

Analyzing HEI10 Dosage Effect on CO formation in an allopolyploid species: Camelina sativa

Marie Casado, Greta Sandmann, Patrick Grillot, Eric Jenczewski

► To cite this version:

Marie Casado, Greta Sandmann, Patrick Grillot, Eric Jenczewski. Analyzing HEI10 Dosage Effect on CO formation in an allopolyploid species: Camelina sativa. Plant and Animal Genome Congress PAG31, Jan 2024, San diego (Californie), France. . hal-04507641

HAL Id: hal-04507641 https://hal.inrae.fr/hal-04507641v1

Submitted on 16 Mar 2024 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Analyzing HEI10 Dosage Effect on CO formation in an allopolyploid species: Camelina sativa

Marie Casado¹, Greta Sandmann¹, Julie Guerin¹, Aurélie Chambon¹, Patrick Grillot¹ and Eric Jenczewski¹



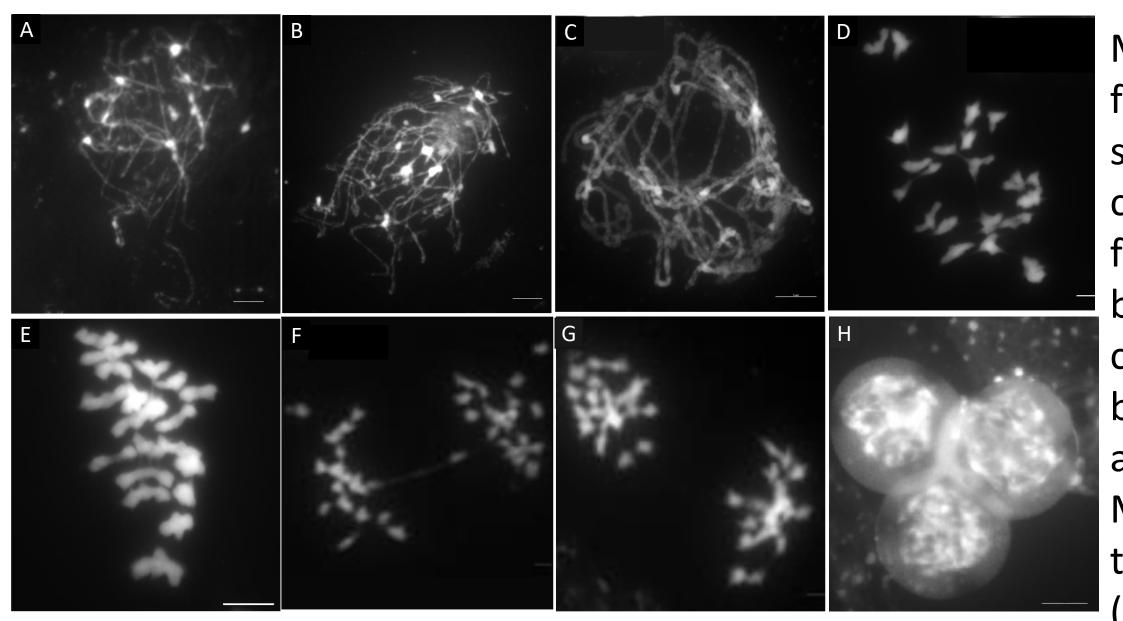
Abstract

Meiotic recombination is a fundamental process for all sexual eukaryotes; it is also crucial for plant breeding as it enables genetic material to be reshuffled between individuals (and species) via the formation of crossovers (COs). To date, most progress in deciphering the mechanisms of CO formation has been achieved through the analysis of knockout (KO) mutants in diploid plants such as Arabidopsis thaliana. Our working hypothesis is that the study of meiosis in allopolyploid species -which generally contain each gene in multiple copies- could provide additional information, in particular by making it possible to study the effect of gene dosage. In this work, we have evaluated the extent to which manipulating the number of functional copies encoding HEI10 (Enhancer of Cell Invasion n°10), a protein involved in the main COs pathway¹, could modify COs number in Camelina sativa, a hexaploid (2n=40) oleaginous species carrying 3 CsaHEI10 is an excellent candidate for this study, as it has been shown that the addition of extra copies of. HEI10 in *A.thaliana* leads to a significant increase in COs number, suggesting a dosage effect^{2,3}.

As meiosis has never been described in C.sativa, we first characterized several key hallmarks of male meiosis in this species using (immuno-)cytology approaches. Then, we modulated the number of functional HEI10 copies downwards by generating (from 0 to 6) CRISPR-Cas9 KO alleles, and upwards by adding extra HEI10 copies from C.sativa and A.thaliana. Analyzing the meiotic behavior in these different genetic backgrounds - which show varying numbers of functional HEI10 copies - provides further insight into the level of HEI10 required to maintain obligatory CO, and the possibility of compensation between copies.

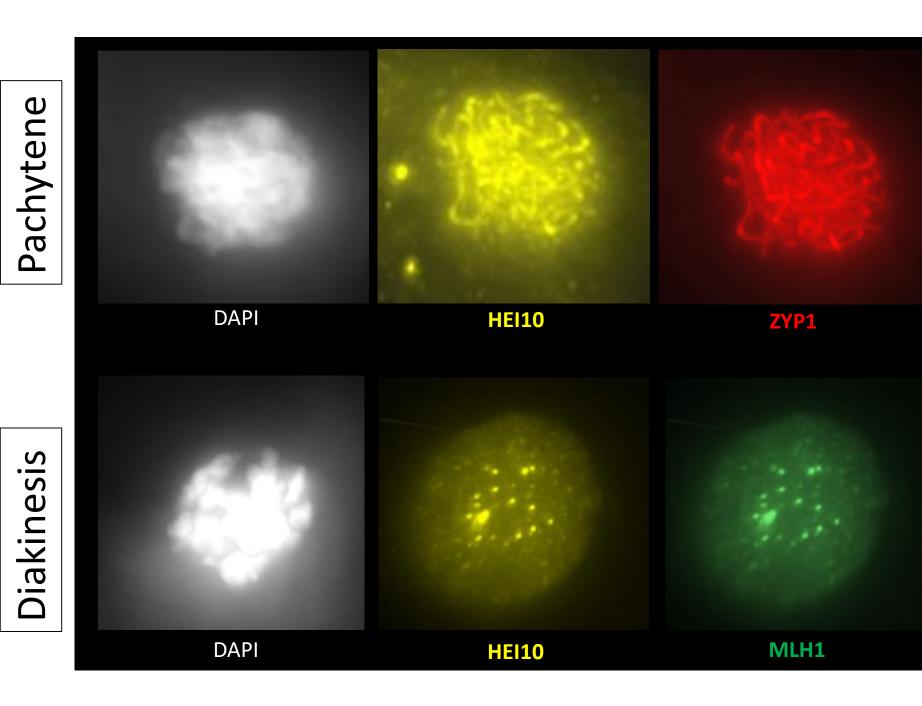
C.sativa shows a diploid-like meiotic behavior with about 1 class I CO per bivalent

In *C.sativa*, HEI10 shows a specific dynamics modelled as **coarsening**



A: Leptotene, B: Zygotene, C : Pachytene, D: Diakinesis, E: Metaphase I, F: Anaphase I, G: Telophase, H : microspore tetrades, scale = $5 \mu m$

Male meiosis in *C.sativa* follows the expected sequence of events⁴ : chromosome axes are formed (A,B) and joined by the synaptonemal complex (C); 20 bivalents are observed at metaphase I (E). Meiosis II ends up with a tetrad of microspores (H).

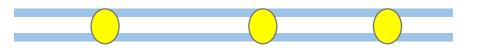


In the early stages, HEI10 shows a multitude of small foci, some being brighter than others. At pachytene, all foci colocalize with the synaptonemal complex (ZYP1).

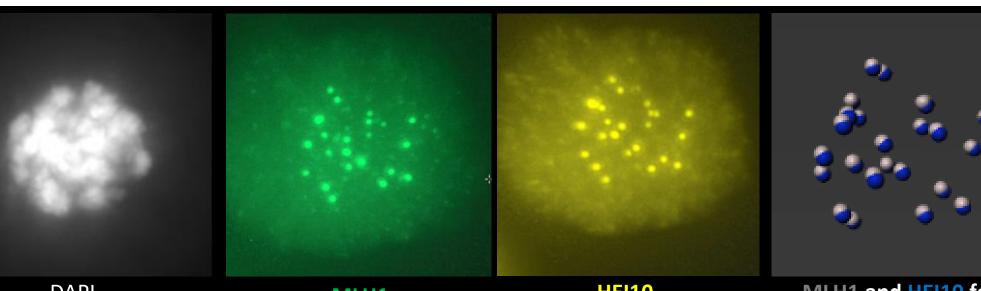
From early to late pachytene, the number of HEI10 foci decreases drastically, to form a small number of large foci. This dynamic could be modeled by a diffusion-mediated **coarsening model**^{3,5} in which larger HEI10 foci grow at the expense of smaller ones.



At diakinesis, HEI10 foci perfectly colocalized with MLH1 foci, spotting where class I COs are formed.



3D immunocytological analyses of MLH1 and HEI10 shows that *C.sativa var* Celine only produces **20,8**+/-**2,2** class I CO/cell.



HEI10 shows a dosage effect on the number of COs in C.sativa

Decreasing *CsaHEI10* copy number

Using different combinations of WT and KO mutant alleles, we show that a single functionnal allele of HEI10 is not sufficient to safeguard the obligatory CO.

MLH1 and HEI10 foci

 \rightarrow No more

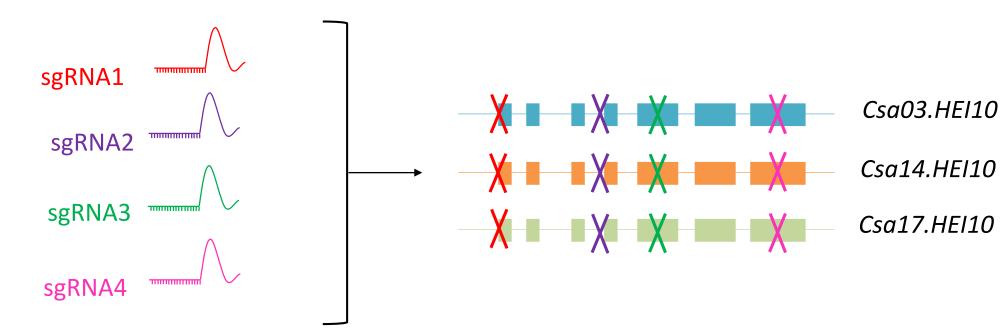
class I COs

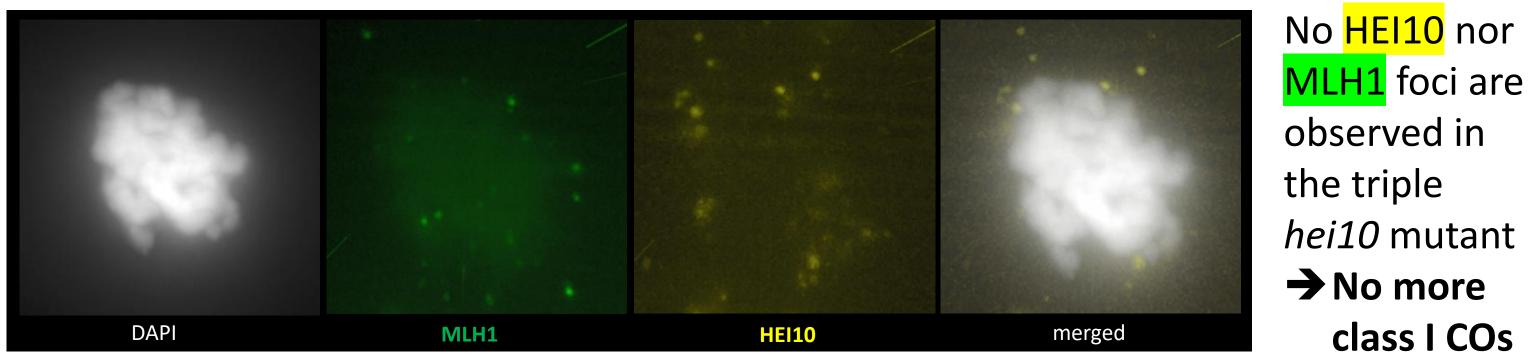
Class II

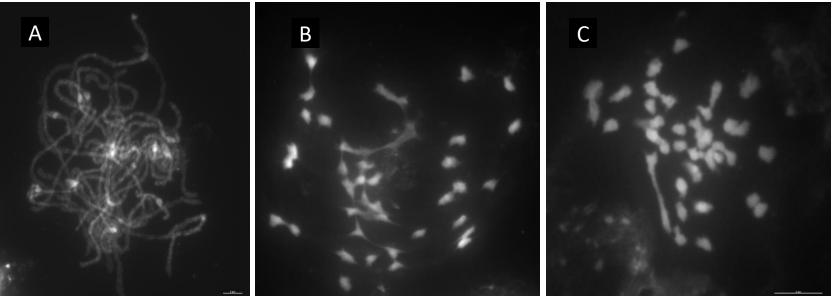
COs

Generating a series of *hei10 null* mutant alleles in *C.sativa*

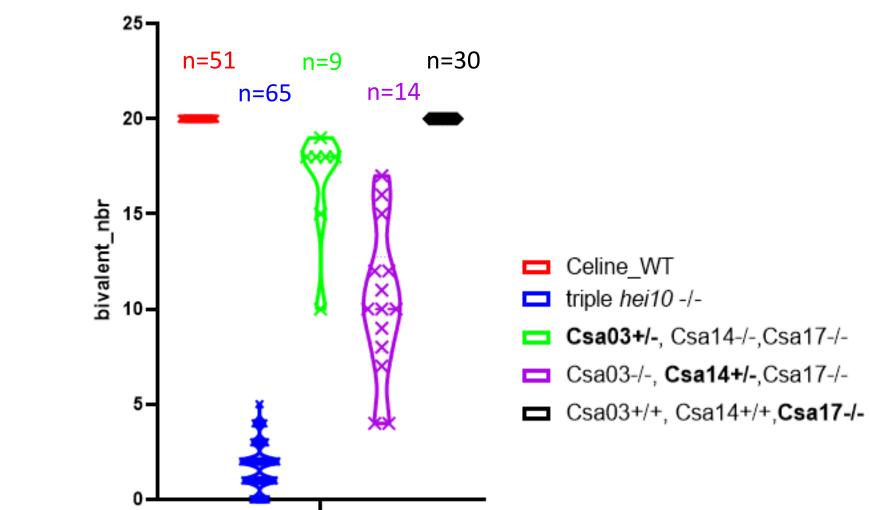
The three genes encoding HEI10 in *C.sativa* var Celine, were succesfully knockedout using the CRISPR/Cas9 strategy.

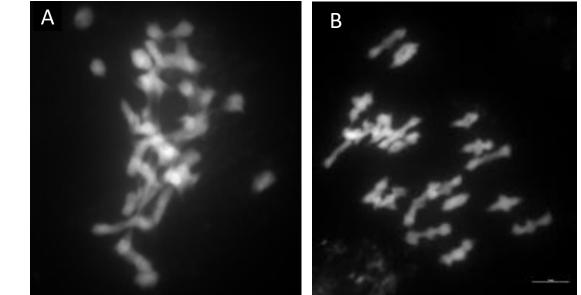




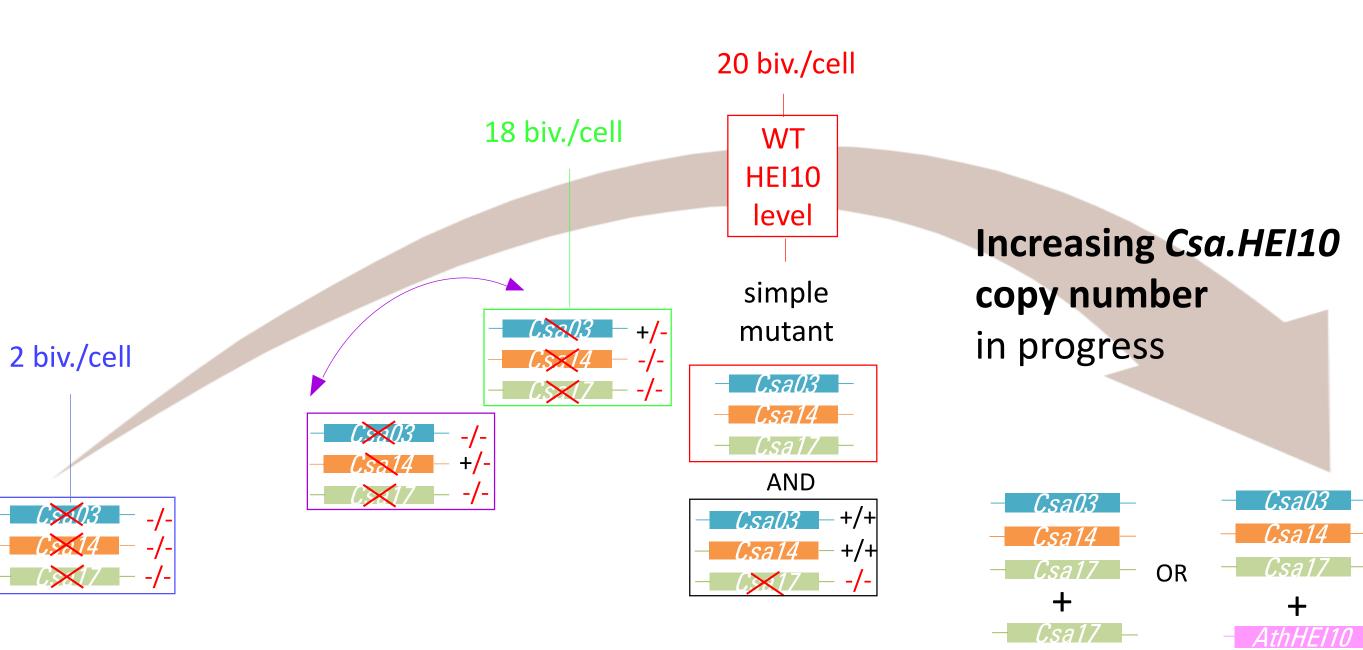


The triple *hei10* mutant shows a huge reduction in bivalent **number**, down to 2 bivalents per cell on average





A : Metaphase I of Csa03+/-, Csa14-/-, Csa17-/-B : Metaphase I of Csa03+/+, Csa14+/+, Csa17-/-



A: Pachytene, B: Diakinesis, C: Metaphase I

TAKE HOME MESSAGES

□ WT *C.sativa* shows about 1 class I CO per bivalent

- As in other species, HEI10 is required for the Class I CO pathway
- In contrast to A. thaliana, one single functional allele of HEI10 is not sufficient to safeguard the obligatory CO

• One missing copy (out of 3) does not affect the number of bivalents

UHypothesis to test next :

- How many functional HEI10 copies are required to restore the mandatory CO?
- Does the addition of extra HEI10 copies enhance COs numbers ?
- Is this enhancement proportional to the number of HEI10 copies added or will it be very small or null compared to WT?

• Do the 3 copies contribute differently to COs formation ?

Bibliography :

- 1- Chelysheva L, Vezon D, Chambon A, Gendrot G, Pereira L, Lemhemdi A, Vrielynck N, Le Guin S, Novatchkova M, Grelon M. 2012. 2- Ziolkowski PA, Underwood CJ, Lambing C, Martinez-Garcia M, Lawrence EJ, Ziolkowska L, Griffin C, Choi K, Franklin FCH, Martienssen RA, et al. 2017.
- 3-Morgan C, Fozard JA, Hartley M, Henderson IR, Bomblies K, Howard M. 2021.
- 4- Mercier R, Mézard C, Jenczewski E, Macaisne N, Grelon M. 2015.
- 5-Durand S, Lian Q, Jing J, Ernst M, Grelon M, Zwicker D, Mercier R. 2022.