



HAL
open science

“Ectomycorrhizal exploration type” could be a functional trait explaining the spatial distribution of tree symbiotic fungi as a function of forest humus forms

Fadwa Khalfallah, Lucie Bon, Mohamed El Mazlouzi, Mark R. Bakker, Nicolas Fanin, Richard Bellanger, Bernier Frédéric, Adinda de Schrijver, Catherine Ducatillon, Fotelli M.N., et al.

► To cite this version:

Fadwa Khalfallah, Lucie Bon, Mohamed El Mazlouzi, Mark R. Bakker, Nicolas Fanin, et al.. “Ectomycorrhizal exploration type” could be a functional trait explaining the spatial distribution of tree symbiotic fungi as a function of forest humus forms. *Mycorrhiza*, 2024, 34, pp.203-216. 10.1007/s00572-024-01146-8. hal-04592027

HAL Id: hal-04592027

<https://hal.inrae.fr/hal-04592027v1>

Submitted on 17 Jan 2025

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

1 **“Ectomycorrhizal exploration type” could be a functional trait explaining the spatial**
2 **distribution of tree symbiotic fungi as a function of forest humus forms**

3

4 Khalfallah F.^{1,2}, Bon L.³, El Mazlouzi M.^{3,15}, Bakker M.R.³, Fanin N.³, Bellanger R.⁴, Bernier F.⁵,
5 De Schrijver A.⁶, Ducatillon C.⁴, Fotelli M.N.⁷, Gateble G.⁴, Gundale M.J.⁸, Larsson M.⁸, Legout
6 A.², Mason W.L.⁹, Nordin A.¹⁰, Smolander A.¹¹, Spyroglou G.⁷, Vanguelova E.I.¹², Verheyen K.¹³,
7 Vesterdal L.¹⁴, Zeller B.², Augusto L.^{3*}, Derrien D.², Buée M.^{1*}

8 ¹ Université de Lorraine, INRAE, IAM, F-54000 Nancy, France.

9 ² INRAE, BEF, F-54000 Nancy, France.

10 ³ INRAE, ISPA, Bordeaux Sciences Agro, F-33140, Villenave d’Ornon, France.

11 ⁴ INRAE, 1353 UEVT, Site de la Villa Thuret, 06600 Antibes, France

12 ⁵ INRAE, 0570 UEFP, Domaine de l’Hermitage, 33610 Cestas Pierroton, France

13 ⁶ AgroFoodNature HOGENT, Departement Biowetenschappen en Industriële Technologie,
14 9090 Melle, Belgium

15 ⁷ Forest Research Institute Hellenic Agricultural Organization Dimitra, Vassilika, 57006
16 Thessaloniki, Greece

17 ⁸ Department of Forest Ecology and Management, Swedish Univ. of Agricultural Sciences, 901-
18 83 Umeå, Sweden

19 ⁹ Forest Research, Northern Research Station, Roslin, Midlothian, Scotland EH25 9SY, UK

20 ¹⁰ Umeå Plant Science Centre (UPSC), Department of Forest Genetics and Plant Physiology,
21 Swedish University of Agricultural Sciences, 901-83 Umeå, Sweden

22 ¹¹ Natural Resources Institute Finland (Luke), Latokartanonkaari 9, 00790 Helsinki, Finland

23 ¹² Forest Research, Alice Holt, Alice Holt Lodge, Farnham, GU10 4LH, UK

24 ¹³ Forest & Nature Lab, Ghent University, Gontrode, 9090 Melle, Belgium

25 ¹⁴ Department of Geosciences and Natural Resource Management, University of Copenhagen,
26 1958 Frederiksberg C, Denmark

27 ¹⁵ Current address : IEES, Université Paris Est Créteil, CNRS, INRAE, IRD, 94010 Créteil 94010,
28 France

29

30

31 * Corresponding authors: marc.buee@inrae.fr and laurent.augusto@inrae.fr

32

33

34

35

36

37 **Summary**

38 In European forests, most tree species form symbioses with ectomycorrhizal (EM) and
39 arbuscular mycorrhizal (AM) fungi. The EM fungi are classified into different morphological
40 types based on the development and structure of their extraradical mycelium. These
41 structures could be root extensions that help trees to acquire nutrients. However, the
42 relationship between these morphological traits and functions involved in soil nutrient
43 foraging is still under debate.

44 We described the composition of mycorrhizal fungal communities under 23 tree species
45 in a wide range of climates and humus forms in Europe and investigated the exploratory
46 types of EM fungi. We assessed the response of this tree extended phenotype to humus
47 forms, as an indicator of the functioning and quality of forest soils. We found a significant
48 relationship between the relative proportion of the two broad categories of EM
49 exploration types (short- or long-distance) and the humus form, showing a greater
50 proportion of long-distance types in the least dynamic soils. As past land-use and host tree
51 species are significant factors structuring fungal communities, we showed this relationship
52 was modulated by host trait (gymnosperms versus angiosperms), soil depth and past land
53 use (farmland or forest).

54 We propose that this potential functional trait of EM fungi be used in future studies to
55 improve predictive models of forest soil functioning and tree adaptation to environmental
56 nutrient conditions.

57

58 **Introduction:**

59 For several million years, gymnosperm and angiosperm trees have co-evolved with symbiotic
60 fungal partners, which help trees grow on diverse types of soil and dominate terrestrial
61 ecosystems (Taylor et al., 2009; Augusto et al., 2014). Most tree species form symbioses with
62 either arbuscular mycorrhizal (AM) or ectomycorrhizal (EM) fungi (Brundrett and Tedersoo,
63 2018), and some fungal partners (notably EM fungi) play a key role in organic matter (OM)
64 foraging and provide nutrients to the trees (Smith & Read, 2010). Fueled by host
65 photosynthetic carbon, these fungi form vast mycelial networks that extend into the soil in
66 search of nutrients and water (Colpeart et al., 1992; Ekblad et al., 2013). Chen et al. (2016)
67 suggested that AM trees produce more roots to improve nutrient foraging, while EM trees
68 produce more mycorrhizal fungal hyphae to optimize this function. In this way, tree together
69 with its associated mycorrhizal fungi can be considered as a meta-organism or the backbone
70 of the holobiont (Lee et al., 2019; Vandenkoornhuyse et al., 2015). For EM symbiosis, it was
71 hypothesized that the extent of extraradical mycelial proliferation in the soil reflects the
72 spatial exploration capacities of fungi (Agerer, 2006; Finlay & Read, 1986). Ectomycorrhizal
73 fungi are, therefore, a root extension of trees, thereby extending the plant's phenotype
74 (Fernandez et al., 2022). Agerer (2001) classified EM mycelial systems according to the
75 patterns of different exploration types of fungal species based on the amounts of emanating
76 hyphae or the presence and differentiation of rhizomorphs, i.e. cord-forming mycelium.
77 Briefly, the 'contact' and 'short-distance' types produce only few or short and non-aggregated
78 hyphae in the near vicinity of the root tip. Conversely, 'long-distance' and 'medium-distance'
79 types form long or aggregated cords with extensive mycelia (Agerer, 2001; 2006). Tree
80 phenotypes could be extended to integrate EM fungal traits enhancing plant nutrient
81 acquisition. In certain studies, specific EM fungi, corresponding to characteristic exploration

82 types, have been found to reflect different strategies of colonization along the soil profile
83 (Anderson et al., 2014; Genney et al., 2006; Pickles and Anderson, 2016). As an example,
84 Kluting et al. (2019) reported that soil microhabitats support functionally distinct fungal
85 communities with respect to trophic mode and growth morphology, with short-distance EM
86 species being most closely associated with organic layers, while mat-forming EM taxa, in
87 particular *Sarcodon* genera, being generally dominant in mineral horizons. Consecutively,
88 these morphological traits of EM fungi have been proposed as functional traits that could link
89 EM hyphal morphologies to different nutrient acquisition capacities (Agerer, 2006; Defrenne
90 et al., 2019; Zanne et al., 2020). Moreover, an increasing number of studies based on
91 functional trait-based approaches have shown that EM fungi have different nutrient foraging
92 strategies (Koide et al., 2014), such as enzymatic activities (Buée et al., 2007; Talbot et al.,
93 2015), nitrogen uptake (Hobbie and Colpaert, 2005; Kranabetter et al., 2015; Lilleskov et al.,
94 2011), hydrophobicity (Unestam and Sun, 1995; Hobbie et al., 2022) or carbohydrate-active
95 enzyme-encoding genes (Štursová et al., 2012; Barbi et al., 2016; Maillard et al., 2018; 2023).
96 Most of these interpretations are based on studies targeting a single forest site or a single tree
97 species, with their own characteristics, such as soil fertility or the specificity of tree cover.
98 Consequently, the validation of the EM exploration type as a functional trait has not yet been
99 validated in a wide range of forest ecosystems bringing together diverse tree species, soil
100 types and climates in the same study.

101 Forests functioning and productivity result from the interactions between climate, soil
102 properties, tree species and ecosystem management (Baldrian, 2017). As forest soils are often
103 nutrient poor, tree roots and their mycorrhizal partners must forage different soil layers to
104 mobilize sufficient amounts of nutrients (Legout et al., 2020). Both partners can colonize the
105 forest floor made up of slowly decaying plant residues, and take up the organic nutrients

106 released during decomposition (Lindahl and Tunlid, 2015). They can also prospect nutrients
107 available in the mineral profile of the soil, following the alteration of minerals or the
108 decomposition of organic matter at depth. The concept of humus form was proposed by soil
109 scientists as a descriptor of forest soil functioning, bringing together several biological
110 processes taking place at the interface between plants, soil organisms and soil (Paul, 1984;
111 Ponge, 2013). Consequently, some authors proposed the “Humus Index” as a numerical score
112 which could be used as an integrator of ecosystem functioning (Ponge 2002; Ponge and
113 Chevalier, 2006; Zanella et al., 2011). Humus form is based on a morphological description of
114 OM accumulation along the soil profile, in particular through the thickness of the forest floor.
115 In mull humus forms, dead leaves are rapidly degraded at the soil surface and OM is
116 incorporated by fauna into the deeper mineral horizons, ensuring high availability of nutrients
117 for micro-organisms and plant roots. In contrast, in mor or moder, organic matter accumulates
118 in the forest floor, concentrating nutrient resources, such as plant debris, fauna and microbial
119 necromass on the surface. The question of whether the functional diversity of EM fungal
120 communities and their exploration capacities directly depend on humus index has never been
121 investigated across a large range of forest types and humus forms. In some cases, this question
122 is all the more challenging as the past land use has a lasting influence on the soil fertility of
123 recent forests (Dupouey et al., 2002; Dambrine et al., 2007; Koerner et al., 1997; Verheyen et
124 al., 1999).

125 As the diversity of EM fungal communities is strongly influenced by soil, forest cover and
126 climate parameters (Pérez-Izquierdo et al., 2021), it is essential to improve trait-based studies
127 in mycorrhizal ecology. Indeed, study of fungal traits makes it possible to identify convergent
128 functional responses of distinct mycorrhizal fungal communities (Chaudhary et al., 2022). Our
129 main objective was to study how EM exploration types respond to humus forms. Ultimately,

130 the aim was to test if the exploration types are functional traits of EM fungi related to the
131 spatial distribution of nutrient resources over the soil profile and to the spatial organization
132 of fungal species within the soil. For this purpose, we studied soil EM fungal communities,
133 using a metabarcoding approach, in seven European countries (9 forest sites) with a total of
134 34 monospecific tree plantations (nested in nine forest sites, each structured as a “common
135 gardens”). Two types of past land-use were compared: five sites had been established on
136 formerly forested soils (hereafter referred to as “ancient forests”) and the other four on
137 former agricultural land (i.e. cropland, shrubland or grassland; hereafter “recent forests”). A
138 total of 23 different tree species were studied (nine EM conifers, two AM conifers, eight EM
139 broadleaves and four AM broadleaves), covering a wide range of climates and humus forms
140 providing broad ecological soil categories. First, we tested if past land-use and tree species are
141 significant factors driving the composition of mycorrhizal fungal communities; second, we
142 used exploratory types as fungal traits potentially correlated with the humus index; and third
143 we evaluated the trait response to humus index, distinguishing between ancient and recent
144 forests. Finally, we aim to propose a conceptual framework to explain the spatial distribution
145 of mycorrhizal fungal species by linking the morphological trait of hyphae (as a potential
146 functional trait for organic nutrient foraging) to the humus index, based on two tree
147 phylogenetic and functional types: conifers and broadleaves (representative of gymnosperms
148 and angiosperms).

149

150

151 **Materials & Methods**

152 *Site characteristics and soil sampling*

153 The study was conducted in nine forest sites in Europe along a latitudinal gradient from Greece
154 to Scandinavia (i.e. Finland and Sweden). Within these sites, structured as common gardens,
155 monospecific tree plantations were established at least 30 years ago on ancient forest sites
156 (former forestry lands), or on former agricultural lands (shrubland, grassland or cropland sites)
157 leading to constitute recent forest sites (Table S1). In each site, we studied two to six tree
158 species, which were selected in order to have pure AM-species or pure ECM-species, thus
159 excluding tree species with both AM and ECM symbionts (e.g. *Eucalyptus* species) or N-fixing
160 tree species (i.e. *Alnus* or *Acacia* species). In total, 34 plots were sampled in all the common
161 gardens, corresponding to 23 different tree species: nine coniferous ectomycorrhizal trees,
162 eight broadleaved ectomycorrhizal trees, two coniferous endomycorrhizal trees and four
163 broadleaved endomycorrhizal trees. Because a few tree species were found in several
164 common gardens (Table S1), it resulted in 34 combinations of site-species, which constituted
165 our study design.

166 During sampling, a detailed description of the humus type was carried out using a classification
167 adapted from those of Brêthes et al. (1995) and Zanella et al. (2011), as summarized in Table
168 S2. This *in situ* analysis was subsequently used to define the humus index for each plot (Table
169 S3) according to Ponge & Chevalier (2006). Sampling was carried out between March 2020
170 and July 2020 (Greece, Southern France, Belgium, Central France), and between March 2021
171 and July 2021 (western France, England, Sweden, Finland, Denmark). Each year, the samplings
172 were carried out from the southern sites to the northern sites in order to ensure that the
173 phenological stages of the trees were comparable. In each plot, six bulk soil samples (6.5 cm
174 in diameter) were collected from three depths after removal of the forest floor: 0–10 cm, 10–
175 30 cm and 30–50 cm. In the Sweden and Finland sites, the bedrock was present at 30 or 40
176 cm, preventing us from collecting samples in the 30–50 cm soil layer. The six soil samples were

177 merged in the field to constitute one representative composite sample. All composites
178 samples were kept at low temperature (using an electric cool box) until being brought in the
179 lab. Then, samples were sieved to 2 mm to remove roots, stones and organic debris,
180 homogenized and sub-sampled for subsequent analysis. Part of the sub-samples was air-dried
181 and used for physical-chemical soil analyses. The remaining sub-samples were quickly
182 transferred at -80°C for molecular analyses. Soil analyses were carried out by the Laboratoire
183 d'Analyse des Sols d'Arras, INRAE, France (data in Table S3). The cation exchange capacity
184 (CEC) was determined according to the cobaltihexamine method. The pH was determined by
185 the water method using a soil/water ratio of 1:5 (w/v). Soil organic carbon (SOC) and total
186 nitrogen (N) contents were measured using a CHN analyzer, after quantification of carbonates.
187 The content in available phosphorus (P) content was determined according to the Olson
188 method (1957).

189

190 *DNA extraction, PCRs and sequencing*

191 Genomic DNA was extracted using 500 mg of frozen soil samples (n = 163) using the FastDNA™
192 SPIN Kit for Soil (MP Biomedicals, Solon, OH, USA). The manufacturer's instructions were
193 followed with a few modifications. Adapted from Luis et al. (2004), silica-DNA pellets were
194 washed two times with a solution of guanidine thiocyanate (5.5 M, pH 7) to eliminate any
195 inhibitors of polymerase, then the final pellets were resuspended in 1 ml of guanidine
196 thiocyanate, transferred to the SPIN filter and centrifuged, before continuing with the
197 manufacturer's instructions again. Amplification of fungal ITS1 region was performed using
198 the forward ITS1F (5'-CTTGGTCATTTAGAGGAAGTAA) and reverse ITS2 (5'-
199 GCTGCGTTCTTCATCGATGC) primers (Gardes and Bruns, 1993; White et al., 1990).

200 PCR was carried out in a total volume of 40 µl including 4 µl of template DNA using REDTaq
201 ReadyMix (Sigma-Aldrich, France). DNA extracts were diluted 1:100 to limit the effect of any
202 PCR inhibitors extracted from soil. PCR amplification was run following this program: 5 min at
203 94°C, 35 cycles of 1 min at 94 °C, 1 min at 52 °C, 1 min at 72°C and a final extension of 10 min
204 at 72°C. PCR barcode amplifications were confirmed using gel electrophoresis. The
205 multiplexing and the Illumina MiSeq sequencing were done by the PGBT platform (Bordeaux,
206 France).

207 Raw sequences processing was performed using FROGS (Find Rapidly OTU with Galaxy
208 Solution) pipeline (Escudié et al., 2018). Briefly, overlapping reads were merged using PEAR
209 (Magoč et al., 2011), and adapters were removed with cutadapt. Paired-end reads were
210 merged using VSEARCH (Rognes et al., 2016). Reads were then clustered with SWARM (Martin,
211 2011), chimera were detected with VSEARCH, and singletons and chimera were removed.
212 Taxonomic assignment was performed against UNITE Fungi 8.2 (Abarenkov et al 2010), and
213 OTUs with affiliations (<90% coverage or <80% identity) were filtered out. Finally, the number
214 of sequences was 982 788 for a total of 1,268 fungal OTUs. Minimum number of total reads
215 in any sample was set at 7121, 7% of the samples were excluded, only 138 samples were finally
216 used for subsequent analyses.

217

218 *Fungal community analysis and exploration type assignation*

219 The main fungal guilds were defined on the basis of taxonomic affiliation (genus level) using
220 the Fungal Traits database (Pöhlme et al., 2020) with manual curative control. A total of 403
221 ectomycorrhizal OTUs have been identified, corresponding to 48 different genera (Table 1).
222 Each genus has been assigned to an exploration type according to Agerer (2001; 2006) and
223 the dEEMY database (Agerer and Rambold, 2011). The exploration types can be separated into

224 two simplified categories based on the length of the mycelium (and presence of rhizomorphs):
225 “short distance” (SD) or “long distance” (LD). The category SD was composed by contact,
226 delicate and coarser short-distance types (*sensu* Agerer 2001). Contact exploration types have
227 few short hyphae while short distance types may have relatively larger emanating hyphae, but
228 no rhizomorphs are formed. The category LD was composed by medium-distance fringe,
229 medium-distance smooth, mat-forming and long-distance exploration types (Agerer, 2001). In
230 detail, medium rhizomorphs can be divided into three subtypes according to their
231 rhizomorphs’ characteristics (fringe, smooth and mat-forming types). Long distance types are
232 characterized by rather smooth ectomycorrhizae with few but highly differentiated
233 rhizomorphs (Agerer 2001). To calculate the relative frequencies of the two main exploratory
234 groups (LD and SD) and the corresponding LD/SD ratios, we used the presence and absence of
235 each EM genus, weighted by the number of species identified within each genus.

236

237 *Data Visualization and statistical analyses*

238 Statistical analyses and data representations were performed using R software (R Core Team,
239 2022) version 3.15. Nonmetric multidimensional scaling (NMDS, Bray–Curtis distance metric),
240 PERMANOVA (adonis2 function in the VEGAN package in R; Oksanen et al., 2013) and analyses
241 of similarity (ANOSIM) were performed to test the differences in the structure of the microbial
242 communities. The relationship between humus indexes and the proportion of long
243 exploratory type (LD/SD) was performed using Pearson correlation. Comparative analyses of
244 data were calculated using Kruskal-Wallis tests and we applied correction using the methods
245 of Benjamini-Hochberg (Benjamini and Yekutieli, 2001). Data representations were performed
246 using ggplot package (Wickham, 2016).

247

248 *Availability of data*

249 The data generated for this study have been deposited with the Sequence Read Archive and
250 are available under the bioproject number PRJNA933461.

251

252

253 **Results**

254 In total, we identified 463 EM OTUs present in one or more soil layers: 407 in the 0–10 cm
255 layer, 383 in the 10–30 cm layer and 356 in the 30–50 cm layer (Figure 1). Based on the
256 classification of Agerer (2006), the number for the SD exploration category (sum of contact
257 and short distance exploration types) varied between 194 OTUs (for 30–50 cm) and 207 OTUs
258 (for 0–10 cm). The number for the LD exploration category (sum of medium and long distance
259 exploration types with differentiated rhizomorphs) ranged from 162 OTUs (for 30–50 cm) to
260 199 OTUs (for 0–10 cm). For all the sites studied, we obtained a balanced distribution of LD
261 and SD - i.e. half-half -, in all three soil depths (Figure 1). These 463 OTUs were grouped into
262 51 genera, classified equally into SD and LD category exploration types: 25 LD, 25 SD and 1
263 unknown (Table 1). Depending on the site and associated tree species, the EM genera were
264 distributed within different but overlapping ranges of the humus index: 3–8 under coniferous
265 and 2–7 under broadleaves (Table S3). We did not find any EM genera under the six AM tree
266 species, which were characterized by relatively low humus indices: 4–5 under coniferous and
267 1–4 under broadleaves, with the exception of *Fraxinus americana* on the Belgian site, which
268 corresponded to a humus index of 7 (Figure S1).

269 Differences in the composition of EM fungal OTUs were visualized using a non-metric
270 multidimensional scale (NMDS), revealing strong clustering by site (Figure 2). The results of
271 the PERMANOVA model testing the effect of different variables on the structures of the EM

272 fungal communities showed a significant effect ($P < 0.05$) of past land-use, as well as the
273 interaction of past land use with the host species (angiosperms or gymnosperms) (Figure 2;
274 Table S4). By independently analyzing data from ancient and recent forests, our results
275 showed a significant effect of the host tree on EM communities ($P < 0.05$) in both types of
276 forest: recent and old forests (Table S4).

277 Whatever the past land-use, the humus indices of conifer and hardwood stands were
278 positively correlated with the LD/SD ratios (Figure 3A). This relationship was significant for the
279 10–30 cm and 30–50 cm layers. Although broadleaves are distributed over a lower humus
280 index range than conifers, they generally have higher LD/SD ratios than conifers. Separate
281 analysis of EM fungal communities in ancient and recent forests revealed that the relationship
282 was no longer significant for the two deepest layers in recent forests (Figure 3B). Conversely,
283 although the number of sites observed was lower, the positive relationship between the
284 humus index and the LD/SD ratio remained significant in the two deepest soil layers for the
285 ancient forest sites (Figure 3C). A comparison of soil analyses between ancient and recent
286 forests revealed significant differences in C/N and CEC, regardless of the horizons studied
287 (Table S5). In addition, we measured a significant negative correlation between the humus
288 index and certain soil fertility parameters such as N and CEC in ancient forests, illustrating the
289 link between low soil fertility and humus form (high humus index) in these sites (Table S6).
290 Interestingly, in recent forests (i.e. soils previously used for agriculture, pasture or shrubland),
291 this relationship is reversed, particularly for nitrogen, since a significant positive correlation is
292 measured between soil N content and humus index.

293

294

295 **Discussion**

296 *Factors driving the composition of mycorrhizal fungal communities*

297 In this study, we investigated the relationships between environmental factors, mycorrhizal
298 fungal communities and their host partners across 23 different tree species, represented by
299 34 monospecific stands distributed in nine sites across a European transect. Unsurprisingly,
300 we did not find EM fungi in any of the six AM species, whether conifers (*Cupressus* genus) or
301 hardwoods (*Acer*, *Celtis* and *Fraxinus* genera). In addition, the forest floors of these AM species
302 were mostly characterized by relatively low humus indices, between 1 and 5. Conversely, EM
303 tree species had humus indices between 2 and 9, with the majority of plots above an index of
304 5. These observations are consistent with the results of previous studies showing faster leaf
305 litter decomposition in AM stands compared with EM stands (Midgley et al., 2010; Phillips et
306 al., 2013). In line with our first hypothesis, we found that EM fungal assemblages were strongly
307 dependent on the host plant trait (angiosperm or gymnosperm), and soil fertility as a
308 consequence of past land-use (ancient or recent forests). These results, reported in previous
309 studies (Moora et al., 2014; van der Linde et al., 2018), highlight the need to consider these
310 factors, and their interactions, as important co-variables when studying the functional
311 diversity of EM fungi and their traits. However, there are still major gaps in our knowledge of
312 changes in fungal diversity with regard to associated functional response. Response traits
313 could explain the suitability of fungi to habitats and their ability to influence ecosystem
314 services (Koide et al., 2014). Here, we used a trait-based approach to improve our
315 understanding of the mechanisms by which EM fungi adapt and coexist in different
316 environments or niches (Lajoie and Kembel, 2019).

317

318 *Relationships between EM morphological traits and humus forms*

319 Although the richness of EM fungi decreased slightly with soil depth, we found that the
320 proportions of LD and SD exploratory types remained relatively well balanced and comparable
321 in the three soil layers studied (i.e., 50/50 ratio). Based on a gradient of forest plots
322 characterized by humus forms ranging from mesomull to humimor, we demonstrated that this
323 LD/SD balance changed significantly with the humus index, a proxy of soil fertility which is
324 negatively correlated with Ellenberg's fertility indices (Lalanne et al., 2010), and CEC or N
325 (present study). Whereas we had not observed any correlation within the forest floor (data
326 not shown), an ecological niche that can be considered a "pantry", the average proportion of
327 EM fungal OTUs belonging to the LD group increased with humus index from 37.2% (humus
328 index 2) to 55.9% (humus index 8). This strong trend, already apparent in the 0–10 cm layer,
329 was significant in the 10–30 and 30–50 cm layers for both conifer and hardwood stands. These
330 results suggest that EM fungi colonizing deeper soil layers explore a greater distance i) to reach
331 the pool of nutrients that are available in the forest floor (i.e. the pantry), particularly when
332 humus forms have changed from mull to moder; or ii) to explore and reach more effectively
333 dispersed nutrient resources in deeper and less fertile layers. Supporting the first hypothesis,
334 previous studies reported that long-distance and medium-distance exploration types directed
335 their growth to organic resource patches (Cairney, 1992; Hobbie & Agerer, 2010).
336 Consecutively, forest fungi may adapt their local mycelial proliferation and spatial distribution
337 related to the vertical distribution of resources (Rosling et al., 2003; Lindahl et al., 2007). In
338 the topsoil, the proximity of organic nutrient resources may explain why the link between the
339 long exploration type and the humus index was weaker. In a coastal pine forest, Kluting et al.
340 (2019) found that short-distance EM OTUs (e.g. *Tylospora* or *Cenococcum*) are more often
341 associated with organic layers (top soil), while OTUs with a medium- and long-distance mat-
342 forming type of exploration, such as *Sarcodon*, are found in mineral and deeper horizons with

343 little presence of roots. Moreover, Peay et al (2011) found that LD exploration types were
344 more frequent in soils with low root density, while SD exploration types were more frequent
345 in soils featuring a high root density in Bishop pine forests. Jørgensen et al. (2023) conducted
346 a 'cafeteria experiment' (i.e. mesh bags filled with different soil and sand substrates) to
347 monitor the EM fungi foraging patterns by incubating these bags in mature *Picea abies* forests.
348 They observed systematic differences in extraradical mycelium proliferation among genera in
349 different substrates. Consequently, they suggested that exploration types are not consistent
350 predictors of soil foraging, proposing that these variations are related to differences in
351 mycelial longevity and the mobility of targeted resources. The artificial nature of that
352 experiment could explain these results, particularly for the observation of adaptive traits
353 requiring long response times. In addition, we cannot rule out two types of adaptation of EM
354 fungal communities in response to resource availability in the soil: i) changes in species
355 composition to favor certain exploratory types and/or ii) changes in extraradical mycelium
356 proliferation among genera as a function of substrates. Our results support the idea that it is
357 necessary to study ectomycorrhizal species traits in order to understand the response of fungi
358 to their environment and to identify the factors that can modify these traits (i.e. trait
359 response). Nevertheless, our study focused on European forests dominated by EM tree
360 species. The objective of establishing a relationship between EM morphological characteristics
361 and humus forms could be extended to other regions of the globe, but mainly in the holarctic
362 zone. Indeed, Tedersoo et al (2010) have highlighted contrasting distribution patterns of EM
363 fungal lineages around the world. For these reasons, if our interpretations can be applied to
364 the forests of the northern hemisphere which harbor the greatest number of EM lineages,
365 future studies should be carried out for tropical forests with *Dipterocarpaceae* and
366 *Caesalpinaceae* hosts and austral areas characterized by other host taxa. In addition, vast

367 forest areas are dominated byAM tree species (Schimann et al., 2017; Davison et al., 2022),
368 requiring the study of other traits.

369

370 *Trait response of ectomycorrhizal fungi to past land-use and soil fertility*

371 Previous studies have reported that the fertility of forest soils explains fungal diversity and
372 causes shifts in mycorrhizal community with decreasing nutrient availability (Kranabetter et
373 al., 2009; Cox et al., 2010). Interestingly, the positive relationship between the humus index
374 and the proportion of long exploration type was partially disrupted in recent forests that had
375 developed on former agricultural or grassland soils. Unlike tree plantations on formerly forest
376 soil, the proportion of LD exploration types at former agricultural land remains stable, at
377 around 50%, between humus index 3 and humus index 9. In recent forests, soil functioning is
378 still in a transitional phase and humus types, normally based on the morphological description
379 of the distribution of OM along the soil profile, do not reflect the actual location of nutrient
380 resources, as organic matter cycles are understood on a secular scale (Balesdent et al., 2018).

381 Past land-use and some human activities have significant and long-lasting effects on soil
382 fertility, due to the persistence of available nutrient such as phosphorus and nitrogen in deep
383 soil horizons, which also has direct impacts on soil organisms (Compton & Boone, 2000; Jussy
384 et al., 2002; Sciama et al., 2009; Jangid et al., 2011; Fichter et al., 2014), including fungal
385 communities and specific ectomycorrhizal genera (Diedhiou et al., 2009; Kjølner et al., 2012;
386 Klavina et al., 2022; Khokon et al., 2023). In a review, Lilleskov et al. (2019) reported that fungal
387 communities primarily composed of EM exploratory types that make up the SD group were
388 characteristic of nitrogen-rich forests, after atmospheric N deposition. In line with this review,
389 we found higher cation exchange capacities and nitrogen and phosphorus contents in former
390 agricultural soils, which could explain the reduction in LD ectomycorrhizal genera in recent

391 forest soils, even in blocks with a high humus index. In addition, our results indicated that the
392 influence of past land-use had a greater effect on the functional diversity of EM fungal
393 communities in the deepest horizons, i.e. down to a depth of 50 cm. Interestingly, it has been
394 reported that EM abundance corresponding to SD exploration types (e.g. *Tylospora* spp.)
395 increased in more fertile soils of *Picea*-dominated forests (Sterkenburg et al., 2015). Similarly,
396 Guo et al (2021) observed that N fertilization induced strong effects on mycelial growth
397 characteristics in pine stands, corresponding to a shift from LD to SD exploration types.

398

399 *Elements for future studies*

400 Hobbie and Agerer (2010) demonstrated EM fungi with longest exploration types (i.e. long-
401 distance, medium-distance mat and medium distance fringe) were more enriched in ¹⁵N than
402 EM fungi with shortest exploration type (i.e. medium-distance smooth, short-distance and
403 contact). Furthermore, the relationship between organic N use and N isotopes has been
404 illustrated by Lilleskov et al. (2002), revealing that the ¹⁵N of sporocarps was highest for EM
405 fungi that use proteins, and lowest in non-users of proteins. These last authors suggested that
406 EM fungal species vary in their individual response to soil fertility and in their ability to utilize
407 nitrogen bound in soil organic matter (N-SOM). Our results confirmed the limited role of EM
408 fungi in organic N exploration under fertile conditions (Högberg, 2007; Peter et al., 2001;
409 Vesala et al., 2021), resulting in a reduction in mycelial biomass production, i.e. a lower
410 proportion of LD exploratory types (Wallander and Nylund, 1992; Nilsson et al., 2003; Sims et
411 al., 2007). This general model supports the 'ecological market' theory of ectomycorrhizal
412 symbiosis, which advocates for an increasing carbon investment by trees in ectomycorrhizal
413 biomass (and mycorrhization rate) when soil N bioavailability decreases (Franklin et al., 2014;
414 Vicca et al., 2012), and this even though tree growth is ultimately reduced. This mechanism

415 may explain the stabilization of the “rhizomorph strategy” in high index humus *via* a feedback
416 effect of nitrogen depletion.

417 Because there is a large variation in fungal community structure among regions and tree
418 species, the use of functional traits would make possible to determine common community
419 responses to environmental constraints or variables. The exploratory types could be proposed
420 as adaptive response to fertility and spatial distribution of nutrients in the soils. In turn, this
421 trait response can influence the functioning of the ecosystem (Koide et al., 2014) and notably
422 for soil carbon storage and potentially drought, as recently suggested (Castaño et al., 2023).

423 Here, we were able to establish a spatial distribution model of hyphal exploration types in
424 correlation with the humus index, as a proxy of soil fertility, to predict the foraging patterns
425 of EM fungi along the vertical soil profile (Figure 4). Confirming our hypotheses, hyphal
426 exploration type could be interpreted as a functional trait for soil exploration and foraging of
427 OM-linked nutrient. Indeed, we have shown that, in the deepest horizons, the proportion of
428 long exploratory types was higher when soil fertility was low and humus index high, and thus
429 nutrient resources which were mainly located in the forest floor at a distance from to EM
430 hyphae sampled at depth. These correlations lead to two potentially complementary
431 interpretations: the 'rhizomorphic' strategy in the deep layers makes it possible to reach the
432 reserve of nutrients available at the surface in the forest floor or to facilitate efficient
433 exploration in the deeper and least fertile soil layers. We propose that this adaptive trait of
434 EM fungi to nutrient availability in mineral soil horizons (Figure 4) may be strongly affected by
435 past land-use and potential associated fertilizer inputs. We propose to take better account EM
436 exploration types in future studies of forest fungal ecology, as a strategy of OM foraging and
437 nutrient mobilization, in particular N-SOM. However, it seems important to consider a
438 potential limitation to metabarcoding studies using soil DNA, as the presence of detected taxa

439 in soil does not guarantee that these fungal species colonize roots and form EM morphotypes.
440 Additionally, as suggested by Jørgensen et al. (2023), there may be different growth rates of
441 extraradical hyphae between exploration types as well as a level of phenotypic plasticity
442 within the same EM genus or exploratory type. We conclude that assessing fungal exploration
443 types as indicators of ecosystem functioning or assessing the response of these potential traits
444 to environmental factors could provide relevant knowledge to improve predictive models of
445 soil carbon storage under different forest management scenarios (e.g. succession, mixed
446 forests, plantation, migration). Therefore, more studies are needed coupling trait ecology
447 approaches with directly quantitative methods, such as metatranscriptomics (Auer et al.,
448 2023) and isotopic analyses (Lilleskov et al., 2002; Pellitier et al., 2021; Maillard et al., 2023).

449

450

451 **Acknowledgements**

452 This study was made possible by a large, continent-scale, field campaign. Therefore, we
453 gratefully acknowledge all the persons involved in fieldwork, samples handling, and laboratory
454 analyses: Rémi Borelle, Coralie Chesseron, Jean-Luc Denou, Céline Gire, Catherine Lambrot,
455 Julien Langrand, Tania Maxwell, Sylvie Milin, Alain Mollier, Arnaud Reichard, and Samir
456 Moutama (France); Yasmin Jaber, Stamatis Rafail Tziaferidis, Giorgos Xanthopoulos, Fani Lyrou
457 (Greece); Kris Ceunen (Belgium); Frank Ashwood, Sue Benham, Chris Reynolds, Liz Richardson,
458 Mark Oram, Tom Pettingale and Andrew Ross (England); Ming Yu and Haifeng Zheng
459 (Denmark); Dorothea Zannantonio (Sweden); Raino Lievonen (Finland). We particularly thank
460 Lucie Bon and Mohamed El-Mazlouzi, who help us at organizing and leading the field missions.
461 We also thank Bruno Ringeval and Lucas Auer for their help at collecting global climate data
462 and at performing statistical analyses. F.K. held a PhD fellowship awarded by the Grand Est

463 Region and the “Agence Nationale de la Recherche” (ANR) as part of the ANR project “CARbon
464 functional Traits and their Optimisation” - CARTON - (AAPG2019 CES32). This project was
465 supported by funds obtained from the CARTON project and the ANR as part of the
466 ‘Investissements d’Avenir’ program (ANR-11-LABX-0002-01, Lab of Excellence ARBRE).

467

468 **Conflict of interest**

469 The authors declare that they have no conflict of interest.

470

471 **Author contributions**

472 L.A. initiated and coordinated the ANR “CARTON” project. He selected the studied common
473 gardens and interacted with the scientists in charge of those sites (M.N.F., G.S., C.D., R.B.,
474 G.G., A.L., B.Z., A.D.S., K.V., E.V., B.M., L.V., A.N., M.J.G., M.L., A.S.), who provided data,
475 ensured monitoring, and supported the field campaign (hosting, field work, tools, shipments,
476 etc.). M.B., D.D. and F.K. designed this study, embedded in the CARTON project. F.K. carried
477 out the molecular biology experiments, handled data, and performed data analyses
478 (bioinformatics and statistics). M.B. and F.K. analysed and interpreted the results. M.B. wrote
479 the manuscript with the help of L.A., D.D., N.F., F.K., M.R.B., and all other authors.

480

481

482 **References**

483 Abarenkov K, Nilsson RH, Larsson K-H, Alexander IJ, Eberhardt U, Erland S, Høiland K,
484 Kjølner R, Larsson E, Pennanen T, *et al.* (2010) The UNITE database for molecular

485 identification of fungi – recent updates and future perspectives. *The New Phytologist* 186:
486 281–285.

487 Agerer R (2001) Exploration types of ectomycorrhizae. *Mycorrhiza*.11: 107–114.

488 Agerer R (2006) Fungal relationships and structural identity of their ectomycorrhizae.
489 *Mycological Progress* 5: 67–107.

490 Agerer R, Rambold G (2004) DEEMY – An Information System for Characterization and
491 Determination of Ectomycorrhizae.

492 Anderson IC, Genney DR, Alexander IJ (2014) Fine-scale diversity and distribution of
493 ectomycorrhizal fungal mycelium in a Scots pine forest. *New Phytologist* 201: 1423–1430.

494 Auer L, Buée M, Fauchery L, Lombard V, Barry KW, Clum A, ... Martin FM (2023)
495 Metatranscriptomics sheds light on the links between the functional traits of fungal guilds
496 and ecological processes in forest soil ecosystems. *New Phytologist*
497 (<https://doi.org/10.1111/nph.19471>)

498 Augusto L, Davies TJ, Delzon S, De Schrijver A (2014) The enigma of the rise of
499 angiosperms: can we untie the knot? *Ecology Letters* 17: 1326–1338.

500 Baldrian P (2017) Forest microbiome: diversity, complexity and dynamics. *FEMS*
501 *Microbiology Reviews* 41: 109–130.

502 Balesdent J, Basile-Doelsch I, Chadoeuf J, Cornu S, Derrien D, Fekiacova Z, & Hatté C.
503 (2018) Atmosphere–soil carbon transfer as a function of soil depth. *Nature* 559: 599-602.

504 Barbi F, Prudent E, Vallon L, Buee M, Dubost A, Legout A, Marmeisse R, Fraissinet-
505 Tachet L, Luis P (2016) Tree species select diverse soil fungal communities expressing
506 different sets of lignocellulolytic enzyme-encoding genes. *Soil Biology and Biochemistry*
507 100: 149–159.

508 Benjamini Y, Yekutieli D (2001) The control of the false discovery rate in multiple testing
509 under dependency. *The Annals of Statistics* 29: 1165–1188.

510 Brethes A, Brun JJ, Jabiol B, Ponge J, Toutain F (1995) Classification of forest humus
511 forms: a French proposal. *Annales des Sciences Forestières* 52: 535–546.

512 Brundrett MC, Tedersoo L (2018) Evolutionary history of mycorrhizal symbioses and
513 global host plant diversity. *New Phytologist* 220: 1108–1115.

514 Buée M, Courty PE, Mignot D, Garbaye J (2007) Soil niche effect on species diversity and
515 catabolic activities in an ectomycorrhizal fungal community. *Soil Biology and*
516 *Biochemistry* 39: 1947–1955.

517 Cairney JWG (1992) Translocation of solutes in ectomycorrhizal and saprotrophic
518 rhizomorphs. *Mycological Research* 96: 135–141.

519 Castaño C, Suarez-Vidal E, Zas R, Bonet JA, Oliva J, Sampedro L (2023) Ectomycorrhizal
520 fungi with hydrophobic mycelia and rhizomorphs dominate in young pine trees surviving
521 experimental drought stress. *Soil Biology and Biochemistry* 178: 108932.

522 Chaudhary VB, Holland EP, Charman-Anderson S, Guzman A, Bell-Dereske L, Cheeke
523 TE, Corrales A, Duchicela J, Egan C, Gupta MM, et al (2022) What are mycorrhizal traits?
524 *Trends in Ecology & Evolution* 37: 573–581.

525 Chen W, Koide RT, Adams TS, DeForest JL, Cheng L, Eissenstat DM (2016) Root
526 morphology and mycorrhizal symbioses together shape nutrient foraging strategies of
527 temperate trees. *Proceedings of the National Academy of Sciences* 113: 8741–8746.

528 Colpaert JV, Van Assche JA, Luijckens K (1992) The growth of the extramatrical mycelium
529 of ectomycorrhizal fungi and the growth response of *Pinus sylvestris* L. *New Phytologist*
530 120: 127–135.

531 Compton JE, Boone RD (2000) Long-Term Impacts of Agriculture on Soil Carbon and
532 Nitrogen in New England Forests. *Ecology* 81: 2314–2330.

533 Cox F, Barsoum N, Lilleskov EA, Bidartondo MI (2010) Nitrogen availability is a primary
534 determinant of conifer mycorrhizas across complex environmental gradients. *Ecology*
535 *Letters* 13: 1103–1113.

536 Dambrine E, Dupouey J-L, Laüt L, Humbert L, Thinon M, Beaufils T, Richard H (2007)
537 Present Forest Biodiversity Patterns in France Related to Former Roman Agriculture.
538 *Ecology* 88: 1430–1439.

539 Davison J, Vasar M, Sepp SK, Oja J, Al-Quraishy S, Bueno CG, et al., Zobel M (2022)
540 Dominance, diversity, and niche breadth in arbuscular mycorrhizal fungal communities.
541 *Ecology* <https://doi.org/10.1002/ecy.3761>

542 Defrenne CE, Philpott TJ, Guichon SHA, Roach WJ, Pickles BJ, Simard SW (2019) Shifts
543 in Ectomycorrhizal Fungal Communities and Exploration Types Relate to the Environment
544 and Fine-Root Traits Across Interior Douglas-Fir Forests of Western Canada. *Frontiers in*
545 *Plant Science* 10.

546 Diedhiou AG, Dupouey J-L, Buée M, Dambrine E, Laüt L, Garbaye J (2009) Response of
547 ectomycorrhizal communities to past Roman occupation in an oak forest. *Soil Biology and*
548 *Biochemistry* 41: 2206–2213.

549 Dupouey JL, Dambrine E, Laffite JD, Moares C (2002) Irreversible Impact of Past Land
550 Use on Forest Soils and Biodiversity. *Ecology* 83: 2978–2984.

551 Ekblad A, Wallander H, Godbold DL, Cruz C, Johnson D, Baldrian P, Björk RG, Epron D,
552 Kieliszewska-Rokicka B, Kjøller R, et al (2013) The production and turnover of
553 extramatrical mycelium of ectomycorrhizal fungi in forest soils: role in carbon cycling.
554 *Plant and Soil* 366: 1–27.

555 Escudié F, Auer L, Bernard M, Mariadassou M, Cauquil L, Vidal K, Maman S, Hernandez-
556 Raquet G, Combes S, Pascal G (2018) FROGS: Find, Rapidly, OTUs with Galaxy Solution.
557 *Bioinformatics* 34: 1287–1294.

558 Fernandez M, Vernay A, Henneron L, Adamik L, Malagoli P, Balandier P (2022) Plant N
559 economics and the extended phenotype: Integrating the functional traits of plants and
560 associated soil biota into plant–plant interactions. *Journal of Ecology* 110: 2015-2032.

561 Fichtner A, von Oheimb G, Härdtle W, Wilken C, Gutknecht JLM (2014) Effects of
562 anthropogenic disturbances on soil microbial communities in oak forests persist for more
563 than 100 years. *Soil Biology and Biochemistry* 70: 79–87.

564 Finlay RD, Read DJ (1986) The Structure and Function of the Vegetative Mycelium of
565 Ectomycorrhizal Plants. *New Phytologist* 103: 157–165.

566 Franklin O, Näsholm T, Högberg P, Högberg MN (2014) Forests trapped in nitrogen
567 limitation – an ecological market perspective on ectomycorrhizal symbiosis. *New*
568 *Phytologist* 203: 657–666.

569 Gardes M, Bruns TD (1993) ITS primers with enhanced specificity for basidiomycetes -
570 application to the identification of mycorrhizae and rusts. *Molecular Ecology* 2: 113–118.

571 Genney DR, Anderson IC, Alexander IJ (2006) Fine-scale distribution of pine
572 ectomycorrhizas and their extramatrical mycelium. *New Phytologist* 170: 381–390.

573 Guo W, Ding J, Wang Q, Yin M, Zhu X, Liu Q, Zhang Z, Yin H (2021) Soil fertility controls
574 ectomycorrhizal mycelial traits in alpine forests receiving nitrogen deposition. *Soil Biology*
575 *and Biochemistry* 161: 108386.

576 Hobbie EA, Agerer R (2010) Nitrogen isotopes in ectomycorrhizal sporocarps correspond
577 to belowground exploration types. *Plant and Soil* 327: 71–83.

578 Hobbie EA, Colpaert JV (2003) Nitrogen availability and colonization by mycorrhizal fungi
579 correlate with nitrogen isotope patterns in plants. *New Phytologist* 157: 115-126.

580 Hobbie EA, Bendiksen K, Thorp NR, Ohenoja E, Ouimette AP (2022) Climate records,
581 isotopes, and C: N stoichiometry reveal carbon and nitrogen flux dynamics differ between
582 functional groups of ectomycorrhizal fungi. *Ecosystems* 25: 1207–1217.

583 Högberg P (2007) Nitrogen impacts on forest carbon. *Nature* 447: 781–782.

584 Jangid K, Williams MA, Franzluebbers AJ, Schmidt TM, Coleman DC, Whitman WB
585 (2011) Land-use history has a stronger impact on soil microbial community composition
586 than aboveground vegetation and soil properties. *Soil Biology and Biochemistry* 43: 2184–
587 2193.

588 Jörgensen K, Clemmensen KE, Wallander H, Lindahl BD (2023) Do ectomycorrhizal
589 exploration types reflect mycelial foraging strategies? *New Phytologist* 237: 576–584.

590 Jussy JH, Koerner W, Dambrine E, Dupouey JL, Benoit M (2002) Influence of former
591 agricultural land use on net nitrate production in forest soils. *European Journal of Soil
592 Science* 53: 367–374.

593 Kranabetter JM, Hawkins BJ, Jones MD, Robbins S, Dyer T, Li T (2015) Species turnover
594 (β -diversity) in ectomycorrhizal fungi linked to uptake capacity. *Molecular Ecology* 24:
595 5992-6005.

596 Khokon AM, Janz D, Polle A (2023) Ectomycorrhizal diversity, taxon-specific traits and
597 root N uptake in temperate beech forests. *New Phytologist* 239: 739–751.

598 Kjøller R, Nilsson L-O, Hansen K, Schmidt IK, Vesterdal L, Gundersen P (2012) Dramatic
599 changes in ectomycorrhizal community composition, root tip abundance and mycelial
600 production along a stand-scale nitrogen deposition gradient. *New Phytologist* 194: 278–
601 286.

602 Klavina D, Tedersoo L, Agan A, Adamson K, Bitenieks K, Gaitnieks T, Drenkhan R (2022)
603 Soil fungal communities in young Norway spruce-dominant stands: footprints of former
604 land use and selective thinning. *European Journal of Forest Research* 141: 503–516.

605 Klütting K, Clemmensen K, Jonaitis S, Vasaitis R, Holmström S, Finlay R, Rosling A (2019)
606 Distribution patterns of fungal taxa and inferred functional traits reflect the non-uniform

607 vertical stratification of soil microhabitats in a coastal pine forest. *FEMS Microbiology*
608 *Ecology* 95: fiz149.

609 Koerner W, Dupouey JL, Dambrine E, Benoit M (1997) Influence of Past Land Use on the
610 Vegetation and Soils of Present Day Forest in the Vosges Mountains, France. *Journal of*
611 *Ecology* 85: 351–358.

612 Koide RT, Fernandez C, Malcolm G (2014) Determining place and process: functional traits
613 of ectomycorrhizal fungi that affect both community structure and ecosystem function. *New*
614 *Phytologist* 201: 433–439.

615 Korkina IN, Vorobeichik EL (2016) The humus index: A promising tool for environmental
616 monitoring. *Russian journal of ecology* 47, 526-531.

617 Kranabetter JM, Friesen J, Gamiet S, Kroeger P (2009) Epigeous fruiting bodies of
618 ectomycorrhizal fungi as indicators of soil fertility and associated nitrogen status of boreal
619 forests. *Mycorrhiza* 19: 535–548.

620 Lajoie G, Kembel SW (2019) Making the Most of Trait-Based Approaches for Microbial
621 Ecology. *Trends in Microbiology* 27: 814–823.

622 Lalanne A, Bardat J, Lalanne-Amara F, Ponge J-F (2010) Local and regional trends in the
623 ground vegetation of beech forests. *Flora - Morphology, Distribution, Functional Ecology*
624 *of Plants* 205: 484–498.

625 Lee SJ, Morse D, Hijri M (2019) Holobiont chronobiology: mycorrhiza may be a key to
626 linking aboveground and underground rhythms. *Mycorrhiza* 29: 403-412.

627 Legout A, Hansson K, van der Heijden G, Laclau J-P, Mareschal L, Nys C, Nicolas M,
628 Saint-André L, Ranger J (2020) Chemical fertility of forest ecosystems. Part 2: Towards
629 redefining the concept by untangling the role of the different components of biogeochemical
630 cycling. *Forest Ecology and Management* 461: 117844.

631 Lilleskov EA, Hobbie EA, Fahey TJ (2002) Ectomycorrhizal fungal taxa differing in
632 response to nitrogen deposition also differ in pure culture organic nitrogen use and natural
633 abundance of nitrogen isotopes. *New Phytologist* 154: 219-231.

634 Lilleskov EA, Hobbie EA, Horton TR (2011) Conservation of ectomycorrhizal fungi:
635 exploring the linkages between functional and taxonomic responses to anthropogenic N
636 deposition. *Fungal ecology* 4: 174-183.

637 Lilleskov EA, Kuyper TW, Bidartondo MI, Hobbie EA (2019) Atmospheric nitrogen
638 deposition impacts on the structure and function of forest mycorrhizal communities: A
639 review. *Environmental Pollution* 246: 148–162.

640 Lindahl BD, Ihrmark K, Boberg J, Trumbore SE, Högberg P, Stenlid J, Finlay RD (2007)
641 Spatial separation of litter decomposition and mycorrhizal nitrogen uptake in a boreal forest.
642 *New Phytologist* 173: 611–620.

643 Lindahl BD, Tunlid A (2015) Ectomycorrhizal fungi – potential organic matter
644 decomposers, yet not saprotrophs. *New Phytologist* 205: 1443–1447.

645 van der Linde S, Suz LM, Orme CDL, Cox F, Andreae H, Asi E, Atkinson B, Benham S,
646 Carroll C, Cools N, et al. (2018) Environment and host as large-scale controls of
647 ectomycorrhizal fungi. *Nature* 558: 243–248.

648 López B, Sabaté S, Gracia CA (2001) Vertical distribution of fine root density, length
649 density, area index and mean diameter in a *Quercus ilex* forest. *Tree Physiology* 21: 555–
650 560.

651 Luis P, Walther G, Kellner H, Martin F, Buscot F (2004) Diversity of laccase genes from
652 basidiomycetes in a forest soil. *Soil Biology and Biochemistry* 36: 1025–1036.

653 Magoč T, Salzberg SL (2011) FLASH: fast length adjustment of short reads to improve
654 genome assemblies. *Bioinformatics* 27: 2957–2963.

655 Maillard F, Didion M, Fauchery L, Bach C, Buée M (2018) N-Acetylglucosaminidase
656 activity, a functional trait of chitin degradation, is regulated differentially within two orders
657 of ectomycorrhizal fungi: Boletales and Agaricales. *Mycorrhiza* 28: 391–397.

658 Maillard F, Kohler A, Morin E, Hossann C, Miyauchi S, Ziegler-Devin I, Gérard D, Angeli
659 N, Lipzen A, Keymanesh K, et al (2023) Functional genomics gives new insights into the
660 ectomycorrhizal degradation of chitin. *New Phytologist* 238: 845–858.

661 Martin CE. 2011. Adapting swarm intelligence for the self-assembly and optimization of
662 networks. University of Maryland, College Park.

663 Midgley MG, Brzostek E, Phillips RP (2015) Decay rates of leaf litters from arbuscular
664 mycorrhizal trees are more sensitive to soil effects than litters from ectomycorrhizal
665 trees. *Journal of Ecology* 103: 1454-1463.

666 Moora M, Davison J, Öpik M, Metsis M, Saks Ü, Jairus T, Vasar M, Zobel M (2014)
667 Anthropogenic land use shapes the composition and phylogenetic structure of soil
668 arbuscular mycorrhizal fungal communities. *FEMS Microbiology Ecology* 90: 609–621.

669 Nilsson LO, Wallander H (2003) Production of external mycelium by ectomycorrhizal
670 fungi in a norway spruce forest was reduced in response to nitrogen fertilization. *New
671 Phytologist* 158: 409–416.

672 Oksanen J (2010) *Vegan : community ecology package*. <http://vegan.r-forge.r-project.org/>.

673 Olsen SR, Watanabe FS (1957) A Method to Determine a Phosphorus Adsorption
674 Maximum of Soils as Measured by the Langmuir Isotherm. *Soil Science Society of America
675 Journal* 21: 144–149.

676 Paul EA (1984) Dynamics of organic matter in soils. *Plant and Soil* 76: 275–285.

677 Peay KG, Kennedy PG, Bruns TD (2011) Rethinking ectomycorrhizal succession: are root
678 density and hyphal exploration types drivers of spatial and temporal zonation? *Fungal
679 Ecology* 4: 233–240.

680 Pellitier PT, Zak DR, Argiroff WA, Upchurch RA (2021) Coupled shifts in ectomycorrhizal
681 communities and plant uptake of organic nitrogen along a soil gradient: an isotopic
682 perspective. *Ecosystems* 24: 1976-1990.

683 Pérez-Izquierdo L, Rincón A, Lindahl BD, Buée M (2021) Chapter 13 - Fungal community
684 of forest soil: Diversity, functions, and services. In: Asiegbu FO, Kovalchuk A, eds. *Forest*
685 *Microbiology*. Forest Microbiology. Academic Press, 231–255.

686 Peter M, Ayer F, Egli S (2001) Nitrogen addition in a Norway spruce stand altered
687 macromycete sporocarp production and below-ground ectomycorrhizal species
688 composition. *New Phytologist* 149: 311–325.

689 Phillips RP, Brzostek E, Midgley MG (2013) The mycorrhizal-associated nutrient
690 economy: a new framework for predicting carbon–nutrient couplings in temperate
691 forests. *New Phytologist* 199: 41-51.

692 Pickles BJ, Anderson IC (2016) Spatial ecology of ectomycorrhizal fungal communities.
693 In: *Molecular Mycorrhizal Symbiosis*. John Wiley & Sons, Ltd, 363–386.

694 Pöhlme S, Abarenkov K, Henrik Nilsson R, Lindahl BD, Clemmensen KE, Kauserud H,
695 Nguyen N, Kjøller R, Bates ST, Baldrian P, et al (2020) FungalTraits: a user-friendly traits
696 database of fungi and fungus-like stramenopiles. *Fungal Diversity* 105: 1–16.

697 Ponge J-F (2003). Humus forms in terrestrial ecosystems: a framework to biodiversity. *Soil*
698 *Biology and Biochemistry* 35: 935–945.

699 Ponge J-F (2013) Plant–soil feedbacks mediated by humus forms: A review. *Soil Biology*
700 *and Biochemistry* 57: 1048–1060.

701 Ponge J-F, Chevalier R (2006) Humus Index as an indicator of forest stand and soil
702 properties. *Forest Ecology and Management* 233: 165–175.

703 Ponge J-F, Chevalier R, Loussot P (2002) Humus Index. *Soil Science Society of America*
704 *Journal* 66: 1996–2001.

705 Rdc T. 2010. R: A language and environment for statistical computing. (No Title).

706 Rognes T, Flouri T, Nichols B, Quince C, Mahé F (2016) VSEARCH: a versatile open
707 source tool for metagenomics. *PeerJ* 4: e2584.

708 Rosling A, Landeweert R, Lindahl BD, Larsson K-H, Kuyper TW, Taylor AFS, Finlay RD
709 (2003) Vertical distribution of ectomycorrhizal fungal taxa in a podzol soil profile. *New*
710 *Phytologist* 159: 775–783.

711 Sciana D, Augusto L, Dupouey J-L, Gonzalez M, Moares Domínguez C (2009) Floristic
712 and ecological differences between recent and ancient forests growing on non-acidic soils.
713 *Forest Ecology and Management* 258: 600–608.

714 Schimann H, Bach C, Lengelle J, Louisanna E, Barantal S, Murat C, Buée M (2017)
715 Diversity and structure of fungal communities in neotropical rainforest soils: the effect of
716 host recurrence. *Microbial ecology* 73: 310-320.

717 Sims SE, Hendricks JJ, Mitchell RJ, Kuehn KA, Pecot SD (2007) Nitrogen decreases and
718 precipitation increases ectomycorrhizal extramatrical mycelia production in a longleaf pine
719 forest. *Mycorrhiza* 17: 299–309.

720 Smith SE, Read DJ (2010) *Mycorrhizal Symbiosis*. Academic Press.

721 Sterkenburg E, Bahr A, Brandström Durling M, Clemmensen KE, Lindahl BD (2015)
722 Changes in fungal communities along a boreal forest soil fertility gradient. *New Phytologist*
723 207: 1145–1158.

724 Štursová M, Žifčáková L, Leigh MB, Burgess R, Baldrian P (2012) Cellulose utilization in
725 forest litter and soil: identification of bacterial and fungal decomposers. *FEMS*
726 *Microbiology Ecology* 80: 735–746.

727 Talbot JM, Martin F, Kohler A, Henrissat B, Peay KG (2015) Functional guild classification
728 predicts the enzymatic role of fungi in litter and soil biogeochemistry. *Soil Biology and*
729 *Biochemistry* 88: 441–456.

730 Taylor LL, Leake JR, Quirk J, Hardy K, Banwart SA, Beerling DJ (2009) Biological
731 weathering and the long-term carbon cycle: integrating mycorrhizal evolution and function
732 into the current paradigm. *Geobiology* 7: 171–191.

733 Tedersoo L, May TW, Smith ME (2010) Ectomycorrhizal lifestyle in fungi: global
734 diversity, distribution, and evolution of phylogenetic lineages. *Mycorrhiza* 20: 217-263.

735 Unestam T, Sun YP (1995) Extramatrical structures of hydrophobic and hydrophilic
736 ectomycorrhizal fungi. *Mycorrhiza* 5: 301-311.

737 Vandenkoornhuysen P, Quaiser A, Duhamel M, Le Van A, Dufresne A (2015) The
738 importance of the microbiome of the plant holobiont. *New Phytologist* 206: 1196-1206.

739 Verheyen K, Bossuyt B, Hermy M, Tack G (1999) The land use history (1278–1990) of a
740 mixed hardwood forest in western Belgium and its relationship with chemical soil
741 characteristics. *Journal of Biogeography* 26: 1115–1128.

742 Vesala R, Kiheri H, Hobbie EA, van Dijk N, Dise N, Larmola T (2021) Atmospheric
743 nitrogen enrichment changes nutrient stoichiometry and reduces fungal N supply to
744 peatland ericoid mycorrhizal shrubs. *Science of The Total Environment* 794: 148737.

745 Vicca S, Luysaert S, Peñuelas J, Campioli M, Chapin III FS, Ciais P, Heinemeyer A,
746 Högberg P, Kutsch WL, Law BE, et al (2012) Fertile forests produce biomass more
747 efficiently. *Ecology Letters* 15: 520–526.

748 Wallander H, Nylund J-E (1992) Effects of excess nitrogen and phosphorus starvation on
749 the extramatrical mycelium of ectomycorrhizas of *Pinus sylvestris* L. *New Phytologist* 120:
750 495–503.

751 White, Bruns T, Lee S, Taylor J (1990.) White, T. J., T. D. Bruns, S. B. Lee, and J. W.
752 Taylor. Amplification and direct sequencing of fungal ribosomal RNA Genes for
753 phylogenetics. In: 315–322.

754 Wickham H (2016) Data Analysis. In: Wickham H, ed. Use R! ggplot2: Elegant Graphics
755 for Data Analysis. Cham: Springer International Publishing, 189–201.

756 Zanine A, Jabiol B, Ponge JF, Sartori G, De Waal R, Van Delft B, Graefe U, Cools N,
757 Katzensteiner K, Hager H, et al (2011) A European morpho-functional classification of
758 humus forms. *Geoderma* 164: 138–145.

759 Zanne AE, Abarenkov K, Afkhami ME, Aguilar-Trigueros CA, Bates S, Bhatnagar JM,
760 Busby PE, Christian N, Cornwell WK, Crowther TW, et al (2020). Fungal functional
761 ecology: bringing a trait-based approach to plant-associated fungi. *Biological Reviews* 95:
762 409–433.

763

764