

# **Why incorporate plant architecture into trait-based ecology?**

Marilyne Laurans, François Munoz, Tristan Charles-Dominique, Patrick Heuret, Claire Fortunel, Sandrine Isnard, Sylvie-Annabel Sabatier, Yves Caraglio, Cyrille Violle

## **To cite this version:**

Marilyne Laurans, François Munoz, Tristan Charles-Dominique, Patrick Heuret, Claire Fortunel, et al.. Why incorporate plant architecture into trait-based ecology?. Trends in Ecology & Evolution, 2024, 39 (6), pp.524-536.  $10.1016/j.tree.2023.11.011$ . hal-04390151

# **HAL Id: hal-04390151 <https://hal.inrae.fr/hal-04390151v1>**

Submitted on 16 Oct 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



[Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License](http://creativecommons.org/licenses/by-nc/4.0/)

## **Why incorporate plant architecture into trait-based ecology?**

- 3 Marilyne LAURANS<sup>1,2#\*</sup>, François MUNOZ<sup>3#</sup>, Tristan CHARLES-DOMINIQUE<sup>2,4</sup>, Patrick HEURET<sup>2</sup>,
- 4 Claire FORTUNEL<sup>2</sup>, Sandrine ISNARD<sup>2</sup>, Sylvie SABATIER<sup>1,2</sup>, Yves CARAGLIO<sup>1,2#</sup>, Cyrille VIOLLE<sup>5#</sup>
- 5 <sup>1</sup> CIRAD, UMR AMAP, F-34398 Montpellier France
- 6 <sup>2</sup> AMAP, Univ Montpellier, CIRAD, CNRS, INRAE, IRD, Montpellier, France
- <sup>3</sup> LiPhy, Univ Grenoble-Alpes, CNRS, Grenoble, France
- 8<sup>4</sup> CNRS UMR7618, Institute of Ecology and Environmental Sciences, Paris, Sorbonne University,

Paris, France

- <sup>5</sup> CEFE, Univ Montpellier, CNRS, EPHE, IRD, Montpellier, France
- 
- # Marilyne LAURANS and François MUNOZ are co-first authors, Yves Caraglio and Cyrille Violle are

co-last authors.

\* Corresponding Author: Marilyne LAURANS, UMR AMAP CIRAD, TA A-51/PS2 Boulevard de la Lironde, 34398 MONTPELLIER CEDEX 5 (FRANCE), Phone +334 67 61 65 86 Email: [marilyne.laurans@cirad.fr](mailto:marilyne.laurans@cirad.fr)

#### **Abstract**

 Trait-based ecology has improved our understanding of the functioning of organisms, communities, ecosystems and beyond. However, its predictive ability remains limited as long as phenotypic integration and temporal dynamics are not considered. We highlight how the morphogenetic processes that shape the 3-dimensional development of a plant during its lifetime affect its performance. We show that the diversity of architectural traits allows us to go beyond organ-level traits in capturing the temporal and spatial dimensions of ecological niches and informing community assembly processes. Overall, we argue that consideration of multilevel topological, geometrical and ontogenetic features provides a dynamic view of whole-plant phenotype and a relevant framework for investigating phenotypic integration, plant adaptation and performance, and community structure and dynamics.

#### **Keywords**

Plant architecture; functional trait; ontogeny; phenotypic integration; community structure

#### **Consideration of phenotypic integration is urgently needed in trait-based ecology**

 Trait-based ecology has gained momentum over the last decades through systematic comparison of genotypes and species across scales and organizational levels (organ, organism, population, community and beyond). This comparative approach has come with the rise of standardized protocols [1] for identifying and measuring organ-level traits that are expected to reflect whole- organism functioning [2]. The promise of deciphering mechanisms with this approach has contributed to its success. For instance, traits depicting the morphology and chemical composition of a leaf are tightly linked to leaf-level metabolism across species worldwide [3]. Nonetheless, functional traits remain poor predictors of local adaptation, plant performance, population

 dynamics and community assembly in natural ecosystems [4-10]. This failure can be explained by the disintegration of the phenotype into elementary (often organ-level) features in trait-based ecology. Interestingly, botanists have long worked on comprehensive descriptions of **plant architecture** (see Glossary) [11] with explicit consideration of **phenotypic integration** across organs, time and space.

 Plants are modular organisms built by three key processes: growth, branching and duplication of branching systems (i.e. **reiteration)**. Stems and roots are **axes** growing in a given direction through successive phases of elongation and development of specific organs. For instance, stems are generally made of a series of annual shoots, which are themselves a series of internodes 10 and nodes bearing acquisitive, reproductive and defensive organs such as leaves, flowers and spines respectively. Axes can have very different patterns of development, as well as specific functions across species and within the same individual. Most plant species thus have several types of stems and roots, called **axis categories**, each with distinct morphological attributes and functions (**Box 1**). Plant architecture describes the properties of these different types of axes, the way they are organised with each other (**topology**) and throughout the plant's development (**ontogeny**) [12]. Plant architecture varies greatly between and within species [13-15] **(Box 1).** This variability can be characterized at different temporal and spatial resolutions using a set of dedicated tools and concepts initially developed for aboveground tree parts [16]. They were later extended to other growth forms (e.g., shrubs, herbs, mosses) [13, 14, 17, 18] and to belowground parts [19, 20]. The morphological specialization of axes [12], for example, can be used to characterize their functional 21 differences both between and within species [13]. In this article we mostly address the aboveground 22 architecture of woody plants for which we have more knowledge and references. We argue that bridging modern trait-based ecology and plant architecture approaches is a frontier research agenda that will enable us to tackle pivotal issues in plant ecology and evolutionary biology.

 A comprehensive quantification of whole-plant functioning is a priority in trait-based ecology so as to further assess the phenotypic drivers of local adaptation and elucidate diversity-ecosystem relationships. Functional traits are most often recorded at the organ level, but they are assumed to capture whole-plant functioning [2]. Nonetheless, because architectures vary among and within species [21] and the distribution of organs within the plant varies in consequence [22], this basic assumption is poorly validated (**Figure 1A**). Phenotypic integration is partly captured by analyzing multiple trait-trait relationships [23, 24], but these can be blurred by interspecific differences in plant architecture, as exemplified in allometric and biomass allocation patterns **(Figure 1B-C**). A direct consequence of the failure to consider plant architecture is a poor estimation of whole-plant metabolism (**Figure 1D**) and reproductive effort (**Figure 1E**). Ultimately, these biases in quantifying whole-plant functioning and performance from plant functional traits alone have major implications for characterizing community dynamics. As a notable example, community assembly studies [25] have analyzed trait patterns to infer processes at the community level using a list of commonly used traits. Single traits and organ-level traits can, by nature, hardly capture all the processes involved in resource partitioning between competing plants (**Figure 1F**). Despite the potential of plant architecture to capture the multidimensional facets of plant phenotype and spatio-temporal variation in ecological strategies, it is still mostly seen as a static description of crown, stem or root dimensions [26, 27]. Plant architecture can capture the myriad events a plant endures over its lifetime, as well as the mechanisms involved in resource capture and use. Incorporating plant architecture in the theoretical corpus of trait-based approaches is thus of critical importance given 21 the limited progress made over the past decade in explaining community-level mechanisms.

 Comparative approaches seek generalities. They are based on trait screening programs covering large sets of organisms [28] to the detriment of precision and consideration of mechanisms. Conversely, plant architecture approaches offer a comprehensive understanding of

1 the plant phenotype but their context dependency and the massive amount of information they may require a priori seem incompatible with the philosophy of modern trait-based ecology. We argue that a bridge must be built between these two disciplines, and we aim to lay its foundations.

#### **Plant growth and persistence through the lens of architecture**

 As sessile organisms, plants spread vertically and laterally depending on local environmental heterogeneity in resource and space availability. Current trait-based approaches largely focus on organ-level functions and usually overlook the distribution and number of acquisitive and reproductive organs across space and time [29-31]. Ecological studies often reduce plant architecture to its geometrical aspects [27, 32, 33], describing stature or allometric traits and forgetting the nature and organization of the axes. In root architecture studies, these two components are more often considered, but the focus is more on anchorage and acquisitive functions than spatio-temporal occupancy [34]. Thus despite its relevance for exploring the various constraints that drive plant form across ecological scales [35, 36], this allometric view of plant architecture does not fully capture the differences among species and individuals in their ability to explore and colonize belowground and aboveground space in the course of their **ontogeny**, nor the many other functions performed by plant architecture [37]. Plant ontogeny can be divided into several stages, from germination to senescence, each being characterized by a specific branching pattern and specific morphological trait values [38]. The **architecture-based approach (Figure 2)** is a framework that explicitly acknowledges different patterns of plant development in space and time to better understand the linkages between plant structure, function and performance [12, 39]. Below, we introduce the key **morphogenetic processes** involved in plant growth and persistence that should be considered in future ecological studies.

#### *Growth strategies arise from the structural and functional differentiation of axes*

 Hallé and Oldeman [16] distinguished 23 **architectural models** to represent basic plant growth strategies for exploring space, competing for light and reaching and occupying the canopy. Architectural models capture developmental constraints on plant organization but their links with ecological performance have not been extensively explored. Analysis of their components has revealed types of stems and roots, known as axis categories, based on morphological traits [12]. Interestingly, morphological differentiation strongly mirrors functional differentiation, with each axis category contributing to one or several functions (**Box 1**), including space exploration [40], resource acquisition [22, 37], vegetative or sexual reproduction [41], resource storage [42], response to disturbance [43], defence against herbivores [44], and mechanical [45] and hydraulic properties [46]. For example, trunks and branches with long internodes are responsible for most vertical and lateral exploration, respectively, while a large number of twigs and short shoots augments leaf area and contributes to flower and fruit production [22, 47]. The relative proportions of the axis categories change as ontogeny progresses through the juvenile, adult, mature and senescent stages. The nature, distribution and proportion of axis categories as well as their relative biomass at a given stage are critical components of plant functional strategies for resource capture and space occupancy [20]. As well as differentiating axes, we need to analyze, integrate and scale up growth and branching dynamics from shoot to whole-plant level. This is essential for disentangling and understanding the many sources of variability in crown properties (such as crown size and shape or foliage arrangement) [48, 49] and whole-plant performances. Incorporating these features in trait-based studies should help improve our understanding of growth and ecological strategies.

#### *The pivotal role of reiteration in persistence strategies*

 A key aspect of plant architecture is the reiteration process, which can greatly extend the plant's lifespan [50]. Reiteration can occur as part of the developmental sequence of the species (sequential reiteration), or in response to a change in resource availability (adaptive reiteration) or a disturbance (traumatic reiteration). Traumatic reiteration or resprouting is part of the recovery process following mechanical damage or in response to biotic or abiotic stress [51]. Traumatic reiteration usually begins when **epicormic shoots** become active after disturbance. It helps to increase species performance in disturbance-prone ecosystems [52]. While identifying reiteration patterns is doable in practice at species level [12], large-scale screening programs have yet to be conducted to determine which mechanisms promote reiteration abilities, how reiteration abilities trade-off with other key traits (e.g. reproduction) and how reiteration promotes species performance along environmental gradients. Vegetative propagation and clonal multiplication mainly result from total reiteration. They are frequently associated with a specialized organ, namely a horizontal stem (a stolon or a rhizome) that provides a means to escape from competition and to forage for nutrients far from the mother plant [53]. Vegetative reproduction is expressed differently across and within species, notably in relation to changing environmental conditions. For example, vegetative reproduction by resprouting is closely related to carbohydrate storage in belowground or aboveground organs [54]. Klimesova et al*.* [55] provide a valuable handbook of overlooked plant **modularity** traits, but their approach does not address their temporal and ontogenetic variation. Consideration of architecture provides a common framework for characterizing the diversity of processes and structures involved in growth and persistence throughout the plant life cycle.

- 
- 
- 

#### *Architecture sheds light on growth dynamics*

 Empirical studies in plant architecture over the last four decades have provided valuable knowledge about the link between architecture, growth, survival, and the environment [56]. Plant morphology captures individual responses to environmental factors, similarly to the way tree rings record secondary growth variations. The analysis of plant structure at a given time conveys key information not only on the past developmental trajectory but also on future potential growth and survival [57]. Conducting a retrospective analysis based on morphological descriptors is a powerful and accessible way to assess and analyze individual performance in trait-based or demographic approaches [10, 58]. Similarly, by characterizing morpho-anatomical markers [59, 60] such as scars left by buds, branches, flowers and inflorescences we can retrospectively analyze the dynamics of the axis elongation, branching and flowering that determine individual growth, survival and reproduction [61].

 Plant architecture can be characterized at different levels of organization and in varying degrees of detail by a set of generic **architectural traits** (e.g. number of branching orders, axis categories, reiteration patterns) **(Box 1)**. As with any functional trait [62, 63], the choice of which architectural traits to measure must depend on the ecological context of the study (notably the biotic and abiotic filters concerned), and on the research question. Interesting work has been done on savannah vegetation in this regard [64]but has yet to be carried out on other ecosystems. We emphasize that considering plant architecture does not only involve measuring architectural traits related to space occupancy, resource foraging and persistence strategies. The architecture-based approach (**Figure 2**) can also be used for identifying and selecting individual plants according to their ontogenetic stage or for standardizing functional trait measurements per axis category (**Box 1**). If functional traits are measured on several axes types, patterns of intra-individual trait variation can be analyzed and compared across species or individuals in different contexts. Lastly, given their plasticity and  integrative nature, some architectural traits allow easy assessment of individual whole-plant performance.

#### **Integrating functional traits into a whole-plant developmental context**

*Architecture as a framework for assessing phenotypic integration*

 Studying plant architecture highlights the intraspecific trait variation and phenotypic integration that are critical to species' adaptation and distribution along environmental gradients [65-67]. Functional traits influence individual performance through their separate and joint effects on different functions [2, 68], and these effects depend on the ontogenetic stage and the environmental context [69]. By describing topological and geometrical relationships across elementary units, plant architecture offers a hierarchical and modular framework for progressing toward a cause-effect analysis of trait covariations. By considering traits related to axis and branching patterns, we can assess the impact of organ-level traits on whole-plant traits, which in turn influence individual performance [49, 70]. Corner's rules [11], encompassing a wide range of organ-level and architectural traits, enable us to build key hypotheses about the linkages between plant architecture and functioning [71, 72]. Architectural traits are assumed to be pivotal in phenotypic networks because of the many competing functions they fulfil and constraints they impose [64, 65].

 *Ontogenetic and phenological variation in functional traits and strategies: towards an eco-devo perspective*

 Variations in plant traits during ontogeny and their ecological implications are increasingly documented [73]. By addressing the ontogenetic trajectories of individual plants in their environmental context, architecture analysis provides an opportunity to integrate the role of

developmental variation into the ecological context ("eco-devo", [74]) and thereby gain insights into

functional differentiation during lifetime.

 Throughout its ontogeny, a plant experiences a range of environmental conditions and is subject to distinct abiotic and biotic conditions which can drive ontogenetic changes in functional traits [75, 76]. On a seasonal time scale, plants also adjust their functioning through the **phenology** of growth and branching processes, so this is also a source of variation in functional traits [77]. Plant architecture provides markers of ontogenetic stages that enable us to track the adjustments made by the plant at various time scales and levels of organization.

 Developmental plasticity is also a key aspect of a plant's response to its environment. The ontogenetic sequence of organ growth and production can be adjusted in terms of the nature and duration of its stages [56, 78]. The ontogenetic trajectory of an individual thus depends both on its genotype and on its plastic response to environmental conditions **(Figure 3)**. For instance, low resource availability generally slows the individual's progress throughout the developmental sequence, and this can delay the expression of the first branches or reproductive structures. It is crucial to consider this ontogenetic plasticity (*sensu* [79]) for interpreting trait variations and understanding their underlying evolutionary and ecological mechanisms [62].

#### **How can plant architecture inform community structure and dynamics?**

 To elucidate the drivers of the structure and dynamics of a population or community one must aggregate information collected at the individual level [4, 80]. However, most past and current approaches in population ecology have ignored such information for lack of knowledge about the linkage between organismal physiology and demographic parameters [4]. In community ecology, the widespread use of organ-level traits is an obstacle to fully capturing processes associated with whole-organism resource capture and acquisition, notably resource competition. Below, we

 highlight the need to integrate plant architecture into the theoretical corpus of population and community ecology, given its capacity to provide ecophysiological information at the whole-plant level and throughout the plant's lifespan (**Figure 4**).

#### *Architecture-dependent population dynamics*

 One way to address population dynamics is to incorporate the influence of functional traits on vital rates, i.e. on the probabilities of survival, growth, mortality and reproduction in populations. Taking this Integral Projection Model (IPM) approach, Merow et al. [81] acknowledge how an individual's state affects its vital rates, and make population projections based on these rates. IPMs are usually built by predicting vital rates from state variables related to size or age and from environmental drivers. Since architecture reflects the way individual plants develop, forage for resources and reproduce in space and time, architectural traits can directly and functionally determine vital rates in IPMs (**Figure 4A**). For instance, Charles-Dominique et al. [82] showed that whole-plant architectural traits (e.g. spatial density of branches, plant module sizes) and the rate of ontogenetic variation in morphological traits had a significant effect on population cover in five shrub species. Using morphological traits rather than age to delimit ontogenetic stages stems from earlier population dynamics studies [83].

#### *Architecture mediates plant interactions and assembly*

 In a community ecology perspective, species niche differences shape the outcome of biotic interactions among individuals and thereby contribute to species coexistence [84]. Plant-plant interactions occur between neighboring individuals and involve competition for aboveground and belowground resources (light, water, nutrients) and attacks by shared herbivores and pathogens [85]. There is a link between individual vital rates and the density and functional composition of  their forest neighborhood (using the Neighborhood Crowding Index - NCI - [86]). Because the variation in architectural traits should reflect variation in competitive ability across individuals and throughout their lifetime [87], NCI should also capture architectural differences and complementarity among neighboring plants (**Figure 4B**) [88]. This agenda should help us better understand the assembly and coexistence of individuals with different architectural strategies.

 Taking a process-based perspective, we can make interaction coefficients in community dynamics models (derived, for example, from the Lotka-Volterra (LV) equations) depend on architectural trait variations among individuals in the community [25]. Recently, Chalmandrier et al. [89] proposed an LV-based model of community dynamics in which the parameters depend on functional trait values. We suggest exploring the influence of architectural traits that drive competitive ability (e.g. reiteration pattern, growth direction) on LV parameters. Recent advances in 3D modelling and simulation also offer new perspectives for designing virtual experiments of multispecies growth dynamics and identifying the influence of architectural diversity on the relative abundance and biomass of coexisting species.

#### *Variations in the functional structure of plant communities along environmental gradients*

 Variations in composition, in terms of architectural traits, along environmental gradients reflect the way different architectural trait values best fit different environmental conditions (**Figure 4C**). Using community assembly modelling approaches, e.g., based on null models, we can test how architectural trait composition responds to environmental filtering [90]. For instance, in a field experiment, Churski et al. [90] showed that the long-term survival of temperate tree species is influenced by their capacity to branch in response to herbivore predation. Variations in light conditions, especially during ecological succession, change architectural trait composition in a forest [91].

 At a broad spatial scale, functional biogeography aims to characterize the macroecological patterns and processes underlying the variation of functional composition across assemblages [92]. These recent advances have focused on characterizing functional variation in terms of organ-level morphological and ecophysiological traits. But plant life forms also vary across biomes depending on the abiotic environment [13], and plant architecture research should integrate broad-scale variations in organ-level properties with variations in plant structure. Understanding the way communities respond to environmental variation lays the foundation for predicting their response to future conditions. This is a paramount objective for assessing the impacts of pervasive global change. Functional diversity, especially the presence of functionally original strategies [93], can ensure a community's resilience to changing environmental conditions. Whether and how the hypervolume (n-dimensional geometrical shape) representing the functional composition of communities along key dimensions varies under climate change reflects whether and how those communities can adapt over time **(Figure 4D)**. Because architectural traits contribute to the communities' responses to key resource gradients and disturbances, we expect them to contribute to their responses to expected changes in climatic conditions and anthropogenic disturbance.

#### **Concluding remarks**

 Plant structure and development are important but overlooked drivers of individual performance and community assembly along environmental gradients. The architecture-based approach can significantly contribute to a better understanding of the variation between and within species, regardless of their growth form. This is a promising avenue for tackling major challenges in trait- based ecology such as the roles of time and space in species interactions [73, 94]. It is now standard 22 practice to revisit community ecology using a trait-based approach [95]. But such approaches are mostly based on organ-level functional traits, neglecting key aspects of plant architecture. By

 characterizing below- and above-ground plant architecture, we can take into account individual- level functional variations that better capture fundamental niche differences between neighboring individuals. A second challenge is to gain insight into the drivers and ecological implications of intraspecific trait variation [96]. The three-dimensional structure of a plant at a given time yields markers of different sources of inter- and intra-individual trait variation (e.g. ontogeny, biotic or abiotic environmental conditions). This makes it a powerful tool for disentangling cause and effect. A third and pivotal challenge concerns the shift to an integrated view of plant phenotype. Architectural traits can integrate multiple trait dimensions into a multilevel phenotypic network in a biologically coherent way. This requires further understanding of the functional space defined by architectural and organ-level traits, and of whole-plant metabolic or allometric rules (see **Outstanding questions**). We therefore encourage future research to identify the relevant architectural traits to measure, depending on ecological context, plant growth form and research question. We believe that lidar technology can make a significant contribution to this goal by providing metrics on the topological and geometrical properties of plant parts [35, 97, 98]. Finally, we argue that the dynamic, integrative view of plants proposed in the present work creates a much-needed bridge between trait-based, ontogenetic and demographic approaches.

#### **Acknowledgements**

 We thank all those involved in the fruitful discussions that led to this manuscript: C. Edelin, J. Prosperi, L. Gaume, O.Taugourdeau, E. Nicolini, N. Rowe, F. Anthelme, M. Millan, R. Pélissier, P. Couteron, T. Fourcaud and G. Vincent. We also thank the editor and three anonymous reviewers for 22 their insightful comments on the manuscript. This work has benefited from an `Investissements d'Avenir' grant managed by Agence Nationale de la Recherche (CEBA, ref. ANR-10-LABX- 25-01).

#### **Glossary**

 **Architectural model**: inherent growth strategy defined by a combination of four morphological characteristics observed on trunk and branches: growth pattern, branching pattern, morphological differentiation of axes and presence or absence of sexual organs. These models were first defined on tropical trees and then applied to temperate trees and to other growth forms (lianas, shrubs and herbs).

**Architectural traits:** traits describing plant structure at different organizational levels.

 **Architectural unit**: refers to the elementary structural organization of a given species described by the number, characteristics and spatial arrangement of axis categories. This concept can be used for both shoot and root branching systems.

 **Architecture-based approach:** refers to different ways of taking the plant structure into account depending on the research question. It includes consideration of intra- and inter-individual trait variations and screening for new phenotypic traits that cover overlooked dimensions of plant strategies.

 **Axis**: stem or root growing in a given direction and whose development can be expressed in different ways.

 **Axis category:** a type of root or stem characterized by its morpho-anatomical and functional properties within the tree architecture (e.g. a short shoot whose function is resource acquisition and reproduction). To identify an axis category, one must observe traits like growth direction, branching pattern, phyllotaxis and the presence or absence of sexual organs. There is a limited number of axis categories for a given species.

**Epicormic shoot**: a shoot growing from a long dormant meristem or bud.

 **Modularity**: the repetition of morphological elements (node/internode, annual shoot, axis, architectural unit) through growth, branching and reiteration.

 **Morphogenetic process**: a biological process by which a cell, tissue or organism develops its organization and shape. The different components of the plant (phytomer, growth unit, annual shoot, axis and architectural unit for shoots; axis and architectural unit for roots) are generated by three main morphogenetic processes : growth, branching and reiteration.

**Ontogeny**: the sequence of developmental phases characterizing a species' life cycle, independently

of the individual's chronological age.

**Phenology**: the study of the timing of recurrent biological events, its causes with regard to biotic

and abiotic forces, and the relations between phases in the same or different species.

**Phenotypic plasticity**: the ability of an organism to change its morphology, physiology and behavior

in response to environmental change.

**Phenotypic integration:** refers to the patterns of functional, developmental and/or genetic

correlations among different traits in a given organism.

 **Plant architecture**: describes the nature and relative arrangement of the different parts of a plant in space and time.

 **Reiteration**: a morphogenetic process generating the partial or total duplication of an architectural unit. This process may be part of the normal development sequence of the plant or may occur in response to stressful conditions. Reiteration can be expressed in the root system as well as the shoot system. The ability to reiterate has been well described in trees but is also expressed in herbs, lianas and shrubs.

**Topology**: the relative arrangement of morphological elements at different spatial scales.

## **Figure 1. Biased estimation of whole-plant functioning and performance when plant architecture**

**is overlooked.** 

 All research fronts in trait-based ecology (right column) would benefit from plant architecture analysis. This is theoretically illustrated here by the following: a weak correlation between organ- level and whole plant functions (A); the architecture-dependence of allocation (B), allometry patterns (C), the ontogenic trajectory of whole-plant metabolism (e.g., photosynthesis) (D), and reproductive effort (E). As a consequence, the way species' ecological differences are assessed can strongly influence quantification of the mechanisms underlying species coexistence (e.g. the niche overlap of species may vary with or without consideration of architecture) (F). Simplified trait values of hypothetical species are shown. Letters a, b, c and d represent different species.



**Phenotypic integration** 

**Determinants of allometric** and allocational scaling

Ontogenetic variation of traits

Understanding and prediction of plant performance

**Biotic and abiotic** interactions across time

#### **Box 1 – How to describe the variability in plant architecture?**

 The intra- and interspecific variability of plant architecture has three main causes (Figure I-A-C) [13- 15]. It can be described by morphological characteristics related to the key processes of growth, branching and reiteration (Figure I-D) at different levels of organization, depending on the question and the scale of the processes being studied. By combining different architectural traits, we can perform various types of categorizations: across species, different architectural types can be identified and used to classify the architectural diversity observed within a community or a clade 8 [13, 21]. For a given species, the variability of axis traits is used to define axis categories (Figure I-E). Their number indicates the degree of functional specialization of the axes. Figure I-F shows the importance of considering the axis categories and intra-individual gradients expressed during ontogeny when designing strategies for sampling functional traits**.** Considerable work remains to be done to better understand how axis differentiation influences intra- and interspecific variations in functional traits and whole-plant performance [70, 99].

 **Figure I- Variability in tree architecture**. *Sources of variability (A-C*): (A) Phenotypic plasticity: two walnut trees (*Juglans regia*) of the same age and the same population have very different architectures owing to differences in local plant density (adapted from Sabatier Sylvie, PhD thesis, University of Montpellier, 1999). (B) Intraspecific genotypic diversity: these forms of Cypress (fastigiate, intermediate and horizontal) correspond to different genotypes, making them heritable (adapted from [100]). (C) Interspecific diversity: a profile of a forest in French Guiana shows the wide diversity of architectures across the forest's vertical strata (adapted from Oldeman R.A.A, PhD thesis, University of Paris, 1978).

 *Methodological approach (D-F):* (D) Examples of architectural traits selected for describing and classifying the great architectural diversity of trees, shrubs and herbs in the genus *Euphorbia*; (figure modified from [87]); (E) Axis categories are defined according to distinguishing morphological criteria in the shoot and root systems of a theoretical species. Each axis category contributes to one or several functions; (F) Distribution of axis categories within the plant. The histograms show the proportion of leaf area borne by each axis category for each vertical layer. Leaves of the upper layer (the most often sampled in trait-based studies) are attached to axes of three different categories and should express differences in leaf trait values. Note that axis categories do not correspond to branching order. The discriminating criteria chosen to characterize axis categories are not exhaustive and could vary between species.



## (D) Examples of architectural traits



## (E) Axis categories



## (F) Whole-plant architecture



 $\mathbf{1}$ 

 $\overline{2}$ 

#### **Figure 2- The many dimensions of the architecture-based approach.**

 The three sides of the triangle below represent the three main axes of integration of the architecture-based approach in trait-based ecology. The architecture-based approach considers a 4 plant as a multilevel organism and explicitly includes the temporal dimension through ontogenetic and phenological processes. It makes it possible to account for inter and intra-individual trait variations, such as those in leaf size and shape, between ontogenetic stages (left side of triangle) or between axis categories (right side of triangle). Three axis categories are represented and colored in black, pink and orange on the right side of the triangle. The architecture-based approach also enables one to document phenotypic traits related to overlooked functions in functional strategies. The diversity of functions performed by plant structures is displayed below the triangle.



#### **Figure 3 – Disentangling age, ontogenetic stage and size across environments.**

 The ontogenetic trajectories of a theoretical tree as a function of its environment, here represented by different light conditions. The trajectory is represented in a two-dimensional space with the age 4 of the tree on the x-axis and its ontogenetic stage on the y-axis. Ontogenetic stage can be defined according to the degree of complexity of the plant structure in terms of branching, reiteration and flowering. The individual's architecture, age and dimensions constitute three complementary features. In full sun conditions, as with an isolated tree, the ontogenetic stages progress rapidly and the tree will reach maturity relatively quickly and remain relatively small. In intermediate shade, the ontogenetic stages progress more slowly, adulthood will be reached later and the adult tree will be larger. Under limiting conditions (closed canopy) the transition from one stage to another is slower and the full sequence is not necessarily achieved. Different trees of equivalent age may display widely different ontogenetic stages and growth dynamics. These different individual states can be characterized by a sequence of coded architectural traits, allowing the diversity of trajectories to be modelled at population or community level.



 $\mathbf{1}$ 

#### **Figure 4 - How do architectural traits contribute to population and community structure and**

#### **dynamics?**

 Depending on the scale and level of organization, different ecological processes and architectural traits are at play. (A) Population dynamics. Architectural traits may be related to reiteration (presence/absence; partial or total; number of total reiterated complexes), branching (presence/absence; maximum order), epicormics (presence/absence; orientation) or crown mortality (%). They can be combined to inform population dynamics, using Integral Projection Models (IPMs) that consider changes in individual trait values in a stage-structured population. Dot sizes are proportional to the number of individuals in the population. (B) Local plant-plant interactions and species coexistence and (C) functional structure of communities along environmental gradients. The architectural traits to be measured at these scales partially depend on the ecological context, biotic and abiotic filters particularly. For example, in the savannah ecosystem with its regular fire events and herbivore pressure, resprouting ability, index of cage architecture and height of fifth fork are key functional traits. (D) Community responses to climate change: the response of community functional composition to climate change is represented by the shift in trait hypervolumes (from yellow to blue) within a three-dimensional trait space involving architectural traits related, for example, to persistence strategies (types of reiteration in shoot and root systems, nature of axis categories) or resource foraging.

**B Community structure** 









## **Highlights**

- Plant functional traits are considered indicators of whole-plant functioning but are often measured only at the organ level and at a single developmental stage.
- The way plants build shoot and root axes determines their ability to explore and acquire resources in space and time in interaction with environmental conditions.
- The three-dimensional architecture of an individual plant at a given time reflects its ontogenetic stage and its environmental conditions, thus providing a powerful tool to disentangle the different sources of intraspecific trait variation.
- Plant architecture offers a hierarchical and modular framework for characterizing integrated plant phenotype and structuring plant trait networks.
- Plant architectural traits reflect key functional responses to abiotic and biotic conditions and should be considered when studying community assembly processes.

### **Outstanding questions**

- 
- What are the critical above- and belowground architectural traits that drive whole-plant performance across growth forms and ecological contexts? ● How do above- and belowground architectural traits coordinate with each other and with the major functional dimensions described so far? 7 • How does inter- and intraspecific variation in functional and architectural traits during ontogeny contribute to niche-based dynamics and community assembly? **•** How does the distribution of architectural traits vary across environmental gradients? 10 • How does architectural diversity influence the spatial and temporal dynamics of multi- trophic interactions (including plant-herbivore and plant-microbe interactions)? 12 • How does the diversity of architectures in plant communities affect ecosystem functioning?

## **References**

- 1. Garnier, E. *et al.* (2016) Plant Functional Diversity Organism traits, community structure, and ecosystem
- properties, Oxford University Press.
- 2. Violle, C. *et al.* (2007) Let the concept of trait be functional! *Oikos* 116 (5), 882-892.
- 3. Wright, I.J. *et al.* (2004) The worldwide leaf economics spectrum. *Nature* 428, 821-827.
- 4. Salguero-Gómez, R. *et al.* (2018) Delivering the promises of trait-based approaches to the needs of
- demographic approaches, and vice versa. *Funct. Ecol.* 32 (6), 1424-1435.
- 5. Paine, C.E.T. *et al.* (2015) Globally, functional traits are weak predictors of juvenile tree growth, and we
- do not know why. *J. Ecol.* 103 (4), 978-989.
- 6. Wright, S.J. *et al.* (2010) Functional traits and the growth mortality trade-off in tropical trees. *Ecology* 91 (12), 3664-3674.
- 7. Poorter, L. *et al.* (2008) Are functional traits good predictors of demographic rates ? Evidence from five neotropical forests. *Ecology* 89 (7), 1908-1920.
- 8. Shipley, B. *et al.* (2016) Reinforcing loose foundation stones in trait-based plant ecology. *Oecologia* 180 (4), 923-931.
- 9. Yang, J. *et al.* (2018) Why functional traits do not predict tree demographic rates. *Trends Ecol. Evol.* 33 (5), 326-336.
- 10. Levionnois, S. *et al.* (2023) Soil variation response is mediated by growth trajectories rather than
- functional traits in a widespread pioneer Neotropical tree. *Peer Community Journal* 3.
- 11. Corner, E.J.H. (1949) The Durian theory or the origin of the modern tree. *Ann. Bot.* 13, 367–414.
- 12. Barthélémy, D. and Caraglio, Y. (2007) Plant architecture: A dynamic, multilevel and comprehensive approach to plant form, structure and ontogeny. *Ann. Bot.* 99, 375-407.
- 13. Anest, A. *et al.* (2021) Evolving the structure: climatic and developmental constraints on the evolution of plant architecture. A case study in Euphorbia. *New Phytol.* 231 (3), 1278-1295.
- 14. Bruy, D. *et al.* (2018) Evolution of plant architecture, functional diversification and divergent evolution
- in the genus *Atractocarpus* (Rubiaceae) for New Caledonia. *Frontiers in Plant Science* 9 (1775).
- 15. Chomicki, G. *et al.* (2017) Evolution and ecology of plant architecture: integrating insights from the
- fossil record, extant morphology, developmental genetics and phylogenies. *Ann. Bot.* 120 (6), 855-891.
- 16. Hallé, F. and Oldeman, R.A.A. (1970) Essai sur l'architecture et la dynamique de croissance des arbres tropicaux, Masson.
- 17. Coudert, Y. *et al.* (2017) Multiple innovations underpinned branching form diversification in mosses.
- *New Phytol.* 215 (2), 840-850.
- 18. Labadie, M. *et al.* (2023) Spatio-temporal analysis of strawberry architecture: insights into the control of branching and inflorescence complexity. *J. Exp. Bot.* 74 (12), 3595-3612.
- 19. Atger, C. and Edelin, C. (1994) Premières données sur l'architecture comparée des systèmes racinaires et caulinaires. *Canadian Journal of Botany* 72 (7), 963-975.
- 20. Torres, C.D. *et al.* (2022) Assessing coordinated intra-specific variation in root/shoot traits in two
- herbaceous species based on architecture and ontogeny. *Folia Geobotanica* 57 (3), 167-180.
- 21. Salmon, C. *et al.* (2023) Architectural traits underlie growth form diversity and polycarpic versus
- monocarpic life histories in Cerberiopsis (Apocynaceae). *Bot. J. Linn. Soc.* 202 (4), 510-528.
- 22. de Haldat du Lys, A. *et al.* (2023) If self-shading is so bad, why is there so much? Short shoots reconcile costs and benefits. *New Phytol.* 237 (5), 1684-1695.
- 23. Poorter, H. *et al.* (2014) Trait correlation networks: a whole-plant perspective on the recently criticized leaf economic spectrum. *New Phytol.* 201 (2), 378-382.
- 24. He, N. *et al.* (2020) Plant trait networks: Improved resolution of the dimensionality of adaptation.
- *Trends Ecol. Evol.* 35 (10), 908-918.
- 25. Kraft, N.J.B. *et al.* (2015) Plant functional traits and the multidimensional nature of species coexistence.
- *Proceedings of the National Academy of Sciences* 112 (3), 797-802.
- 26. Jucker, T. *et al.* (2022) Tallo: A global tree allometry and crown architecture database. *Global Change*
- *Biol.* 28 (17), 5254-5268.
- 27. Williams, L.J. *et al.* (2017) Spatial complementarity in tree crowns explains overyielding in species
- mixtures. *Nature Ecology & Evolution* 1, 0063.
- 28. Keddy, P.A. (1992) A pragmatic approach to functional ecology. *Funct. Ecol.* 6 (6), 621-626.
- 29. Ottaviani, G. *et al.* (2020) The neglected belowground dimension of plant dominance. *Trends Ecol. Evol.*  35 (9), 763-766.
- 30. Pérez-Harguindeguy, N. *et al.* (2013) New handbook for standardised measurement of plant functional traits worldwide. *Aust. J. Bot.* 61, 167-234.
- 31. Iversen, C.M. *et al.* (2017) A global fine-root ecology database to address below-ground challenges in plant ecology. *New Phytol.* 215 (1), 15-26.
- 32. Yang, X.-D. *et al.* (2015) Tree architecture varies with forest succession in evergreen broad-leaved
- forests in Eastern China. *Trees* 29 (1), 43-57.
- 33. Poorter, L. *et al.* (2003) Architecture of 53 rain forest tree species differing in adult stature and shade tolerance. *Ecology* 84 (3), 602-608.
- 34. Lynch, J.P. (2022) Harnessing root architecture to address global challenges. *The Plant Journal* 109 (2), 415-431.
- 35. Lines, E.R. *et al.* (2022) The shape of trees: Reimagining forest ecology in three dimensions with remote
- sensing. *J. Ecol.* 110 (8), 1730-1745.
- 36. Banin, L. *et al.* (2012) What controls tropical forest architecture? Testing environmental, structural and
- floristic drivers. *Global Ecol. Biogeogr.* 21 (12), 1179-1190.
- 37. Pearcy, R.W. *et al.* (2005) Crown architecture in sun and shade environments : assessing function and trade-offs with a three-dimensional simulation model. *New Phytol.* 166, 791-800.
- 38. Jones, C. (1999) An essay on juvenility, phase change, and heteroblasty in seed plants. *Int. J. Plant Sci.*  160 (S6), S105-S111.
- 39. Klimešová, J. *et al.* (2021) Incorporating clonality into the plant ecology research agenda. *Trends Plant Sci.* 26 (12), 1236-1247.
- 40. Huber, H. *et al.* (1999) Spatial structure of stoloniferous herbs: an interplay between structural blue-
- print, ontogeny and phenotypic plasticity. *Plant Ecol.* 141 (1-2), 107-115.
- 41. Harder, L.D. *et al.*, The role of structural and life-history tradeoffs in plant architecture: a model study of
- Protea lepidocarpodendron (Proteaceae), 5 th International Workshop on Structural-Fonctional Plant Models, Napier, New Zealand, 2007.
- 42. Klimešová, J. *et al.* (2018) Belowground plant functional ecology: Towards an integrated perspective.
- *Funct. Ecol.* 32 (9), 2115-2126.
- 43. Chiminazzo, M.A. *et al.* (2023) Why woody plant modularity through time and space must be integrated in fire research? *AoB PLANTS* 15 (3).
- 44. Charles-Dominique, T. *et al.* (2017) The architectural design of trees protects them against large
- herbivores. *Funct. Ecol.* 31 (9), 1710-1717.
- 45. Granados Mendoza, C. *et al.* (2014) Bouldering: an alternative strategy to long-vertical climbing in root-climbing hortensias. *Journal of The Royal Society Interface* 11 (99), 20140611.
- 46. Leigh, A. *et al.* (2011) Structural and hydraulic correlates of heterophylly in Ginkgo biloba. *New Phytol.*  189 (2), 459-470.
- 47. Lauri, P.-E. and Kelner, J.-J. (2011) Shoot type demography and dry matter partitioning: A morphometric
- approach in apple (Malus ×domestica). *Canadian Journal of Botany* 79, 1270-1273.
- 48. Pearcy, R.W. *et al.* (2004) A functional analysis of the crown architecture of tropical forest *Psychotria*
- species : do species vary in light capture efficiency and consequently in carbon gain and growth ? *Oecologia*  139, 163-177.
- 49. Laurans, M. and Vincent, G. (2016) Are inter- and intraspecific variations of sapling crown traits
- consistent with a strategy promoting light capture in tropical moist forest? *Ann. Bot.* 118 (5), 983-996.
- 50. Ishii, H.T. *et al.* (2007) Physiological and ecological implications of adaptive reiteration as a mechanism
- for crown maintenance and longevity. *Tree physiology* 27 (3), 455-462.
- 51. Nicolini, E. *et al.* (2003) Epicormic Branches: a Growth Indicator for the Tropical Forest Tree, Dicorynia
- guianensis Amshoff (Caesalpiniaceae). *Ann. Bot.* 92 (1), 97-105.
- 52. Meier, A.R. *et al.* (2012) Epicormic buds in trees: a review of bud establishment, development and
- dormancy release. *Tree physiology* 32 (5), 565-584.
- 53. Klimešová, J. *et al.* (2018) Horizontal growth: An overlooked dimension in plant trait space. *Perspect.*
- *Plant Ecol. Evol. Syst.* 32, 18-21.
- 54. Poorter, L. *et al.* (2010) Resprouting as a persistence strategy of tropical forest trees: Relations with
- carbohydrate storage and shade tolerance. *Ecology* 91 (9), 2613-2627.
- 55. Klimešová, J. *et al.* (2019) Handbook of standardized protocols for collecting plant modularity traits.
- *Perspect. Plant Ecol. Evol. Syst.* 40, 125485.
- 56. Taugourdeau, O. *et al.* (2012) Retrospective analysis of tree architecture in silver fir (Abies alba Mill.):
- ontogenetic trends and responses to environmental variability. *Annals of Forest Science* 69 (6), 713-721.
- 57. Taugourdeau, O. *et al.* (2011) Deciphering the developmental plasticity of walnut saplings in relation to climatic factors and light environment. *J. Exp. Bot.* 62 (15), 5283-5296.
- 58. Wang, Y. *et al.* (2012) Variation of maximum tree height and annual shoot growth of Smith Fir at various
- elevations in the Sygera Mountains, Southeastern Tibetan Plateau. *PLOS ONE* 7 (3), e31725.
- 59. Dolezal, J. *et al.* (2020) Alpine plant growth and reproduction dynamics in a warmer world. *New Phytol.*  228 (4), 1295-1305.
- 60. Schweingruber, F.H. and Poschlod, P. (2005) Growth rings in herbs and shrubs: life span, age
- determination and stem anatomy, Swiss Federal Research Institute WSL Birmensdorf, Switzerland.
- 61. Baudoux, C. *et al.* (2023) Can the competition dynamics of non-native invaders be reconstructed to
- reveal historical impact? *Biol. Invasions* (in press).
- 62. Zhu, L. *et al.* (2017) Trait choice profoundly affected the ecological conclusions drawn from functional
- diversity measures. *Scientific Reports* 7 (1), 3643.
- 63. Streit, R.P. and Bellwood, D.R. (2023) To harness traits for ecology, let's abandon 'functionality'. *Trends Ecol. Evol.* 38 (5), 402-411.
- 64. Wigley, B.J. *et al.* (2020) A handbook for the standardised sampling of plant functional traits in
- disturbance-prone ecosystems, with a focus on open ecosystems. *Aust. J. Bot.* 68 (8), 473-531.
- 65. Dwyer, J.M. and Laughlin, D.C. (2017) Constraints on trait combinations explain climatic drivers of
- biodiversity: the importance of trait covariance in community assembly. *Ecol. Lett.* 20 (7), 872-882.
- 66. Zhou, J. *et al.* (2022) Global analysis of trait–trait relationships within and between species. *New Phytol.*
- 233 (4), 1643-1656.
- 67. He, D. *et al.* (2021) The importance of intraspecific trait variability in promoting functional niche
- dimensionality. *Ecography* 44 (3), 380-390.
- 68. Sack, L. and Buckley, T.N. (2019) Trait multi-functionality in plant stress response. *Integr. Comp. Biol.* 60 (1), 98-112.
- 69. Garnier, E. *et al.* (2018) Inter- and intra-specific trait shifts among sites differing in drought conditions at
- the north western edge of the Mediterranean Region. *Flora: Morphology, Distribution, Functional Ecology of Plants* 254, 147-160.
- 70. Edwards, E.J. *et al.* (2014) Leaf life span and the leaf economic spectrum in the context of whole plant
- architecture. *J. Ecol.* 102 (2), 328-336.
- 71. Messier, J. *et al.* (2017) Interspecific integration of trait dimensions at local scales: the plant phenotype as an integrated network. *J. Ecol.* 105 (6), 1775-1790.
- 72. Lauri, P.-É. (2018) Corner's rules as a framework for plant morphology, architecture and functioning issues and steps forward. *New Phytol.* 221, 1679-1684.
- 
- 73. Cope, O.L. *et al.* (2022) The role of timing in intraspecific trait ecology. *Trends Ecol. Evol.* 37 (11), 997- 1005.
- 74. Petrone-Mendoza, E. *et al.* (2023) Plant morpho evo-devo. *Trends Plant Sci.* 28 (11), 1257-1276.
- 75. Roggy, J.C. *et al.* (2005) Links between tree structure, and functional leaf traits in the tropical forest tree *Dicorynia guianensis* Amshoff (Caesalpiniaceae). *Annals of Forest Science* 62 (6), 553-564.
- 76. Fortunel, C. *et al.* (2020) Disentangling the effects of environment and ontogeny on tree functional
- dimensions for congeneric species in tropical forests. *New Phytol.* 226 (2), 385-395.
- 77. Fajardo, A. and Siefert, A. (2016) Phenological variation of leaf functional traits within species.
- *Oecologia* 180 (4), 951-959.
- 78. Charles-Dominique, T. *et al.* (2012) Architectural strategies of Rhamnus cathartica (Rhamnaceae) in
- relation to canopy openness. *Botany* 90 (10), 976-989.
- 79. Wright, S.D. and Mc Connaughay, K.D.M. (2002) Interpreting phenotypic plasticity : the importance of
- ontogeny. *Plants species Biology* 17, 119-131.
- 80. Shipley, B. (2009) From Plant Traits to Vegetation Structure: Chance and Selection in the Assembly of
- Ecological Communities, Cambridge University Press.
- 81. Merow, C. *et al.* (2014) Advancing population ecology with integral projection models: a practical guide.
- *Methods in Ecology and Evolution* 5 (2), 99-110.
- 82. Charles-Dominique, T. *et al.* (2014) Using intra-individual variation in shrub architecture to explain
- population cover. *Oikos* 124 (6), 707-716.
- 83. Van Sickle, J. (1977) Analysis of a distributed-parameter population model based on physiological age. *J.*
- *Theor. Biol.* 64 (3), 571-586.
- 84. Levine, J.M. and HilleRisLambers, J. (2009) The importance of niches for the maintenance of species
- diversity. *Nature* 461 (7261), 254-257.
- 85. Trogisch, S. *et al.* (2021) The significance of tree-tree interactions for forest ecosystem functioning. *Basic Appl. Ecol.* 55, 33-52.
- 86. Canham, C.D. *et al.* (2006) Neighborhood analyses of canopy tree competition along environmental gradients in New England forests. *Ecol. Appl.* 16 (2), 550-554.
- 87. Ford, D. (2014) The dynamic relationship between plant architecture and competition. *Frontiers in Plant Science* 5.
- 88. Zambrano, J. *et al.* (2019) Tree crown overlap improves predictions of the functional neighbourhood
- effects on tree survival and growth. *J. Ecol.* 107 (2), 887-900.
- 89. Chalmandrier, L. *et al.* (2021) Linking functional traits and demography to model species-rich
- communities. *Nature Communications* 12 (1).
- 90. Churski, M. *et al.* (2022) Herbivore-induced branching increases sapling survival in temperate forest canopy gaps. *J. Ecol.* 110 (6), 1390-1402.
- 91. Millet, J. *et al.* (1998) Relationship between architecture and successional status of trees in the temperate deciduous forest *Ecoscience* 6 (2), 187-203.
- 92. Violle, C. *et al.* (2014) The emergence and promise of functional biogeography. *Proceedings of the*
- *National Academy of Sciences* 111 (38), 13690-13696.
- 93. Leitão, R.P. *et al.* (2016) Rare species contribute disproportionately to the functional structure of
- species assemblages. *Proceedings of the Royal Society B: Biological Sciences* 283 (1828), 20160084.
- 94. Munné-Bosch, S. (2022) Spatiotemporal limitations in plant biology research. *Trends Plant Sci.* 27 (4), 346-354.
- 95. Mc Gill, B.J. *et al.* (2006) Rebuilding community ecology from functional traits. *Trends in Ecology and Evolution* 21 (4), 178-185.
- 96. Trends in Ecology & EvolutionChacón-Labella, J. *et al.* (2022) How to improve scaling from traits to ecosystem processes. *Trends Ecol. Evol.* 38 (3), 228-237.
- 97. Verbeeck, H. *et al.* (2019) Time for a Plant Structural Economics Spectrum. *Frontiers in Forests and Global Change* 2 (43).
- 98. Martin-Ducup, O. *et al.* (2020) Terrestrial laser scanning reveals convergence of tree architecture with
- increasingly dominant crown canopy position. *Funct. Ecol.* 34 (12), 2442-2452.
- 99. Dang-Le, A.T. *et al.* (2013) Ontogenetic variations in leaf morphology of the tropical rain forest species
- Dipterocarpus alatus Roxb. ex G. Don. *Trees* 27 (3), 773-786.
- 100. Barthélémy, D. *et al.* (1999) Biologie, croissance et développement. In Le cyprès : guide pratique
- (Teissier Du Cros, E. et al. eds), pp. 27-34, Studio Leonardo.
-