SRR25684270](#), [SRR25684268](#) for PAMNT027, PAMNT028, PAMNT030, and PAMNT034 respectively. The versions described in this paper are versions JASERN010000000, JASERQ010000000, JASERO010000000 and JASERR010000000 for PAMNT027, PAMNT028, PAMNT030, and PAMNT034 respectively.

REFERENCES

- Wingett SW, Andrews S. 2018. Fastq screen: A tool for multi-genome mapping and quality control. *F1000Res* 7:1338. <https://doi.org/10.12688/f1000research.15931.2>
- Afgan E, Baker D, Batut B, Van Den Beek M, BouvierD, Ech M, Chilton J, Clements D, Coraor N, Grüning BA, et al. 2019. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. *Nucleic Acids Res* 46:W537–W544. <https://doi.org/10.1016/j.future.2018.04.037>
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
- Mikheenko A, Valin G, Pribelski A, Saveliev V, Gurevich A. 2016. Icarus: visualizer for de novo assembly evaluation. *Bioinformatics* 32:3321–3323. <https://doi.org/10.1093/bioinformatics/btw379>
- Mikheenko A, Saveliev V, Gurevich A. 2016. MetaQUAST: evaluation of metagenome assemblies. *Bioinformatics* 32:1088–1090. <https://doi.org/10.1093/bioinformatics/btv697>
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
- Mikheenko A, Pribelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUAST-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
- Partridge SR, Di Pilato V, Doi Y, Feldgarden M, Haft DH, Klimke W, Kumar-Singh S, Liu J-H, Malhotra-Kumar S, Prasad A, Rossolini GM, Schwarz S, Shen J, Walsh T, Wang Y, Xavier BB. 2018. Proposal for assignment of allele numbers for mobile colistin resistance (MCR) genes. *J Antimicrob Chemother* 73:2625–2630. <https://doi.org/10.1093/jac/dky262>
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz

- M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the prokaryotic genome annotation pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>
11. Bharat A, Petkau A, Avery BP, Chen JC, Folster JP, Carson CA, Kearney A, Nadon C, Mabon P, Thiessen J, Alexander DC, Allen V, El Bailey S, Bekal S, German GJ, Haldane D, Hoang L, Chui L, Minion J, Zahariadis G, Domselaar GV, Reid-Smith RJ, Mulvey MR. 2022. Correlation between phenotypic and in silico detection of antimicrobial resistance in *Salmonella enterica* in Canada using Staramr. *Microorganisms* 10:292. <https://doi.org/10.3390/microorganisms10020292>
 12. Sabnis A, Hagart KL, Klöckner A, Becce M, Evans LE, Furniss RCD, Mavridou DA, Murphy R, Stevens MM, Davies JC, Larrouy-Maumus GJ, Clarke TB, Edwards AM. 2021. Colistin kills bacteria by targeting lipopolysaccharide in the cytoplasmic membrane. *Elife* 10:e65836. <https://doi.org/10.7554/eLife.65836>
 13. Schweizer HP. 2003. Efflux as a mechanism of resistance to antimicrobials in *Pseudomonas aeruginosa* and related bacteria: unanswered questions. *Genet Mol Res* 2:48–62.
 14. Torres DA, Seth-Smith HMB, Joosse N, Lang C, Dubuis O, Nüesch-Inderbinen M, Hinic V, Egli A. 2021. Colistin resistance in gram-negative bacteria analysed by five phenotypic assays and inference of the underlying genomic mechanisms. *BMC Microbiol.* 21:321. <https://doi.org/10.1186/s12866-021-02388-8>
 15. Gabrielli L, Merlo S, Airolidi C, Sperandeo P, Gianera S, Polissi A, Nicotra F, Holler TP, Woodard RW, Cipolla L. 2014. Arabinose 5-phosphate isomerase as a target for antibacterial design: studies with substrate analogues and inhibitors. *Bioorg Med Chem* 22:2576–2583. <https://doi.org/10.1016/j.bmc.2013.08.012>
 16. Heacock-Kang Y, Sun Z, Zarzycki-Siek J, Poonsuk K, McMillan IA, Chuanchuen R, Hoang TT. 2018. Two regulators, PA3898 and PA2100, modulate the *Pseudomonas aeruginosa* multidrug resistance *mexAB-oprM* and *emrAB* efflux pumps and biofilm formation. *Antimicrob Agents Chemother* 62:e01459-18. <https://doi.org/10.1128/AAC.01459-18>
 17. Pamp SJ, Gjermansen M, Johansen HK, Tolker-Nielsen T. 2008. Tolerance to the antimicrobial peptide colistin in *Pseudomonas aeruginosa* biofilms is linked to metabolically active cells, and depends on the PMR and *mexAB-oprM* genes. *Mol Microbiol* 68:223–240. <https://doi.org/10.1111/j.1365-2958.2008.06152.x>