

# Supplementary Information

## Rapid detection and identification of vancomycin sensitive bacteria using an electrochemical apta-sensor

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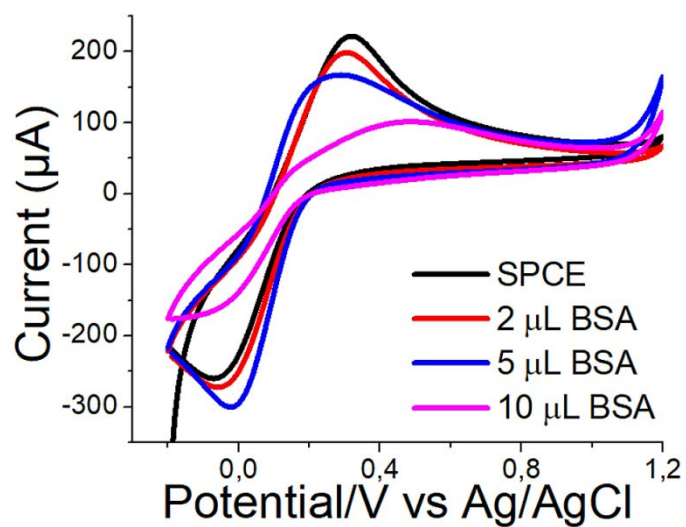
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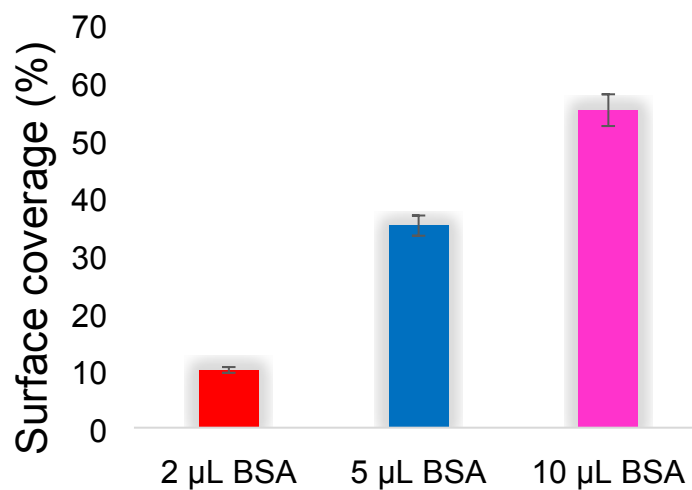
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a)

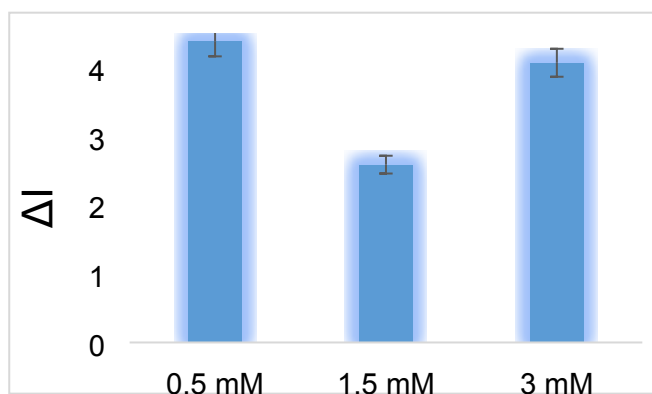


b)

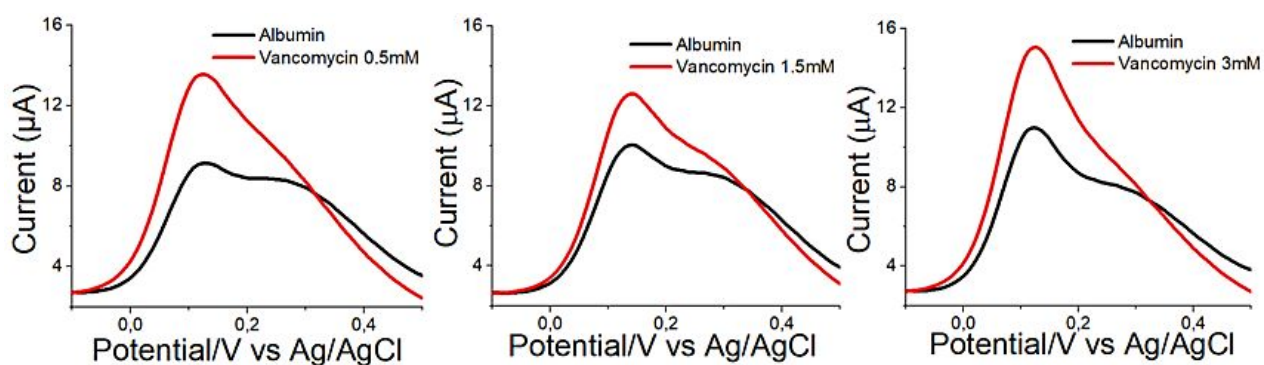


**Figure S1. BSA immobilization.** a) CV obtained using electrode incubated with different volumes of 2 mg/mL BSA. b) Surface coverage (%) versus different volumes of BSA used for functionalization.

a)

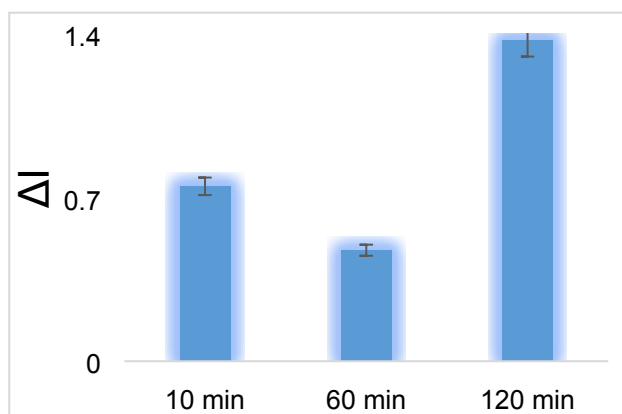


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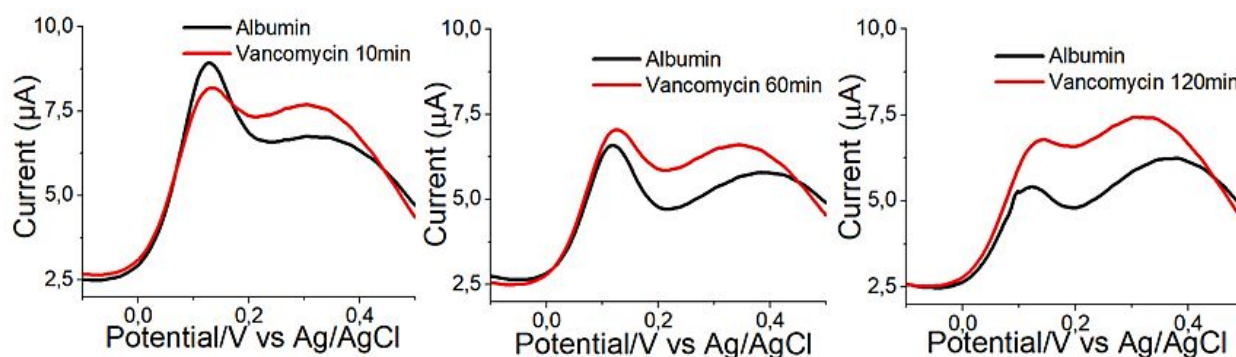


**Figure S2. Vancomycin immobilization: optimization of concentration.** a) The change in current intensity obtained after electrode incubation with different vancomycin concentrations. b) DPV obtained for different vancomycin concentrations. DPV parameters: potential ranged from -0.2 V to 0.5V, pulse amplitude 0.02 V.

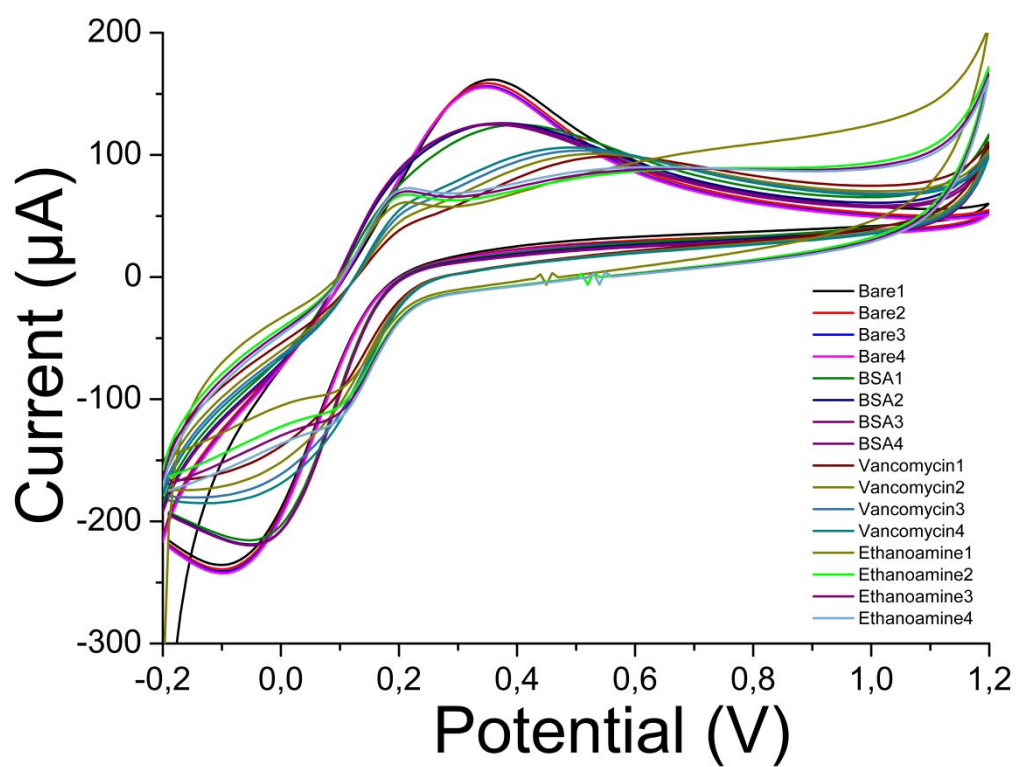
a)



b)



**Figure S3. Vancomycin immobilization: optimization of incubation time.** a) DPV current intensity versus different times of vancomycin incubation onto the working electrode. b) DPV obtained after the addition of vancomycin for different times of incubations.



**Figure S4. Electrode stability tests** using CV at scan rate of 50 mV/s in  $\text{Fe}^{2+/3+}$  redox couple in 0.1 KCl as supporting electrolyte.

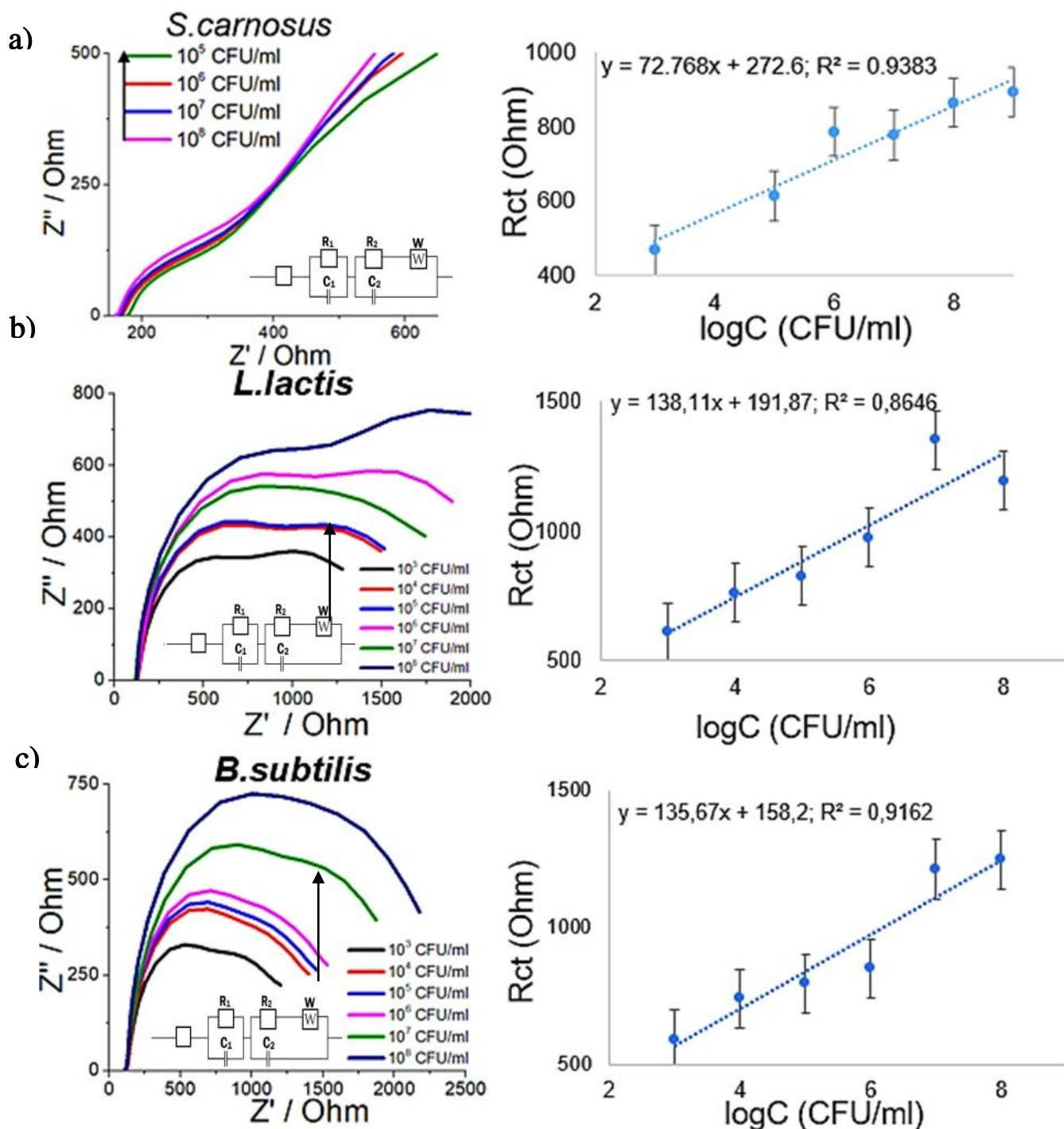
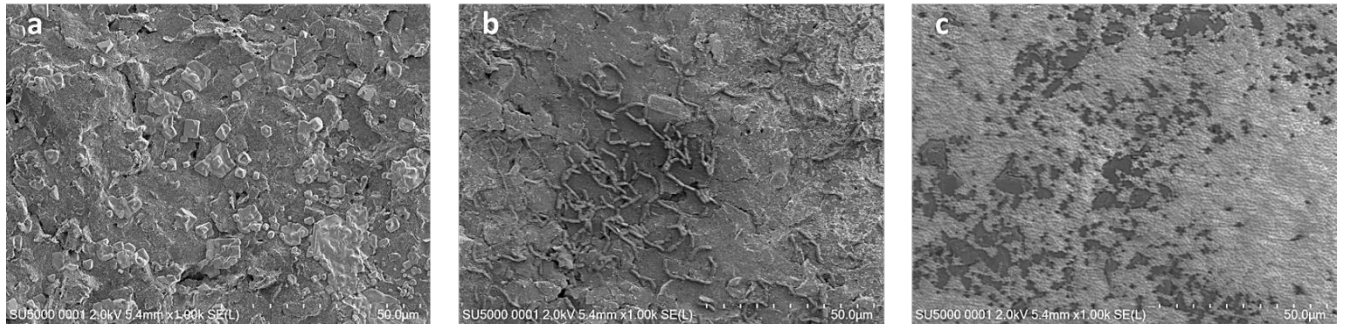
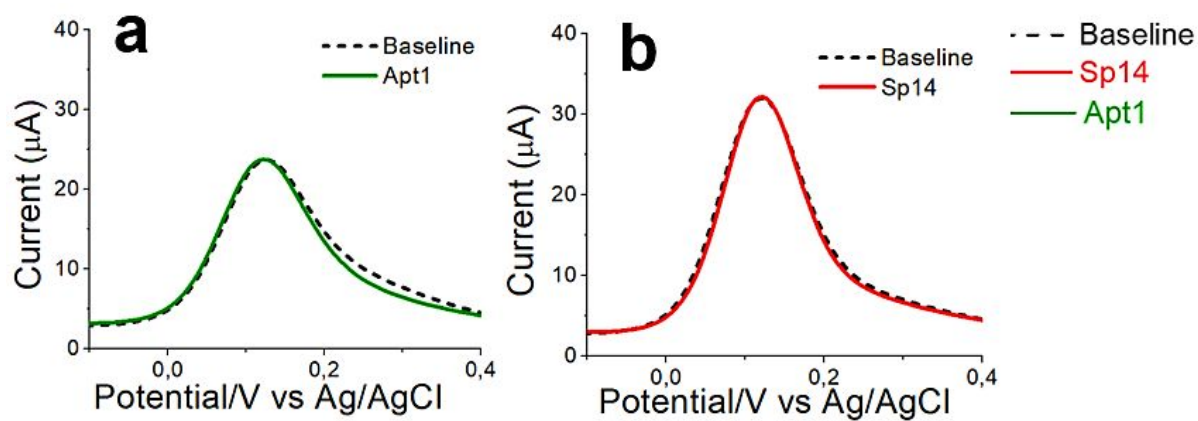


Figure S5. Electrochemical impedance spectroscopy measurements. a) Nyquist plot of impedance spectra of the vancomycin-based biosensor obtained from the increasing amount of *S. carnosus*, gram-positive bacteria in PBS. The calibration plot showing the change of charge transfer resistance,  $R_{ct}$ , as a function of the different concentrations of *S. carnosus*; b) and c) Nyquist plots of impedance spectra of *L. lactis* and *B. subtilis*, respectively.

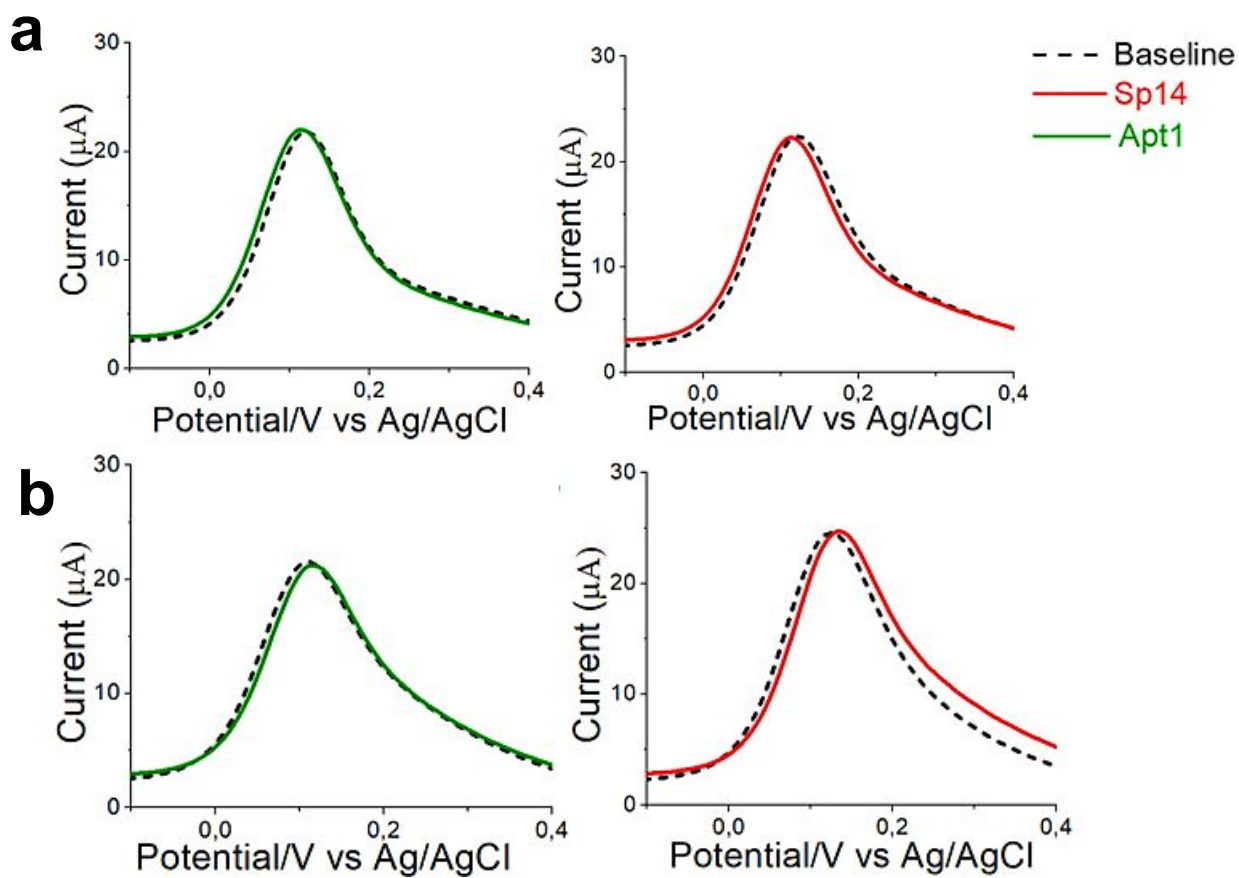


**S6. SEM images of electrodes carrying vancomycin after incubation with *E. coli* (a), *B. cereus* (b) and *S. carnosus* (c) showing the nonadherence of *E. coli* in contrast to two Gram-positive bacteria.**





**Figure S7. Aptamer background signal.** a) DPV obtained after biosensor incubation with 10  $\mu\text{M}$  Apt1; b) DPV obtained after biosensor incubation with 1  $\mu\text{M}$  specific aptamer, Sp14.



**Figure S8. Aptamer background signal in real samples.** a) DPV obtained after biosensor incubation with 10  $\mu\text{M}$  Apt1 and 1  $\mu\text{M}$  specific aptamer, Sp14 in milk; b) DPV obtained after biosensor incubation with 10  $\mu\text{M}$  Apt1 and 1  $\mu\text{M}$  specific aptamer, Sp14 in serum.

**Table S1.** Fitting values of the equivalent circuit elements for Gram positive bacteria detection by EIS Analyzer software. **R1** is the internal resistance of the electrolyte solution, **R2** is the resistance due to electrochemical reactions, **R3** is the charge transfer resistance, and **W1**, the Warburg resistance.

	logC of bacteria	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	W
<i>B. subtilis</i>	3	127.05	588.86	810.65	/
	4	127.23	738.97	919.9	/
	5	120.42	794.61	881.12	/
	6	120.88	848.69	894.83	/
	7	121.72	1210	934.4	/
	8	124.85	1245.1	1172.8	/
<i>L. lactis</i>	3	125.42	608.5	1081.4	/
	4	125.06	760.43	1254.8	/
	5	126.7	825.24	1225.8	/
	6	118.28	973.71	1496.6	/
	7	124.56	1347.9	1743.3	/
	8	125,00	1193.1	1768.6	/
<i>B. cereus</i>	3	156.05	2123.5	231.7	1.31E <sup>-19</sup>
	4	150.01	2153.6	284.22	5.14E <sup>-19</sup>
	5	151.3	2362.5	235.98	8.95E <sup>-19</sup>
	6	159.86	2518.2	265.27	1.45E <sup>-18</sup>
	7	160	2597.9	231.78	2.11E <sup>-18</sup>
<i>S. carnosus</i>	3	178.89	1628,5	155,78	5.7484E <sup>-23</sup>
	4	168.57	1932,6	186,91	1.3653E <sup>-22</sup>
	5	166.36	2204,4	188,73	2.94E <sup>-20</sup>
	6	175.39	2627,6	230,18	1.4154E <sup>-22</sup>
	7	162.13	2695,6	221,41	3.0636E <sup>-22</sup>
<i>S. aureus</i>	3	124.41	238.51	367.82	324.87
	4	114.94	470.99	341.96	340.02
	5	122.85	423.1	602.99	359.95
	6	122.03	585.35	465.28	357.46
	7	121.49	585.35	499.46	367.2
	8	123.18	702.08	580.21	376.38