

# **Winter is not coming: evaluating impacts of changing winter conditions on coregonine reproductive phenology**

Taylor R Stewart, Juha Sakari Karjalainen, Matteo Zucchetta, Chloé Goulon, Orlane Anneville, Mark R Vinson, Josef Wanzenböck, Jason D Stockwell

# **To cite this version:**

Taylor R Stewart, Juha Sakari Karjalainen, Matteo Zucchetta, Chloé Goulon, Orlane Anneville, et al.. Winter is not coming: evaluating impacts of changing winter conditions on coregonine reproductive phenology. Annales de Limnologie - International Journal of Limnology, 2024, 60, pp.17. 10.1051/limn/2024014. hal-04709184

# **HAL Id: hal-04709184 <https://hal.inrae.fr/hal-04709184v1>**

Submitted on 25 Sep 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



[Distributed under a Creative Commons Attribution 4.0 International License](http://creativecommons.org/licenses/by/4.0/)

See discussions, stats, and author profiles for this publication at: [https://www.researchgate.net/publication/383871982](https://www.researchgate.net/publication/383871982_Winter_is_not_coming_evaluating_impacts_of_changing_winter_conditions_on_coregonine_reproductive_phenology?enrichId=rgreq-745c72e4034b12fb5c97f6c873d27470-XXX&enrichSource=Y292ZXJQYWdlOzM4Mzg3MTk4MjtBUzoxMTQzMTI4MTI3Njk2MzkyNEAxNzI1ODg1NjMwNzMz&el=1_x_2&_esc=publicationCoverPdf)

# [Winter is not coming: evaluating impacts of changing winter conditions on](https://www.researchgate.net/publication/383871982_Winter_is_not_coming_evaluating_impacts_of_changing_winter_conditions_on_coregonine_reproductive_phenology?enrichId=rgreq-745c72e4034b12fb5c97f6c873d27470-XXX&enrichSource=Y292ZXJQYWdlOzM4Mzg3MTk4MjtBUzoxMTQzMTI4MTI3Njk2MzkyNEAxNzI1ODg1NjMwNzMz&el=1_x_3&_esc=publicationCoverPdf) coregonine reproductive phenology

**Article** in International Journal of Limnology · September 2024

DOI: 10.1051/limn/2024014



Special issue - Biology and Management of Coregonid Fishes - 2023 Guest editors: Orlane Anneville, Chloé Goulon, Juha Karjalainen, Jean Guillard, Jared T. Myers and Jason Stockwell

RESEARCH ARTICLE

**OPEN A ACCESS** 

# Winter is not coming: evaluating impacts of changing winter conditions on coregonine reproductive phenology

Taylor R. Stewart<sup>1[,](https://orcid.org/0000-0001-6207-7466)2,\*</sup> , Juha Karjalainen<sup>[3](https://orcid.org/0000-0001-9302-1174)</sup> , Matteo Zucchetta<sup>[4](https://orcid.org/0000-0002-5431-6751)</sup> , Chloé Goulon<sup>[5](https://orcid.org/0000-0002-8070-9452)</sup> , Orlane Anneville<sup>5</sup> [,](https://orcid.org/0000-0001-5256-9539) Mark R. Vinson<sup>6</sup> , Josef Wanzenböck<sup>[7](https://orcid.org/0000-0002-9186-5565)</sup> and Jason D. Stockwell<sup>2</sup>

<sup>1</sup> Department of Biology, University of Vermont, Burlington, Vermont, USA<br>
<sup>2</sup> Rubenstein Ecosystem Science Laboratory, University of Vermont, Burlington, Vermont, USA<br>
<sup>3</sup> Department of Biological and Environmental Scie Thonon-Les-Bains, France

 $6$  United States Geological Survey, Great Lakes Science Center, Lake Superior Biological Station, Ashland, Wisconsin, USA  $7$  University of Innsbruck, Mondsee, Austria

Received: 21 January 2024; Accepted: 5 July 2024

Abstract – Fishes in northern latitude lakes are at risk from climate-induced warming because the seasonality in water temperature is degrading, which can change ecosystem properties and the phenology of life-history events. Temperature-dependent embryo development models were developed for a group of cold, stenothermic fishes (Salmonidae Coregoninae) to assess the potential impacts of climate-induced changes in water temperature on cisco (Coregonus artedi) from two populations in Lake Superior (Apostle Islands [USA] and Thunder Bay [Canada]) and one in Lake Ontario (USA), vendace (C. albula) in Lake Southern Konnevesi (Finland), and European whitefish (C. lavaretus) in lakes Southern Konnevesi, Constance (Germany), Geneva (France), and Annecy (France). Water temperatures for each study group were simulated and changes in reproductive phenology across historic (1900–2006) and three future climatic-warming scenarios (2007–2099) were investigated. Models predicted that increases in water temperatures are likely to cause delayed spawning, shorter embryo incubation durations, and earlier larval hatching. Relative changes increased as warming scenarios increased in severity and were higher for littoral as compared to pelagic populations. Our simulations demonstrated that slower cooling in the autumn and (or) more rapid warming in spring can translate into substantial changes in the reproductive phenology of coregonines among our study groups. We expect that the changes in reproductive phenology predicted by our models, in the absence of thermal or behavioral adaptation, will have negative implications for population sustainability.

Keywords: Coregonus / climate change / simulation modeling / reproductive phenology / water temperature

# 1 Introduction

Warming lake temperatures worldwide (O'Reilly et al., 2015; Maberly et al., 2020; Woolway, 2023) are an imminent threat to lacustrine fish (Dahlke et al., 2020). Water temperature directly affects the limnological characteristics of lakes and is a master factor in regulating reproduction, development, physiology, and survival of lacustrine fishes (Gillooly et al., 2002; Brown et al., 2004; Cline et al., 2013; Little et al., 2020). Unlike riverine or marine fishes, lacustrine fish are limited in their ability to evade warming habitats. The vulnerability of fish populations varies among lakes depending on habitat connectivity, the magnitude of thermal regime change, and species and life-stage specific temperature tolerances (Dahlke et al., 2020).

Holarctic fishes are predicted to be at risk because the strong seasonality in water temperature regimes in northern hemisphere lake ecosystems is degrading (Winslow et al., 2017; Sharma et al., 2019; Maberly et al., 2020; Woolway et al., 2021) and their life history is innately linked to seasonal climate patterns (Winder and Schindler, 2004). Disrupted seasonal patterns have resulted in positive feedback loops with warmer winter temperatures and (or) shorter periods of winter ice coverage, earlier and more rapid spring water warming, \*Corresponding author: [taylorstewart@utah.gov](mailto:taylorstewart@utah.gov)

This is an Open Access article distributed under the terms of the Creative Commons Attribution License ([https://creativecommons.org/licenses/by/4.0\)](https://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

warmer and prolonged summer growing seasons and stratification periods, and delayed autumn cooling (Sharma et al., 2019; Maberly et al., 2020; Woolway et al., 2021; Woolway, 2023). These changes in seasonal temperatures and lake mixing have modified lake ecosystem productivity (O'Reilly et al., 2015; Yankova et al., 2017), which can have large impacts on ecosystem structure and function (Carpenter et al., 2011; Bhagowati and Ahamad, 2019).

Most observed freshwater fish responses to climateinduced changes in thermal conditions are shifts in withinlake habitat distributions (Tunney et al., 2014; Guzzo et al., 2017) and seasonal timing of life-history events (Parmesan, 2006; Farmer et al., 2015; Slesinger et al., 2021; Woods et al., 2021). Spawning adults and embryos are hypothesized to be the most temperature-sensitive life-stages in fishes based on the concept of limited cardiorespiratory capacity and thermal tolerance; spawning adults need suitable thermal habitat for offspring to develop and aerobic capacity improves with cardiorespiratory development (Dahlke et al., 2020). Warming waters will also shorten embryo incubation times resulting in earlier larval hatch dates (Reist et al., 2006; Karjalainen et al., 2015; Stewart et al., 2021a). Fish reproductive strategies have generally evolved to account for seasonal changes in resources and allow for the most energy-intensive period of the consumer's reproductive phenology to align with the peak availability of prey resources (Hjort, 1914; Cushing, 1990; Lowerre-Barbieri et al., 2011; Lyons et al., 2015). The spatiotemporal synchrony between larval fish and their prey is believed to be an important driver of interannual variation in fish year-class strength (Cushing, 1990; Nyberg *et al.*, 2001; Straile *et al.*, 2015).

Freshwater coregonines, Salmonidae Coregoninae, are a group of cold, stenothermic fish species of high economic, ecological, and cultural importance widely distributed throughout the northern hemisphere (Bogue, 2001; Nyberg et al., 2001; Zeller et al., 2011; Lynch et al., 2016; Hodgson et al., 2020; Leppi et al., 2023). Over the past several decades, many coregonine populations have declined and are the focus of reintroduction, restoration, and conservation efforts (Zimmerman and Krueger, 2009; Anneville et al., 2015; Bronte et al., 2017). Declines in some coregonine populations throughout their range are related to low early-life survival (Nyberg et al., 2001; Parks and Rypel, 2018). Reasons for declining recruitment have been linked to invasive species (e.g., three-spined stickleback Gasterosteus aculeatus in Lake Constance) and changes in primary production (e.g., re-oligotrophication in Lake Geneva) at local scales (DeWeber et al., 2022; Bourinet et al., 2023). However, factors influencing declining recruitment remain unknown for many bodies of water, but climate-induced increases in water temperatures during critical early-life stages may be an important driver (Nyberg *et al.*, 2001; Jeppesen *et al.*, 2012; Karjalainen et al., 2015; Bourinet et al., 2023).

Coregonines generally spawn near shore in late-autumn or winter (*i.e., ca.* October-January in the northern hemisphere); embryos incubate under ice or in ice-free water and hatch in late-winter or spring (Anneville et al., 2007; Karjalainen et al., 2015; Eshenroder et al., 2016). For autumn-spawning coregonines, the seasonal decrease in water temperature initiates spawning and winter water temperature is positively related to embryo development rate and negatively related to

incubation duration (Colby and Brooke, 1973; Luczyński and Kirklewska, 1984; Gillet, 1989; Karjalainen et al., 2015; Eshenroder et al., 2016; Stewart et al., 2021a). Climate change projections suggest that warming lake temperatures in autumn and winter could alter spawning phenology, incubation time, and subsequent hatching time and survival of early-life stage coregonines (Nyberg et al., 2001; Karjalainen et al., 2015, 2016; Stewart et al., 2021b; Vinson et al., 2023).

To further investigate how warming water temperatures may influence coregonine embryo development, we modeled the response of coregonine spawning phenology and development time to future climatic-warming scenarios for coregonine populations across North America and Europe. We hypothesized that delays in coregonine spawning, as a consequence of changes in autumn or winter cooling, result in altered embryo incubation durations (i.e., number of days between spawning and hatching) and hatching times. Our predictions were that delayed cooling in autumn or winter water temperature would cause delayed spawning times, with one of the following subsequent scenarios for hatching times: (1) shorter winter duration, higher winter water temperatures, decreased embryo incubation durations, and earlier hatching times; or (2) rapid winter cooling, normal winter water temperatures, typical incubation durations, and delayed hatching times. We expected coregonine populations that historically spawn at warmer temperatures *(i.e., lower latitude*) populations) to have higher relative changes in reproductive phenology because of climate warming compared to populations that historically spawn at colder temperatures and are adapted to prolonged incubations (i.e., higher latitude populations).

# 2 Methods

Temperature-dependent embryo development models were developed for eight coregonine study groups, newly fit models were validated with existing in situ observations, lake bottom thermal regimes were developed based on climatic-warming scenarios, and future simulations for study groups were run with validated models (Fig. 1).

#### 2.1 Study locations and species

Study groups included pelagic cisco (Coregonus artedi) from two populations in Lake Superior (Apostle Islands [United States of America (USA)] and Thunder Bay [Canada]) and one in Lake Ontario (USA), pelagic vendace (C. albula) in Lake Southern Konnevesi (Finland), and the littoral species within the European whitefish species complex (C. *lavaretus*) in lakes Southern Konnevesi, Geneva (France), and Annecy (France), and the pelagic species within the European whitefish species complex (C. *lavaretus*) in Lake Constance (Germany; Fig. 2). Study groups were selected based on the availability of water temperature, spawning, and developmental data.

Lakes Superior and Ontario are large, deep lakes (>19,000  $km^2$ ; max depths  $> 244$  m) within the Laurentian Great Lakes system with a diverse range of trophic states, thermal conditions, and habitats. Lake Southern Konnevesi is an oligotrophic lake  $(120 \text{ km}^2; \text{max depth } 56 \text{ m})$  in southern Finland with free-flowing inlet and outlet streams. Lakes



Fig. 1. Workflow diagram describing the use of in situ spawning, hatching, and daily lakebed water temperature data in the model validation process.



Fig. 2. Map showing the location of each study group (LSTB = Lake Superior Thunder Bay, LSAI = Lake Superior Apostle Islands; LOCB = Lake Ontario Chaumont Bay; LSK = Lake Southern Konnevesi; LC = Lake Constance; LG = Lake Geneva; LA = Lake Annecy) sampled in North America (left) and Europe (right).

Constance and Geneva are large, deep, peri-alpine lakes (>530  $km^2$ ; max depths  $> 250$  m) in central and western Europe. Lake Annecy is a small, peri-alpine lake  $(28 \text{ km}^2; \text{max depth})$ 82 m) in western Europe. Lakes Constance, Geneva, and Annecy exhibit significant thermal stratification and have been shifting towards an oligotrophic state over the last 40 years (Bourinet et al., 2023).

## 2.2 Temperature-dependent embryo development model description

Embryo development rates were expressed as the reciprocal number of days from fertilization to a given developmental stage (Colby and Brooke, 1973; Luczyński and



Fig. 3. Predicted daily proportion of embryo development at water temperatures (°C) from validated development to 50% hatching  $(DR_{50})$  models for vendace (Coregonus albula) from Lake Southern Konnevesi, cisco (C. artedi) from lakes Superior and Ontario, and European whitefish (C. lavaretus) from Lake Geneva, and the pelagic species of European whitefish from Lake Constance. Populationspecific  $DR_{50}$  models were fit to data in Stewart *et al.*, (2021a) for C. artedi from lakes Superior and Ontario and C. albula from Lake S. Konnevesi and Stewart et al., in review) for European whitefish from Lake Geneva. Species-specific  $DR_{50}$  models were taken from Eckmann (1987) for C. lavaretus (pelagic) from Lake Constance.

Kirklewska, 1984; Eckmann, 1987). The 50% hatching development stage was chosen to be comparable among study groups. The generalized equation relating rate of development to 50% hatching  $(DR_{50})$  with respect to incubation temperature  $(x; °C)$  is

$$
DR_{50} = ab^x c^{x^2}
$$

where  $a$ ,  $b$ , and  $c$  are polynomial coefficients and the semilog form is

$$
log_{10}DR_{50} = log_{10}a + xlog_{10}b + x^2log_{10}c.
$$

 $DR_{50}$  models for cisco from lakes Superior and Ontario and vendace from Lake Southern Konnevesi were fit with experimental incubation duration data on embryos incubated at constant water temperatures of ca. 2.0, 4.5, 7.0, and 9.0  $^{\circ}$ C (Stewart et al., 2021a; Fig. 3, Tab. 1). An additional model was fit for littoral spawning morphotypes of European whitefish from Lake Geneva using embryo incubation data. Winter temperatures at Lake Geneva in the littoral zone normally decline to only 5–7 °C, so only two incubation temperatures were evaluated (contemporary at 7.0 °C and warmer winters at 9.0 °C). Thus, only a linear model was fit for European whitefish in Lake Geneva. The semilog form (i.e.,  $log_{10}DR_{50}$ ) was fit to obtain model coefficients for each study group with experimental incubation data using the mean number of days from fertilization at each incubation temperature. Furthermore, published  $DR_{50}$  models were used for the littoral and pelagic spawning species of European Whitefish (C. lavaretus macrophthalmus and C. lavaretus wartmanni, respectively) from Lake Constance, vendace from Lake Kosno (Poland), and cisco from Pickerel Lake (USA; Colby and Brooke, 1973; Luczyński and Kirklewska, 1984; Eckmann, 1987; Tab. 1). Hereafter,  $DR_{50}$  models fit to each study group from experimental data (Stewart et al., 2021a, in review) are referred to as population-specific models while  $DR_{50}$  models with published polynomial coefficients for species outside of our study groups (Colby and Brooke, 1973; Luczyński and Kirklewska, 1984; Eckmann, 1987) are species-specific models.

To predict development time to 50% hatching,  $log_{10}DR_{50}$ was calculated for each daily mean temperature since spawning and the antilog10 taken to estimate the daily proportion of development. When the cumulative daily proportions of development equaled one, 50% hatching was assumed to have occurred.

### 2.3 Data sources

Fishery-dependent spawning data were available for Lake Superior (Apostle Islands and Thunder Bay; Tab. A1). Commercial fishers from Lake Superior used gillnets to target spawning aggregations and recorded date, sex, maturity, and gonad condition from a subset of their catch (e.g., first 10 fish; Yule et al., 2008).

Fishery-independent spawning data collected during spawning surveys or broodstock collections were available for lakes Southern Konnevesi, Annecy, Geneva, and Ontario (Tab. A1). Spawning adults were collected using gillnets for lakes Southern Konnevesi, Geneva, and Annecy and trap nets for Lake Ontario at known spawning times and locations. Spawning adults collected were either assessed for gonad condition (i.e., green, ripe, spent) if actively spawning or were transported and held in hatchery ponds fed by lake water until spawning was observed. A daily distribution of spawning intensity was calculated, except for Lake Annecy where only peak spawning was reported. Daily number of ripe spawning individuals was reported for Lake Geneva, while daily gonad condition was reported for lakes Southern Konnevesi and Ontario.

Repeat larval abundance data (i.e., ca. weekly) were gathered for each study group to estimate dates of hatching (Tab. A1). Larval collections were made by either a seine or ichthyoplankton net through ice, or along the surface or at a stratified depth when lakes were ice-free (Perrier et al., 2012; Karjalainen et al., 2019; Lucke et al., 2020; McKenna et al., 2020). All larvae collections began prior to the start of hatching or with low abundance so the date of first capture was assumed to be the start hatch date, except for the Lake Superior (Apostle Islands) study group. Larvae from Lake Superior (Apostle Islands) were sampled on fixed dates annually, presumably during or after peak hatching, and thus the hatch date of individual larvae were back-calculated from total lengths assuming an absolute growth rate of  $0.18$  mm day<sup>-1</sup> (Oyadomari and Auer, 2007) and a length-at-hatch of 9.9 mm (Stewart et al., 2021a). Daily lakebed winter water temperatures were obtained from deployed temperature loggers or sondes (e.g., HOBO<sup>®</sup> Water Temperature Pro v2) for lakes Superior (Apostle Islands), Ontario, Southern Konnevesi, Geneva, and Annecy on known spawning habitat or depth (Table A.1). Only a single winter (2017-18) of lakebed temperature data was available from Chaumont Bay, Lake

Table 1. Development rate equations to 50% hatching DR<sub>50</sub> for cisco (Coregonus artedi from lakes Superior, Ontario, and Pickerel, vendace (C. albula) from lakes Kosno and Southern Konnevesi, and European whitefish (C. lavaretus) from lakes Constance and Geneva. Populationspecific DR<sub>50</sub> models were fit to data for C. artedi from lakes Superior and Ontario, C. albula and C. lavaretus from Lake S. Konnevesi (Stewart et al., 2021a), and C. lavaretus from Lake Geneva (Stewart et al., in review). Species-specific  $DR_{50}$  models were taken for C. artedi from Pickerel Lake (Colby and Brooke, 1973), C. albula from Lake Kosno (Luczyński and Kirklewska, 1984), and C. lavaretus (pelagic and littoral species) from Lake Constance (Eckmann, 1987). The coefficient of determination  $(R^2)$  was provided for models fit in this paper. All coefficients are in common logarithm (log10), and coefficients from Eckmann (1987) were transformed from natural logarithm. x is the incubation temperature in °C. – indicates no value was reported.

Model group	Species	$log_{10}DR_{50}$ =	$R^2$
Lake Superior	C. artedi	$-2.3289 + 0.0717x - 0.0001x^2$	0.97
Lake Ontario	C. artedi	$-2.3836 + 0.0643x + 0.0008x^2$	0.98
Pickerel Lake	C. artedi	$-2.4088 + 0.0720x + 0.0011x^2$	
Lake Kosno	C. albula	$-2.3035 + 0.0651x + 0.0004x^2$	
Lake S. Konnevesi	C. albula	$-2.3664 + 0.0088x + 0.0050x^2$	0.94
Lake S. Konnevesi	C. lavaretus	$-2.4183 + 0.0459x + 0.0032x^2$	0.96
Lake Constance	C. <i>lavaretus</i> (pelagic)	$-2.2419 + 0.1104x - 0.0033x^2$	
Lake Constance	C. lavaretus (littoral)	$-2.3002 + 0.1104x - 0.0031x^2$	
Lake Geneva	C. lavaretus	$-2.1159 + 0.0528x$	0.86

Ontario, and thus water temperatures from satellite surface water [\(https://podaac.ipl.nasa.gov/MEaSUREs-MUR](https://podaac.ipl.nasa.gov/MEaSUREs-MUR)) and U.S. Geological Survey Oswego River gauge (gauge number 04249000; U.S. Geological Survey, 2024) were compared to the observed lakebed winter water temperatures from Chaumont Bay in 2017-2018. The Oswego River gauge provided the closest representation of the lakebed thermal conditions of Chaumont Bay (mean daily difference =  $0.77 \degree C$ ) and was used as the in situ winter water temperatures for the Lake Ontario study group. Daily lakebed winter water temperatures for Lake Superior (Thunder Bay) were provided by the City of Thunder Bay Bare Point Water Treatment Plant, which measures water temperature at intake pipes submerged 733 m from shore at 13-m depth.

#### 2.4 Model validation

Population-specific  $DR_{50}$  models fit from experimental incubation duration data (Stewart et al., in review) were validated using in situ daily lakebed winter water temperatures at typical spawning depths to compare model outputs (i.e., hatching dates) to the observed hatching data from eight coregonine study groups across North America and Europe. Experimental data were not available to fit population-specific models for lakes Annecy and Superior (Thunder Bay), thus the population-specific model from the closest-related study group and lake was used as the best-approximating model. Our models from Lake Geneva and the Apostle Islands in Lake Superior were therefore applied to Lake Annecy and Thunder Bay, respectively.

Initiation and completion of contemporary spawning were based on observed water temperatures (Fig. 1). Coregonine spawning data to calculate the spawning temperatures were either (1) the daily proportion of ripe females leading to spawning or (2) the daily number of spawning individuals on spawning grounds. The onset of spawning was assumed to start on the date when either  $>10\%$  of females captured daily were ripe or when the number of daily spawners was  $>10\%$  of the total number of spawning individuals annually. The end of spawning was calculated as the nearest date to the start of spawning when either  $\langle 10\%$  of females were ripe or when the number of daily spawners was <10% of the total number of spawning individuals annually. If the end of spawning period could not be calculated (e.g., commercial roe fishery ceased once eggs were free flowing or only peak spawning was observed), the spawning period was assumed to end 9 days after the start of spawning (Straile *et al.*, 2015). The water temperatures from the first and last date of the spawning period were taken from in situ daily lakebed winter water temperatures and averaged across all years available to define contemporary start and ending spawning water temperatures. If the years with spawning data did not overlap the years with water temperature data, the mean starting and ending days of year for the spawning period were calculated from all years of available spawning data, and the water temperatures at each mean starting and ending spawning days of year were averaged across all available water temperature years. For Lake Constance, where the dominant species of European whitefish spawns in pelagic water and embryos sink to a depth of 200– 250 m (Straile et al., 2007), spawning start dates were calculated using published equations from the date that the upper 10-m of the water column reached 10 °C isothermal conditions and spawning was assumed to end after 9 days (Straile et al., 2015).

To validate the models for each study group, the mean spawning date was estimated from the calculated spawning start and end water temperatures for each year of available in situ water temperature data (Fig. 1). Mean observed hatching days of year were calculated as a weighted mean with daily larval abundance as the weight across all years available for each study group. The predicted development time to 50% hatching for each study group was calculated using daily water temperatures beginning at the mean spawning date for each population-specific model and, if applicable, the respective species-specific models from literature (Colby and Brooke, 1973; Luczyński and Kirklewska, 1984; Eckmann, 1987). The mean degree-days at hatching from all fitted models were

**Table 2.** Model validation results for population-specific development to 50% hatching models  $(DR_{50})$  for cisco (Coregonus artedi) from lakes Superior (Apostle Island and Thunder Bay) and Ontario, vendace (C. albula) from Lake Southern Konnevesi, and European whitefish (C. lavaretus) from lakes Geneva and Annecy. Starting and ending spawning temperatures (°C) used as model parameters are listed and the model output (i.e., degree-days at hatching) from population-specific and species-specific models were compared to in situ hatching data for each study group. N indicates the number of available years with daily in situ water temperatures and across which spawning temperatures were averaged. Bolded degree-days at hatching indicates the selected  $DR_{50}$  model for each study group, if one was validated. Population-specific  $DR_{50}$ models were fit to data for C. artedi from lakes Superior (Apostle Islands) and Ontario, C. albula and C. lavaretus from Lake S. Konnevesi (Stewart et al., 2021a), and C. lavaretus from Lake Geneva (Stewart et al., in review). Species-specific  $DR_{50}$  models were taken for C. artedi from Pickerel Lake (Colby and Brooke, 1973), C. albula from Lake Kosno (Luczyński and Kirklewska, 1984), and C. lavaretus(pelagic species) from Lake Constance (Eckmann, 1987).



estimated and compared to the mean observed degree-days at hatching for each study group. Degree-days (DD) were calculated using 0 °C as the reference temperature. A model was considered validated if the difference between the model degree-days at hatching and the mean observed degree-days at hatching was within  $\pm 50$  DD. The closest validated model (*i.e.*, either the population-specific model or species-specific model) to the mean observed DD at hatching was selected for each study group.

# 2.5 Climate scenarios

Daily lake bottom water temperatures were simulated within the Inter-Sectoral Impact Model Intercomparison Project phase 2b (ISIMIP2b) local lake sector (Warszawski et al., 2014), using the SimStrat v2.1 lake model (Gaudard *et al.*, 2019) for years 1900–2099 (Fig. A1). To drive the lake model, EWEMBI bias-corrected (Lange, 2019) climate model projections from ISIMIP2b were used, specifically projections from GFDL-ESM2M, HadGEM2-ES, IPSL-CM5A-LR, and MIROC5 for historic (1900–2006) and future periods (2007– 2099) under three representative concentration pathways (RCP): RCP 2.6, 6.0, and 8.5. These pathways included a range of potential future global radiative forcing with RCP 2.6 the lowest-emission scenario and RCP 8.5 the highestemission scenario (IPCC, 2014).

To simulate how warming water temperatures would manifest in each of our study groups, lakes from ISIMIP were used as case studies. A total of 59 modeled case-study lakes from ISIMIP were categorized into climate zones following the definitions of Maberly et al. (2020) and maximum depth zones of  $0-10$ ,  $10-25$ ,  $25-50$ , and  $50+$  m. Water temperatures from case-study lakes were averaged across all four climate model projections by each date, climate zone, and depth category to provide a simulated daily estimate of thermal spawning habitat for each climate zone and lake depth category combination. Each study group was assigned by climate zone and typical coregonine spawning depth, and the respective simulated mean daily water temperatures were used for each study group. Only 19 of the 59 available case-study lakes matched the climate zone and depth categories of our study groups and were used (Tab. A2).

Lakes Geneva and Annecy are deep, peri-alpine lakes where coregonine spawning occurs in shallow waters  $(<5$  m). The contemporary thermal spawning habitats for these two lakes were not well represented by the simulated bottom water temperature scenarios. Lakes Geneva and Annecy were available as modeled case-study lakes through ISIMIP and thus the lake-specific simulated daily water temperatures at the 5-m depth stratum were used, instead of the bottom water temperatures (mean maximum depth  $= 196$  m), as the thermal scenarios for these lakes.

### 2.6 Model simulations

For each simulated year in the model, spawning began on the first day that the 5-day running average of simulated water temperatures dropped below the population-specific start spawning temperature and ended on the first day that the 5-day running average of simulated water temperatures dropped below the population-specific end spawning temperature. Eggs were deposited throughout the defined spawning period according to the proportional rate of daily water temperature change, where larger daily decreases in temperature resulted in higher daily spawner abundances. We modeled 500 female spawners each year, with each female depositing 100 eggs to provide a relative index of cohort size. If the simulated water temperatures did not decrease sufficiently to end spawning, spawning was assumed to end after 20 days. A spawning period of this duration is reasonable for high-latitude coregonine populations (Karjalainen and Marjomäki, 2018). Daily cohorts of deposited eggs were run through the respective validated  $DR_{50}$  model for each study group, and the hatch date was estimated.



Fig. 4. Simulation model anomalies (number of days) for spawning date, incubation length, and hatching date for cisco (Coregonus artedi) from lakes Superior and Ontario and vendace (C. albula) from Lake Southern Konnevesi. Anomalies were calculated for three representative concentration pathways (RCP) from 2007–2099 compared to the mean value from the historical period (1900-2006). The rate of simulated incubation temperature (°C) change per decade is indicated in the top row for each study group and matched to the respective RCP by color. Linear regression equations for each RCP scenario and the coefficient of determination  $(R^2)$  were provided.

To estimate the magnitude of change in reproductive phenology, incubation duration was calculated for each simulated embryo and mean spawning and hatching dates for each study group across historic (1900–2005) and future periods (2006–2099). Mean spawning date, incubation duration, and hatch date anomalies were calculated as deviations from the respective mean trait value during the historical period (1900–2005) for each study group, future year, and RCP scenario. Linear regressions were fit through simulated years to trait anomalies for each study group and RCP scenario. The linear models of the trait anomalies for each RCP scenario were compared using a two-way analysis of variance (R syntax: trait anomaly  $\sim$  year class + RCP scenario) within each study group, and a Tukey post-hoc test was conducted if a significant difference among slopes was found ( $a = 0.05$ ) using the *emmeans* package v.1.10.0 (Lenth, 2021).

All simulations and analyses were performed in R version 4.2.3 (R Core Team, 2023).

# 3 Results

Our population-specific models were fit, validated, and selected for cisco from lakes Superior (Apostle Islands) and Ontario, vendace from Lake Southern Konnevesi, and European whitefish from Lake Annecy (Tabs. 1 and 2). No

model was validated for cisco from Lake Superior (Thunder Bay) and European whitefish from lakes Southern Konnevesi and Geneva because the difference between predicted and observed hatching was not within our confidence range (i.e., ±50 DD), and thus results are not reported for these study groups (Tab. 2).

Study groups with validated  $DR_{50}$  models displayed variable development rates but were similar within species across populations (Fig. 3).  $DR_{50}$  models for European whitefish from Lake Constance had the fastest development rates while models for vendace from Lake Southern Konnevesi had the slowest development rates.

#### 3.1 Spawning time

Model simulations predicted that spawning will be delayed for all study groups except the pelagic species of European whitefish from Lake Constance as climate change scenarios increase water temperatures (Figs. 4 and 5). The RCP 8.5 scenario resulted in the greatest deviation from the mean historical spawning time in all study groups (mean increase =  $0.28$  days year<sup>-1</sup>).

Spawning time of European whitefish from Lake Annecy had the largest response to increased temperature (0.61 days  $year^{-1}$  at RCP 8.5), and the RCP 8.5 scenario resulted in skipped spawning years and the complete absence of adequate



Fig. 5. Simulation model anomalies (number of days) for spawning date, incubation length, and hatching date for European whitefish (Coregonus lavaretus) from lakes Constance (pelagic species) and Annecy. Anomalies were calculated for three representative concentration pathways (RCP) from 2007-2099 compared to the mean value from the historical period (1900–2006). The rate of simulated incubation temperature ( °C) change per decade is indicated in the top row for each study group and matched to the respective RCP by color. Linear regression equations for each RCP scenario and the coefficient of determination  $(R^2)$  were provided.

thermal habitat during the reproductive period required to initiate spawning by 2080 (Fig. 5). Cisco from lakes Superior and Ontario and vendace from Lake Southern Konnevesi had similar delayed spawning responses to increased temperatures within each RCP scenario (Fig. 4). Spawning time of European whitefish from Lake Constance responded similarly among RCP scenarios and had minimal change from historical spawning times ( $< 0.07$  days year<sup> $-1$ </sup>; Fig. 5). All RCP anomaly slope pairwise comparisons were significantly different for spawning date within each study group ( $p < 0.05$ ), except for vendace from Lake Southern Konnevesi between RCP 2.6 and 6.0  $(p = 0.060;$  Fig. 6).

### 3.2 Incubation length

Embryo incubation durations (i.e., number of development days) were predicted to decrease because of increased water temperatures from climate change for all study groups examined (Figs. 4 and 5). The greatest deviation in incubation length from the mean historical incubation length was the RCP



Representative Concentration Pathway Scenarios

Fig. 6. Anomaly slopes for spawning date, incubation length, and hatching date among representative concentration pathway (RCP) scenarios. Study groups included vendace (Coregonus albula), cisco (C. artedi) from lakes Superior and Ontario, the littoral species of European whitefish (C. lavaretus) from Lake Annecy, and the pelagic species of European whitefish (C. lavaretus) from Lake Constance. The p-value from Tukey post-hoc pairwise comparisons within each study group is provided if the RCP p-value from the two-way ANOVA main effect was significant. Error bars indicate 95% confidence interval. \*\*\*  $< 0.001$ ; \*\*  $< 0.01$ ; \* =  $< 0.05$ ; ns = Not Significant (>0.05).



Fig. 7. Theoretical winter incubation periods and responses of embryo demographics under normal (2.0 °C; blue) and hypothetical warm (5.0 °C; orange) winter thermal regimes. The shaded regions indicate spawning periods (left) and hatching periods (right) that may occur between 4 and 5 °C. The 2.0 °C temperature regime is water temperature data collected from Lake Superior at 10-m depth in 2018.

8.5 scenario for all study groups (mean decrease  $= 0.29$  days year-1 ). Incubation length had similar responses within the RCP 8.5 scenario to increased temperatures among all study groups, with vendace from Lake Southern Konnevesi having the greatest response  $(-0.41$  days year<sup>-1</sup>) and the pelagic species of European whitefish from Lake Constance having the smallest response  $(-0.24 \text{ days year}^{-1})$ ; Figs. 4 and 5). Furthermore, all study groups responded similarly to increased temperatures within the RCP 6.0 and 2.6 scenarios (mean = -0.16 and -0.04 days, respectively). All RCP anomaly slope pairwise comparisons were significant for incubation length within each study group ( $p < 0.05$ ; Fig. 6).

### 3.3 Hatching

Model simulation outputs for hatching date were variable among study groups (Figs. 4 and 5). Hatching dates of cisco from Lake Ontario, vendace from Lake Southern Konnevesi, and the pelagic species of European whitefish from Lake Constance were earlier in response to increased temperatures, while hatching dates of European whitefish from Lake Annecy were later and did not change for cisco from Lake Superior as water temperatures increased. The hatching date anomaly slopes among RCP scenarios were not significantly different for cisco from Lake Superior ( $p = 0.949$ ). RCP anomaly slope pairwise comparisons between RCP 2.6 and 6.0 for Lake Ontario cisco  $(p = 0.259)$  were not significant. All other comparisons were significant ( $p < 0.05$ ; Fig. 6).

# 4 Discussion

Our hypothesis that delays in coregonine spawning, as a consequence of changes in autumn and winter cooling, would result in altered embryo incubation durations (i.e., number of days between spawning and hatching) and hatching times was supported for all study groups. Simulation models predicted that climate-induced increases in water temperatures will shift reproductive phenology of coregonines causing (1) delayed spawning, (2) shorter embryo incubation durations, and (3) varying changes to larval hatching dates (Fig. 7). Relative changes were higher with more severe climate change scenarios for littoral species where embryos have higher contemporary incubation water temperatures (e.g., European whitefish from Lake Annecy).

In our simulations, the timing and number of spawning days were regulated by when, and in some instances if, water temperatures cooled to provide adequate spawning conditions. Oocyte development in fish is driven by energy content and is relative to water temperature during the germ-cell-developing season, and final oocyte maturation is a prerequisite for successful fertilization (Nagahama and Yamashita, 2008; Burt et al., 2011; Im et al., 2016). Elevated summer and autumn temperatures could place additional energy demands on adults and require longer feeding periods later into autumn. By shifting spawning to align with cooler water temperatures, coregonines could delay the high energy demand required for oocyte development to reduce competing energy demands during the summer when metabolic rates are greatest. Spawning at suboptimal water temperatures has the potential to induce considerable fertilization failure or embryo mortality if gametes are not adequately mature prior to spawning (Burt et al., 2011). The pre-fertilization thermal environment can also have intergenerational effects and shape offspring phenotypes, and thermally stressful spawning conditions can reduce the size and swimming performance in European whitefish larvae (Kekäläinen et al., 2018). Thus, delayed spawning could be a more efficient long-term life-history strategy for population persistence than the time-intensive evolutionary process of adults and embryos adapting to increased temperatures.

Shifting reproductive timing is a plausible response to warming temperatures. Contemporary coregonine populations exhibit multiple spawning strategies ranging from autumn to spring-spawning stocks (Eronen and Lahti, 1988; Hénault and Fortin, 1991; Schulz et al., 2006; Ohlberger et al., 2008). Thermal habitat used by high-latitude or deep-spawning populations could be less affected by climate change and may still provide adequate spawning and incubation conditions if water temperatures continue to rise, but this may not be the case for low-latitude or littoral-spawning populations. For example, Lake Annecy is near the southern native extent for European whitefish and had the largest shift in spawning time among our study groups, but adequate thermal spawning temperatures were absent by the middle of the 21st century under the worst-case climate scenario. This projection is likely to be applicable for populations within the same geographic region that inhabit lakes with similar morphology and thermal conditions where population-specific  $DR_{50}$  models were not able to be validated, and even more dire projections could be expected for non-native populations south of Lake Annecy (e.g., Lake Garda [Italy]; Volta et al., 2018). However, these southern European lakes are deep ( $>80$  m) with stable thermal refugia outside of the littoral zone (Kelly *et al.*, 2020). Shifting spawning strategies to use deeper habitats could provide adequate thermal conditions for ovulation to initiate, but whether suitable coregonine spawning habitat is available in these deeper waters, including sufficient oxygen levels, is unknown (Jane et al., 2020; Kelly et al., 2020; Desgué-Itier et al., 2023).

Spawning scenarios determined many of the changes in reproduction phenology but embryos are static and unable to evade changes in winter water temperatures post-spawning (Stockwell et al., 2014). Elevated incubation water temperatures accelerated embryo rates of development and hatching time and were not negated by delayed spawning. The frequency of shorter, warmer winters is projected to increase (Sharma et al., 2019), which corresponds with our simulation results suggesting future incubation durations will decrease across all study groups. In the absence of changes to spawning behavior or habitat, warmer and shorter incubations could cause higher coregonine embryo and larvae mortality, increased occurrence of embryo deformities, and smaller and less robust larvae at hatching compared to colder, longer incubations (Stewart et al., 2021a,b). Additionally, reduced ice formation caused by warmer air and water temperatures could decrease embryo survival from increased storm displacement and sediment deposition while decreasing lengths-at-hatching (Kangur et al., 2020; Stewart et al., 2021b). A complex mix of environmental factors during embryogenesis could generate high variability in hatching success and larval fitness - this was not considered in our modeling efforts (Hjort, 1914; Houde, 1989; Marjomäki et al., 2004; Fig. 7).

Coregonine populations where embryos incubate in cold littoral waters (<4 °C) are hypothesized to rely on ice-break up and spring warming to trigger final embryo development stages and hatching (Karjalainen et al., 2015). The rapid increase in spring water temperatures also synchronizes other phenological processes such as the onset of spring plankton blooms (Sommer et al., 2012). Our simulations suggest significant changes in length of incubation and subsequent hatching times, and changes in thermal regimes and time of hatching may result in temporal separation between coregonine larvae and their zooplanktonic prey. Increases in seasonal temperature variability could also cause mismatches with larval zooplankton prey if temperature changes are not heterogeneous across seasons (e.g., a cold winter followed by a warm spring; Straile et al., 2015).

Fish year-class strength is often dependent on larvae surviving from hatch through the first three to six months of life and successfully transitioning from endogenous to exogenous feeding (Hjort, 1914; Houde and Hoyt, 1987; Cushing, 1990). Numerous size-dependent processes strongly influence when larvae need to first feed and their ability to successfully forage (Miller et al., 1988). Coregonine larval body size at hatching is inversely related to incubation water temperature, and length-at-hatch and yolk-sac volume have a negative relationship (Karjalainen et al., 2015; Stewart et al., 2021a). Warmer incubations result in coregonine larvae hatching earlier with smaller lengths and larger yolk-sacs (Stewart et al., 2021a). Increased yolk-sac energy reserves at hatching may help larvae reduce starvation risk but the rate of endogenous feeding (i.e., yolk consumption) is regulated by metabolic demand (Kamler, 2008). Earlier and warmer spring water temperatures in nursery zones, which induce earlier hatching, could accelerate yolk consumption in newly hatched larvae and counteract the physiological trade-off between length-at-hatching and yolk-sac volume. Larvae hatching earlier in the spring may also have reduced swimming abilities, reduced visual acuity, and more gape limitations, which can impact their ability to evade predators and forage efficiently (Miller et al., 1988; Myers et al., 2014). Determining the physiological stress response of hatching earlier and the impact warming nursery water temperatures may have on yolk conversion efficiency is a logical and necessary next step to build on our simulation results.

Our models extend earlier approaches (Colby and Brooke, 1973; Luczyński and Kirklewska, 1984; Eckmann, 1987) by incorporating mechanistic relationships to future climatechange scenarios from a wider range of populations. Because coregonines are highly developmentally plastic and exhibit diverse spawning behaviors (Muir et al., 2013), the transferability of published models to other populations appears limited. High-quality in situ reproductive and embryo development data are critical to fitting and validating new population-specific development models. For example, hatching data for European whitefish in Lake Geneva were only available for a single year with a limited sampling period and likely did not capture the start of hatching. This led to development models underestimating hatching when, in reality, the in situ data likely did not represent the true hatching period and created a limitation in our modeling efforts. Increasing field sampling efforts around these critical life stages will benefit future research and climate-change predictability for both applied and modeling-based methods. Our simulations demonstrated that subtle changes in water temperature could translate into substantial changes in the reproductive phenology of coregonines among our study groups. Long-term changes in environmental conditions during reproductive and development periods could play a large role in generating variability in offspring success (Houde and Hoyt, 1987; Little et al., 2020). The results of our modeling efforts highlight how water temperature is

fundamental in regulating biological and physiological processes, but the impact of these changes is difficult to decipher as coregonines are behaviorally and developmentally plastic (Muir et al., 2013; Karjalainen et al., 2015). Quantifying the relationships between water temperature and coregonine reproductive phenology across a wide range of populations will be useful for managers to determine which populations may be more susceptible to the consequences of climate change and help prioritize conservation and restoration efforts. Unless coregonines exhibit behavioral or thermal adaptations to changing environmental conditions (e.g., McQuinn, 1997; Jørgensen et al., 2006), we expect that the changes in coregonine reproductive phenology predicted by our models will have negative implications for population sustainability throughout the 21<sup>st</sup> century, even under the lowest climate-emission scenario.

# APPENDIX A

Table A.1. Years of in situ data used for model validation.

Study group	Water temperature	Spawning	Hatching
Lake Superior (Apostle Islands)	$2016 - 18$	$2016 - 18$	$2016 - 18$
Lake Superior (Thunder Bay)	$2017 - 21$	$2005; 2007 - 08; 2010$	$2008 - 09$
Lake Ontario	$2012 - 21$	2007; 2017; 2019-20	$2004 - 06$
Lake S. Konnevesi	$2019 - 21$	$2019 - 21$	$2019 - 21$
Lake Geneva	$2010 - 21$	$2016 - 19$	2016
Lake Annecy	$2005 - 20$	$2016 - 19$	$2005 - 07$

Table A.2. Study group and modeled case-study lakes from inter-sectoral impact model intercomparison project that match study group parameters (the lakes that were averaged together by category).





Fig. A.1. Simulated daily lake bottom water temperatures for three representative concentration pathways (RCP) in 2020, 2040, 2060, and 2080 for Lake Superior, Lake Ontario, Lake Southern Konnevesi, Lake Constance, and Lake Annecy.

#### Acknowledgments

We thank Jim McKenna (U.S. Geological Survey Tunison Laboratory of Aquatic Science), Mike Connerton (New York State Department of Environmental Conservation Cape Vincent Fisheries Station), Eric Berglund (Ontario Ministry of Natural Resources and Forestry), Scott Sapper and Dray Carl (Wisconsin Department of Natural Resources Bayfield Fisheries Field Station), Jared Myers (U. S. Fish & Wildlife Service Ashland Fish and Wildlife Conservation Office), and Dan Yule (U.S. Geological Survey Lake Superior Biological Station) for providing coregonine spawning and/or hatching data. Brian Weidel (U.S. Geological Survey Lake Ontario Biological Station), Brenda Lafrancois and Jay Glase (National Park Service), and Walter Turek (City of Thunder Bay) provided winter water temperatures from Chaumont Bay, Lake Ontario, Apostle Islands, Lake Superior, and Thunder Bay, Lake Superior, respectively. Spawning data for Lake Geneva were collected during spawning surveys performed by the Observatory on Lakes for the International Commission for the Protection of Waters of Lake Geneva long-term monitoring program. Spawning data for Lake Annecy were collected by the Observatory on Lakes for the SILA-DDT-professional fishermen-ALP association program. Timo Marjomäki, Jonna Kuha, and staff at the Konnevesi Research Station helped obtain temperature, spawning, and hatching data of Konnevesi vendace and whitefish. This work was funded by the U.S. Geological Survey under Grant No. G16AP00087 to the Vermont Water Resources and Lakes Studies Center. We acknowledge INRAE, French National Research Institute for Agriculture, Food, and Environment, the UMR CARRTEL (INRAE - USMB) and the National Science Foundation (award number 1829451) for supporting a workshop to develop this work. Nicole Berry provided the U.S. Geological Survey solicited review that strengthened the manuscript, as did anonymous peer reviewers. Any use of trade, product, or firm names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

### **References**

- Anneville O, Lainé L, Benker S, Ponticelli A, Gerdeaux D. 2007. Food habits and ontogentic changes in the diet of whitefish larvae in Lake Annecy. Bull Fr Pêche Piscic 21–33.
- Anneville O, Lasne E, Guillard J, Eckmann R, Stockwell JD, Gillet C, Yule DL. 2015. Impact of fishing and stocking practices on coregonid diversity. Food Nutr Sci 06: 1045–1055.
- Bhagowati B, Ahamad KU. 2019. A review on lake eutrophication dynamics and recent developments in lake modeling. Ecohydrol Hydrobiol 19: 155–166.
- Bogue MB. 2001. Fishing the Great Lakes: An Environmental History, 1783–1933. Madison, Wisconsin: Univ of Wisconsin Press.
- Bourinet F, Anneville O, Drouineau H, Goulon C, Guillard J, Richard A. 2023. Synchrony in whitefish stock dynamics: disentangling the effects of local drivers and climate. J Limnol 82.
- Bronte CR, Bunnell DB, David SR, Gordon R, Gorsky D, Millard MJ, Read J, Stein RA, Vaccaro L. 2017. Report from the workshop on coregonine restoration science. U.S. Geological Survey Open-File Report 2017-1081, 23 p, [https://doi.org/](https://doi.org/10.3133/ofr20171081) [10.3133/ofr20171081](https://doi.org/10.3133/ofr20171081)
- Brown JH, Gillooly JF, Allen AP, Savage VM, West GB. 2004. Toward a metabolic theory of ecology. Ecology 85: 1771–1789.
- Burt JM, Hinch SG, Patterson DA. 2011. The importance of parentage in assessing temperature effects on fish early life history: a review of the experimental literature. Rev Fish Biol Fish 21: 377–406.
- Carpenter SR, Stanley EH, Vander Zanden MJ. 2011. State of the world's freshwater ecosystems: physical, chemical, and biological changes. Annu Rev Environ Resour 36: 75–99.
- Cline TJ, Bennington V, Kitchell JF. 2013. Climate change expands the spatial extent and duration of preferred thermal habitat for lake superior fishes. PLoS ONE 8: 1–8.
- Colby PJ, Brooke LT. 1973. Effects of temperature on embryonicdevelopment of lake herring (Coregonus artedii). J Fish Res Board Can 30: 799–810.
- Cushing DH. 1990. Plankton production and year-class strength in fish populations: an update of the match/mismatch hypothesis. Adv Mar Biol 26: 249–293.
- Dahlke FT, Wohlrab S, Butzin M, Pörtner H-O. 2020. Thermal bottlenecks in the life cycle define climate vulnerability of fish. Science 369: 65–70.
- Desgué-Itier O, Melo Vieira Soares L, Anneville O, Bouffard D, Chanudet V, Danis PA, Domaizon I, Guillard J, Mazure T, Sharaf N, Soulignac F, Tran-Khac V, et al., 2023. Past and future climate change effects on the thermal regime and oxygen solubility of four peri-alpine lakes. Hydrol Earth Syst Sci 27: 837–859.
- DeWeber JT, Baer J, Rösch R, Brinker A. 2022. Turning summer into winter: nutrient dynamics, temperature, density dependence and invasive species drive bioenergetic processes and growth of a keystone coldwater fish. Oikos 2022: e09316.
- Eckmann R. 1987. A comparative study on the temperature dependence of embryogenesis in three coregonids (Coregonus spp.) from Lake Constance. Swiss J Hydrol 49: 353–362.
- Eronen T, Lahti E. 1988. Life cycle of winter spawning vendace (Coregonus albula L.) in Lake Kajoonjärvi, eastern Finland. Finn Fish Res 9: 197–203.
- Eshenroder RL, Vecsei P, Gorman OT, Yule DL, Pratt TC, Mandrak NE, Bunnell DB, Muir AM. 2016. Ciscoes (Coregonus, subgenus Leucichthys) of the Laurentian Great Lakes and Lake Nipigon. Gt. Lakes Fish. Comm. Misc. Publ. 1: 156.
- Farmer TM, Marschall EA, Dabrowski K, Ludsin SA. 2015. Short winters threaten temperate fish populations. Nat Commun 6: 7724.
- Gaudard A, Råman Vinnå L, Bärenbold F, Schmid M, Bouffard D. 2019. Toward an open access to high-frequency lake modeling and statistics data for scientists and practitioners-the case of Swiss lakes using Simstrat v2. 1. Geosci Model Dev 12: 3955–3974.
- Gillet C. 1989. Le déroulement de la fraie des principauxpoissons lacustres. Hydroécologie Appliquée 1: 117-143.
- Gillooly JF, Charnov EL, West GB, Savage VM, Brown JH. 2002. Effects of size and temperature on developmental time. Nature 417: 70–73.
- Guzzo MM, Blanchfield PJ, Rennie MD. 2017. Behavioral responses to annual temperature variation alter the dominant energy pathway,

growth, and condition of a cold-water predator. Proc Natl Acad Sci 114: 9912–9917.

- Hénault M, Fortin R. 1991. Early life stages, growth, and reproduction of spring-spawning ciscoes (Coregonus artedii) in Lac des Écorces, Quebec. Can J Zool 69: 1644–1652.
- Hjort J. 1914. Fluctuations in the great fisheries of Northern Europe. Copenhagen.
- Hodgson EE, Hovel RA, Ward EJ, Lord S, Moore JW. 2020. Migratory diversity in an Arctic fish supporting subsistence harvest. Biol Conserv 248: 108685.
- Houde ED. 1989. Comparative growth, mortality, and energetics of marine fish larvae: temperature and implied latitudinal effects. Fish Bull 87: 471–495.
- Houde ED, Hoyt RD. 1987. Fish early life dynamics and recruitment variability. Trans Am Fish Soc.
- Im J, Kong D, Ghil S. 2016. Effects of water temperature on gonad development in the cold-water fish, kumgang fat minnow Rhynchocypris kumgangensis. Cytologia (Tokyo) 81: 311–317.
- IPCC. 2014. Climate Change 2014: Synthesis Report. Geneva, Switzerland: IPCC; 151 pp.
- Jane SF, Hansen GJ, Benjamin K, Leavitt PR, Mincer JL, North RL, Pilla RM, Stetler JT, Williamson CE, Woolway RI, Arvola L, Chandra S, et al., 2020. Environmental Data Initiative.
- Jeppesen E, Mehner T, Winfield IJ, Kangur K, Sarvala J, Gerdeaux D, Rask M, Malmquist HJ, Holmgren K, Volta P, Romo S, Eckmann R, et al., 2012. Impacts of climate warming on the long-term dynamics of key fish species in 24 European lakes. Hydrobiologia 694: 1–39.
- Jørgensen C, Ernande B, Fiksen Ø, Dieckmann U. 2006. The logic of skipped spawning in fish. Can J Fish Aquat Sci 63: 200–211.
- Kamler E. 2008. Resource allocation in yolk-feeding fish. Rev Fish Biol Fish 18: 143.
- Kangur K, Ginter K, Kangur A, Kangur P, Möls T. 2020. How did the late 1980s climate regime shift affect temperature-sensitive fish population dynamics: case study of Vendace (Coregonus albula) in a large north-temperate lake. Water 12: 2694.
- Karjalainen J, Jokinen L, Keskinen T, Marjomäki TJ. 2016. Environmental and genetic effects on larval hatching time in two coregonids. Hydrobiologia 780: 135–143.
- Karjalainen J, Juntunen J, Keskinen T, Koljonen S, Nyholm K, Ropponen J, Sjövik R, Taskinen S, Marjomäki TJ. 2019. Dispersion of vendace eggs and larvae around potential nursery areas reveals their reproductive strategy. Freshw Biol 64: 843–855.
- Karjalainen J, Keskinen T, Pulkkanen M, Marjomäki TJ. 2015. Climate change alters the egg development dynamics in cold-water adapted coregonids. Environ Biol Fishes 98: 979–991.
- Karjalainen J, Marjomäki TJ. 2018. Communal pair spawning behaviour of vendace (Coregonus albula) in the dark. Ecol Freshw Fish 27: 542–548.
- Kekäläinen J, Oskoei P, Janhunen M, Koskinen H, Kortet R, Huuskonen H. 2018. Sperm pre-fertilization thermal environment shapes offspring phenotype and performance. J Exp Biol 221: 1–8.
- Kelly S, Moore TN, de Eyto E, Dillane M, Goulon C, Guillard J, Lasne E, McGinnity P, Poole R, Winfield IJ. 2020. Warming winters threaten peripheral Arctic charr populations of Europe. Clim Change 1–20.
- Lange S. 2019. Trend-preserving bias adjustment and statistical downscaling with ISIMIP3BASD (v1. 0). Geosci Model Dev 12: 3055-3070.
- Lenth RV. 2021. emmeans: Estimated Marginal Means, aka Least-Squares Means. <https://cran.r-project.org/package=emmeans>.
- Leppi JC, Rinella DJ, Wipfli MS, Liljedahl AK, Seitz AC, Falke JA. 2023. Climate change risks to freshwater subsistence fisheries in Arctic Alaska: insights and uncertainty from broad whitefish Coregonus nasus. Fisheries 48: 295–306.
- Little AG, Loughland I, Seebacher F. 2020. What do warming waters mean for fish physiology and fisheries? J Fish Biol 97: 328–340.
- Lowerre-Barbieri SK, Ganias K, Saborido-Rey F, Murua H, Hunter JR. 2011. Reproductive timing in marine fishes: variability, temporal scales, and methods. Mar Coast Fish 3: 71–91.
- Lucke VS, Stewart TR, Vinson MR, Glase JD, Stockwell JD. 2020. Larval Coregonus spp. diets and zooplankton community patterns in the Apostle Islands, Lake Superior. J Gt Lakes Res 46: 1391–1401.
- Luczyński M, Kirklewska A. 1984. Dependence of Coregonus albula embryogenesis rate on the incubation temperature. Aquaculture 42: 43–55.
- Lynch AJ, Cooke SJ, Deines AM, Bower SD, Bunnell DB, Cowx IG, Nguyen VM, Nohner J, Phouthavong K, Riley B, Rogers MW, Taylor WW, et al., 2016. The social, economic, and environmental importance of inland fish and fisheries. Environ Rev 24: 115–121.
- Lyons J, Rypel AL, Rasmussen PW, Burzynski TE, Eggold BT, Myers JT, Paoli TJ, McIntyre PB. 2015. Trends in the reproductive phenology of two Great Lakes fishes. Trans Am Fish Soc 144: 1263–1274.
- Maberly SC, O'Donnell RA, Woolway RI, Cutler MEJ, Gong M, Jones ID, Merchant CJ, Miller CA, Politi E, Scott EM. 2020. Global lake thermal regions shift under climate change. Nat Commun 11: 1–9.
- Marjomäki TJ, Auvinen H, Helminen H, Huusko A, Sarvala J, Valkeajärvi P, Viljanen M, Karjalainen J. 2004. Spatial synchrony in the inter-annual population variation of vendace (Coregonus albula (L.)) in Finnish lakes. Ann Zool Fenn 41: 225–240.
- McKenna JE, Stott W, Chalupnicki M, Johnson JH. 2020. Spatial segregation of cisco (Coregonus artedi) and lake whitefish (C. clupeaformis) larvae in Chaumont Bay, Lake Ontario. J Gt Lakes Res 46: 1485–1490.
- McQuinn IH. 1997. Metapopulations and the Atlantic Herring. Rev Fish Biol Fish 7: 297–329.
- Miller TJ, Crowder LB, Rice JA, Marschall EA. 1988. Larval size and recruitment mechanisms in fishes: toward a conceptual framework. Can J Fish Aquat Sci 45: 1657–1670.
- Muir AM, Vecsei P, Pratt TC, Krueger CC, Power M, Reist JD. 2013. Ontogenetic shifts in morphology and resource use of cisco Coregonus artedi. J Fish Biol 82: 600–617.
- Myers JT, Yule DL, Jones ML, Quinlan HR, Berglund EK. 2014. Foraging and predation risk for larval cisco (Coregonus artedi) in Lake Superior: a modelling synthesis of empirical survey data. Ecol Model 294: 71–83.
- Nagahama Y, Yamashita M. 2008. Regulation of oocyte maturation in fish. Dev Growth Differ 50: S195–S219.
- Nyberg P, Bergstrand E, Degerman E, Enderlein O. 2001. Recruitment of pelagic fish in an unstable climate: studies in Sweden's four largest lakes. Ambio 30: 559–564.
- Ohlberger J, Mehner T, Staaks G, Hölker F. 2008. Is ecological segregation in a pair of sympatric coregonines supported by divergent feeding efficiencies? Can J Fish Aquat Sci 65: 2105– 2113.
- O'Reilly CM, Rowley RJ, Schneider P, Lenters JD, Mcintyre PB, Kraemer BM. 2015. Rapid and highly variable warming of lake surface waters around the globe. Geophys Res Lett 42: 1–9.
- Oyadomari JK, Auer NN. 2007. Influence of rearing temperature and feeding regime on Otolith increment deposition in larval Ciscoes. Trans Am Fish Soc 136: 766–777.
- Parks TP, Rypel AL. 2018. Predator-prey dynamics mediate longterm production trends of cisco (Coregonus artedi) in a northern Wisconsin lake. Can J Fish Aquat Sci 75: 1969–1976.
- Parmesan C. 2006. Ecological and evolutionary responses to recent climate change. Annu Rev Ecol Evol Syst 37: 637–669.
- Perrier C, Molinero JC, Gerdeaux D, Anneville O. 2012. Effects of temperature and food supply on the growth of whitefish Coregonus lavaretus larvae in an oligotrophic peri-alpine lake. J Fish Biol 81: 1501–1513.
- R Core Team, 2023. R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing, <https://www.r-project.org/>.
- Reist JD, Wrona FJ, Prowse TD, Power M, Dempson JB, Beamish RJ, King JR, Carmichael TJ, Sawatzky CD. 2006. General effects of climate change on Arctic fishes and fish populations. AMBIO J Hum Environ 35: 370–380.
- Schulz M, Freyhof J, Saint-Laurent R, Østbye K, Mehner T, Bernatchez L. 2006. Evidence for independent origin of two spring-spawning ciscoes (Salmoniformes: Coregonidae) in Germany. J Fish Biol 68: 119–135.
- Sharma S, Blagrave K, Magnuson JJ, O'Reilly CM, Oliver S, Batt RD, Magee MR, Straile D, Weyhenmeyer GA, Winslow LA. 2019. Widespread loss of lake ice around the Northern Hemisphere in a warming world. Nat Clim Change 9: 227.
- Slesinger E, Jensen OP, Saba G. 2021. Spawning phenology of a rapidly shifting marine fish species throughout its range. ICES J Mar Sci 78: 1010–1022.
- Sommer U, Adrian R, De Senerpont Domis L, Elser JJ, Gaedke U, Ibelings B, Jeppesen E, Lürling M, Molinero JC, Mooij WM. 2012. Beyond the Plankton Ecology Group (PEG) model: mechanisms driving plankton succession. Annu Rev Ecol Evol Syst 43: 429–448.
- Stewart TR, Brun C, Goulon C, Baer J, Karjalainen J, Guillard J, Lasne E. in review. Response of whitefish embryos to thermal conditions diverges between perialpine populations. Int J Limnol. DOI: <https://doi.org/10.1051/limn/2024017>
- Stewart TR, Mäkinen M, Goulon C, Guillard J, Marjomäki TJ, Lasne E, Karjalainen J, Stockwell JD. 2021a. Influence of warming temperatures on coregonine embryogenesis within and among species. Hydrobiologia 848: 4363–4385.
- Stewart TR, Vinson MR, Stockwell JD. 2021b. Shining a light on Laurentian Great Lakes cisco (Coregonus artedi): how ice coverage may impact embryonic development. J Gt Lakes Res 47: 1410–1418.
- Stockwell JD, Yule DL, Hrabik TR, Sierszen ME, Isaac EJ. 2014. Habitat coupling in a large lake system: delivery of an energy subsidy by an offshore planktivore to the nearshore zone of Lake Superior. Freshw Biol 59: 1197-1212.
- Straile D, Eckmann R, Jüngling T, Thomas G, Löffler H. 2007. Influence of climate variability on whitefish (Coregonus lavaretus) year-class strength in a deep, warm monomictic lake. Oecologia 151: 521–529.
- Straile D, Kerimoglu O, Peeters F. 2015. Trophic mismatch requires seasonal heterogeneity of warming. Ecology 96: 2794–2805.
- Tunney TD, McCann KS, Lester NP, Shuter BJ. 2014. Effects of differential habitat warming on complex communities. Proc Natl Acad Sci 111: 8077–8082.
- U.S. Geological Survey, 2024, USGS 04249000, Oswego River at Lock 7, Oswego, NY, in USGS water data for the Nation: U.S. Geological Survey National Water Information System database, accessed 14 September 2021 at [https://doi.org/10.5066/F7P55KJN.](https://doi.org/10.5066/F7P55KJN) [Site information directly accessible at [https://waterdata.usgs.gov/](https://waterdata.usgs.gov/nwis/uv?site_no=04249000&legacy=1) [nwis/uv?site\\_no=04249000&legacy=1\]](https://waterdata.usgs.gov/nwis/uv?site_no=04249000&legacy=1)
- Vinson MR, Herbert ME, Ackiss AS, Dobosenski JA, Evrard LM, Gorman OT, Lyons JF, Phillips SB, Yule DL. 2023. Lake Superior Kiyi reproductive biology. Trans Am Fish Soc 152: 75–93.
- Volta P, Jeppesen E, Sala P, Galafassi S, Foglini C, Puzzi C, Winfield IJ. 2018. Fish assemblages in deep Italian subalpine lakes: history and present status with an emphasis on non-native species. Hydrobiologia 824: 255–270.
- Warszawski L, Frieler K, Huber V, Piontek F, Serdeczny O, Schewe J. 2014. The inter-sectoral impact model intercomparison project (ISI-MIP): project framework. Proc Natl Acad Sci 111:3228–3232.
- Winder M, Schindler DE. 2004. Climate change uncouples trophic interactions in an aquatic ecosystem. Ecology 85: 2100–2106.
- Winslow LA, Read JS, Hansen GJA, Rose KC, Robertson DM. 2017. Seasonality of change: summer warming rates do not fully represent effects of climate change on lake temperatures. Limnol Oceanogr 62: 2168–2178.
- Woods T, Kaz A, Giam X. 2021. Phenology in freshwaters: a review and recommendations for future research. Ecography 44: 1–14.
- Woolway RI. 2023. The pace of shifting seasons in lakes. Nat Commun 14: 2101.
- Woolway RI, Sharma S, Weyhenmeyer GA, Debolskiy A, Golub M, Mercado-Bettín D, Perroud M, Stepanenko V, Tan Z, Grant L. 2021. Phenological shifts in lake stratification under climate change. Nat Commun 12: 1–11.
- Yankova Y, Neuenschwander S, Köster O, Posch T. 2017. Abrupt stop of deep water turnover with lake warming: drastic consequences for algal primary producers. Sci Rep 7: 1–9.
- Yule DL, Stockwell JD, Black JA, Cullis KI, Cholwek GA, Myers JT. 2008. How systematic age underestimation can impede understanding of fish population dynamics: lessons learned from a lake superior Cisco Stock. Trans Am Fish Soc 137: 481–495.
- Zeller D, Booth S, Pakhomov E, Swartz W, Pauly D. 2011. Arctic fisheries catches in Russia, USA, and Canada: baselines for neglected ecosystems. Polar Biol 34: 955–973.
- Zimmerman MS, Krueger CC. 2009. An ecosystem perspective on reestablishing native deepwater fishes in the Laurentian Great Lakes. North Am J Fish Manag 29: 1352–1371.

Cite this article as: Stewart TR, Karjalainen J, Zucchetta M, Goulon C, Anneville O, Vinson MR, Wanzenböck J, Stockwell JD. 2024. Winter is not coming: evaluating impacts of changing winter conditions on coregonine reproductive phenology. Int. J. Lim. 60: 17.