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► To cite this version:

Ameni Ben Abdennebi, Emna Chaabani, Soumaya Bourgou, Majdi Hammami, Moufida Saidani Tounsi, et al.. Fenugreek as a Potential Active Ingredient for the Development of Innovative Cosmetic Formulation. *Cosmetics*, 2025, 12 (1), pp.21. 10.3390/cosmetics12010021 . hal-04918443

HAL Id: hal-04918443

<https://hal.inrae.fr/hal-04918443v1>

Submitted on 29 Jan 2025

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Article

Fenugreek as a Potential Active Ingredient for the Development of Innovative Cosmetic Formulation

Ameni Ben Abdennebi¹, Emna Chaabani¹ , Soumaya Bourgou¹, Majdi Hammami¹, Moufida Saidani Tounsi¹, Othmane Merah^{2,3,*}  and Iness Bettaieb Rebey^{1,*}

- ¹ Laboratory of Aromatic and Medicinal Plants, Borj Cedria Biotechnology Center, BP. 901, Hammam-Lif 2050, Tunisia; benabdennebiameni9@gmail.com (A.B.A.); emna.chaabani@cbbc.rnrt.tn (E.C.); soumaya.bourgou@cbbc.rnrt.tn (S.B.); majdi.hammami@cbbc.rnrt.tn (M.H.); moufida.saidani@cbbc.rnrt.tn (M.S.T.)
- ² Laboratory of Agro-Industrial Chemistry (LCA), Toulouse University, National Research Institute for Agriculture, Food and Environment (INRAE), National Polytechnic Institute of Toulouse (INPT), 31030 Toulouse, France
- ³ Department of Biological Engineering, IUT Paul Sabatier, University Paul Sabatier, 32000 Auch, France
- * Correspondence: othmane.merah@ensiacet.fr (O.M.); iness.bettaieb@cbbc.rnrt.tn (I.B.R.)

Abstract: This study focuses on optimizing the extraction conditions for antioxidants from the fenugreek seeds (*Trigonella foenum-graecum* L.) through ultrasound-assisted extraction with the aim of creating a stable and effective cosmetic formulation. RSM was used to optimize the extraction parameters to ultrasonic power of 60%, with 50% ethanol concentration for 10 min. Under those conditions, the extract showed a phenolic-rich profile, with a total phenolic content equivalent to 18.56 mg GAE/g DM and a radical scavenging activity of 63.24%. Chromatographic analysis further confirmed the abundance of phenolic compounds, with epicatechin identified as the major compound at a concentration of 22.58 mg/g DM. The extract exhibited considerable antibacterial activity for a number of bacterial strains, and it exhibited no cell toxicity on RAW 267.4 cells, supporting its safe use in cosmetic products. The cosmetic formulation maintained high stability, with pH values from 6.25 to 6.35, viscosity values from 7941.69 to 7956.70 cp, and less color change after 90 days of preservation under varied temperature conditions. These findings validate fenugreek extract's potential for producing a stable, eco-friendly, and effective cosmetic product, thus bringing skin health benefits and driving sustainable extraction methods in the cosmetic industry.

Keywords: *Trigonella foenum graecum* L.; ultrasound-assisted extraction; epicatechin; radical scavenging activity; antibacterial activity; cytotoxicity; cosmetic formulation; skin health



Academic Editor: Vasil Georgiev

Received: 19 December 2024

Revised: 19 January 2025

Accepted: 21 January 2025

Published: 24 January 2025

Citation: Abdennebi, A.B.; Chaabani, E.; Bourgou, S.; Hammami, M.; Tounsi, M.S.; Merah, O.; Bettaieb Rebey, I.

Fenugreek as a Potential Active Ingredient for the Development of Innovative Cosmetic Formulation. *Cosmetics* **2025**, *12*, 21. <https://doi.org/10.3390/cosmetics12010021>

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1. Introduction

The skin, being the largest organ of the body, serves as a vital protective shield against harmful pathogens and substances, essentially acting as the body's first line of defense [1]. This protective role is shaped by both internal and external factors [2]. External influences include environmental conditions such as UV radiation, wind, humidity, and air conditioning, as well as physical damage and interactions with microorganisms like bacteria, fungi, viruses, and toxins. On the internal side, genetics, certain medications (like immunosuppressants and contraceptives), antibiotics, and diet all play a part. Furthermore, daily use of cosmetics, skincare products, detergents, soaps, and perfumes also has a notable impact on skin health [3], which can accelerate skin aging through the production of reactive oxygen species (ROS). Oxidative stress, exacerbated by UV exposure, increases

the activity of enzymes that degrade skin fibers, such as collagenase and elastase [4], thereby contributing to a loss of elasticity and tensile strength, leading to the appearance of wrinkles and increased dryness [4]. Oxidative stress also plays a key role in inducing inflammation [5], slowing down the cellular renewal of the epidermis and leading to a reduction in its thickness, which weakens the protective barrier [6]. UV radiation also triggers the production of free radicals (ROS), which are responsible for skin dehydration. To minimize skin damage, it is essential to reduce oxidative stress through the use of antioxidants. In recent years, interest in natural substances has significantly increased, particularly due to the growing distrust of synthetic products. Many industries, including the cosmetics industry, are now turning toward the incorporation of natural molecules in their formulations, as these offer unique biological and chemical properties [7]. Studies have shown that many plants contain a wide variety of active compounds, such as terpenoids, alkaloids, and phenols [8]. These compounds, by neutralizing free radicals, slow down premature skin aging. Polyphenols, in particular, protect the skin from damage caused by these free radicals, thus reducing the development of wrinkles and visible aging indicators while supporting a radiant and balanced skin tone [9].

Trigonella foenum-graecum L. is an herbaceous plant belonging to the Leguminosae family, primarily cultivated in Western Asia, North Africa, Northern India, and the Mediterranean region [10]. Its seeds are widely used as food, spice, galactagogue, and in traditional medicine for the treatment of diabetes [11]. Furthermore, a toxicological study has confirmed the safety of using fenugreek seeds as a dietary supplement [12]. These seeds contain various phytochemicals such as alkaloids, saponins, and flavonoids (rutin, quercetin, vitexin), as well as galactomannans, which contribute to skin hydration through a humectant effect [13]. Numerous studies have demonstrated the anticancer, antimicrobial, antioxidant, and anti-inflammatory properties of fenugreek [14]. Among its active compounds, epicatchin stands out for its ability to protect skin cells from damage caused by free radicals, thereby delaying premature aging and helping to prevent the appearance of wrinkles and fine lines [15]. The extraction of phenolic compounds from fenugreek seeds can be achieved through several techniques. In recent years, ultrasound-assisted extraction (UAE) has gained popularity due to its advantages, including more efficient extraction, minimal solvent use, reduced costs, and low environmental impact. UAE enhances extraction yields by facilitating the diffusion of active compounds through cavitation, a phenomenon where air bubbles rapidly form and collapse under the effect of ultrasound, generating local increases in pressure and temperature [16].

This study focuses on optimizing the ultrasound-assisted extraction (UAE) process for fenugreek seeds using response surface methodology (RSM) to maximize the recovery of phenolic compounds and their antioxidant activity. The application of RSM provides a systematic approach for evaluating and refining key extraction parameters to achieve high yields of bioactive substances with minimal environmental impact. The fenugreek extract obtained under optimal conditions was formerly evaluated for its antioxidant, antimicrobial, and anti-inflammatory activities. The extract was then incorporated into a cosmetic cream formulation, where its stability, bioactivity, and sensory properties were thoroughly assessed. By combining advanced extraction techniques with plant-derived bioactive compounds, this study contributes to the development of innovative skincare products that effectively target skin health while prioritizing environmental sustainability. The findings underscore the role of fenugreek extract as a safe and versatile component for next-generation cosmetic formulations.

2. Materials and Methods

2.1. Plant Material

Fenugreek, or *Trigonella foenum-graecum* L., is a plant renowned for its seeds and numerous uses. These seeds were purchased in dried form from spice merchants, particularly from the Beja province.

2.2. Ultrasound-Assisted Recovery of Bioactive Antioxidants

The ultrasound-assisted extraction was conducted using an ultrasonic bath (Sonorex Digital 10 P, Bandelin, GmbH, Berlin, Germany) with a hydro-ethanolic solvent mixture prepared in varying ratios. Following each extraction, the solution was subjected to centrifugation at 5000 rpm for 10 min. The supernatant, which contained the extracted antioxidants, was then collected for further analysis [17].

2.3. Design of Experiments and Statistical Analysis

We designed an experimental plan using the NemrodW (LPRAI, version 2000) software to optimize the experimental conditions for extracting antioxidants from fenugreek and increase this extraction using ultrasound. With three independent variables and two responses, this design consists of 19 trials arranged using the Box–Behnken model (Table 1).

Table 1. Box–Behnken design.

Exp	Independent Variables			Responses	
	Time (min)	Ultrasonic Power (%)	EtOH/H ₂ O Ratio (% v/v)	Y _{TPC} (mg GAE/g DM)	Y _{PI} (%)
1	5	50	25	18.53	63.70
2	5	70	25	13.85	51.63
3	5	50	75	5.00	35.05
4	5	70	75	6.85	52.61
5	15	50	25	15.07	63.05
6	15	70	25	11.77	51.38
7	15	50	75	9.33	39.34
8	15	70	75	18.68	63.85
9	10	50	50	19.20	62.12
10	10	70	50	19.20	66.21
11	10	60	25	14.57	56.57
12	10	60	75	9.12	50.82
13	5	60	50	15.62	55.84
14	15	60	50	19.75	54.58
15	10	60	50	18.28	58.84
16	10	60	50	19.62	60.42
17	10	60	50	19.40	61.70
18	10	60	50	19.02	60.71
19	10	60	50	19.05	59.17

This plan consists of 19 experiments with 3 independent variables and 2 responses, arranged using a Box–Behnken model (Table 1). The factors investigated include ultrasonic power (X_1), ethanol % (X_2), and extraction time (X_3). Their respective central values are 60%, 50%, and 10 min. The variation steps selected are 10% for X_1 , 25% for X_2 , and 5 min for X_3 . A second-degree polynomial model was used to analyze the change in DPPH free radical scavenging activity (Y_{PI}) and total phenolic content (Y_{TPC}) with respect to the three variables that were chosen, X_1 , X_2 , and X_3 , as indicated in the equation below:

$$Y_i = \beta_0 + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3 + \beta_{1,1}(X_1 X_1) + \beta_{2,2}(X_2 X_2) + \beta_{3,3}(X_3 X_3) + \beta_{1,2}(X_1 X_2) + \beta_{1,3}(X_1 X_3) + \beta_{2,3}(X_2 X_3)$$

- Y_i denotes the observed response variables;
- β_0 is a constant;
- β_1 , β_2 , and β_3 are linear coefficients for X_1 , X_2 , and X_3 , respectively;
- $\beta_{1,1}$, $\beta_{2,2}$, and $\beta_{3,3}$ are coefficients for the quadratic terms $X_1 X_1$, $X_2 X_2$, and $X_3 X_3$, respectively;
- $\beta_{1,2}$, $\beta_{1,3}$, and $\beta_{2,3}$ are coefficients for the interaction terms $X_1 X_2$, $X_1 X_3$, and $X_2 X_3$, respectively.

The statistical analysis was performed to ensure the robustness and reliability of the experimental results, particularly in evaluating the fenugreek extract's effects on microbial growth. Analysis of variance (ANOVA) was used to assess the significance of the independent variables (ultrasonic power, ethanol concentration, and extraction time) and their interactions with the observed responses. ANOVA was conducted with IBM SPSS Statistics (Version 20.0, IBM SPSS Inc., Armonk, NY, USA), followed by Duncan's multiple range test to determine significant differences among means at $p < 0.05$. Statistical tests were applied to data obtained from triplicate experiments.

2.4. Total Phenolic and Flavonoid Contents

The total polyphenol content (TPC) was determined by spectrophotometric analysis using the Folin–Ciocalteu method. The aluminum chloride colorimetric method was used to determine the total flavonoid content of the sample, in accordance with the procedure described by condensed tannins were also measured by subjecting them to depolymerization in the presence of sulfuric acid, followed by a reaction with vanillin [17].

2.5. Chromatographic Phenolic Composition Assessment

Phenolic compounds were identified and quantified using an Agilent 1260 system (Agilent Technologies, Waldbronn, Germany) equipped with a photodiode array detector. Separation was achieved on a Zorbax Eclipse XDB C18 reverse-phase column (Agilent Technologies, Santa Clara, CA, USA), maintained at 25 °C. The mobile phase consisted of HPLC-grade water with 0.1% formic acid (solvent A) and acetonitrile (solvent B) at a flow rate of 0.7 mL/min. The gradient was programmed as follows: 90% A/10% B (0–40 min), 50% A/50% B (40–41 min), 100% B (41–50 min), and 90% A/10% B (50–59 min). Retention times for each compound were annotated to facilitate cross-verification. Quantification was performed using standards at 280 nm, with calibration curves constructed for concentrations from 10 to 1000 µg/mL. Analyses were conducted in triplicate, and results were reported as mg/g of extract.

2.6. Biological Activities

2.6.1. Antioxidant Activities

The evaluation of antioxidant activity was carried out using four distinct methods. First, the total antioxidant capacity was determined by the reduction of molybdenum (Mo^{6+}) to molybdenum (Mo^{5+}) [18]. The antiradical activity was then assessed using the DPPH and ABTS assays, which measure the extracts' ability to neutralize DPPH and ABTS free radicals, with results expressed as a percentage of inhibition [19]. The reducing power of the extracts was evaluated using the ferricyanide method, where the reduction of ferric iron (Fe^{3+}) to ferrous iron (Fe^{2+}) served as an indicator of antioxidant potential. The effective concentration (EC_{50} µg/mL) required to achieve an absorbance of 0.5 at 700 nm was calculated [19]. All tests were conducted in triplicate.

2.6.2. Antibacterial Activity

Assessment of Eco-Extract via Disc Diffusion Technique

The antibacterial efficacy of the fenugreek eco-extract was assessed using the agar diffusion method against several pathogenic bacteria, including *Enterococcus faecalis* (ATCC 29212), *Staphylococcus aureus* (ATCC 25923), *Escherichia coli* (ATCC 8739), and *Salmonella typhimurium* (ATCC 14028). A single colony from each pure culture was transferred to physiological saline. The pathogenic strains were inoculated at 100 µL (10⁸ CFU/mL) into 10 mL of soft agar, which was then overlaid onto a Petri dish containing 50 mL of MH agar [20]. Discs of 6 mm in diameter were placed on the agar surface and loaded with 10 µL of the extract dissolved in 10% DMSO. For determining the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC), serial dilutions of the green fenugreek extract in a 10% aqueous DMSO solution were added to wells inoculated with 10 µL of a bacterial suspension (10⁶ CFU/mL) and Mueller Hinton broth. The choice of 10% DMSO as a solvent was guided by its ability to efficiently solubilize phenolic and flavonoid compounds while remaining non-toxic to the cells and microorganisms tested. The plates were first incubated at 4 °C for 2 h, then at 37 °C for 24 h. The discs designated for positive control were loaded with Streptomycin, while those intended for negative control were impregnated with DMSO. The zones of inhibition of bacterial growth were measured to assess the antibacterial effectiveness of the extract.

Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC)

For determining the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC), serial dilutions of the green fenugreek extract in a 10% aqueous DMSO solution were added to wells inoculated with 10 µL of a bacterial suspension (10⁶ CFU/mL) and Mueller Hinton broth. After overnight incubation at 37 °C, the MIC was determined by the highest dilution, showing no growth. For the MBC, 200 µL from wells with no growth were plated on agar and incubated overnight at 37 °C, with the highest dilution showing no bacterial colonies recorded as the MBC [21].

2.6.3. Anti-Inflammatory Activity

The RAW 264.7 murine macrophage cell line was obtained from the American Type Culture Collection (ATCC) and cultured in Dulbecco's Modified Eagle Medium (DMEM) under standard conditions. After a 24 h incubation period to allow for cell adhesion, the medium was replaced with fresh DMEM containing different concentrations of the extracts. The cells were then incubated for an additional 24 h.

Cell viability was assessed using the resazurin assay [22], and the fluorescence intensity (F) was measured. Cell viability was calculated using the following formula:

$$\text{Cellviability (\%)} = \frac{F_{\text{sample}}}{F_{\text{control}}} \times 100$$

where F_{sample} represents the fluorescence of treated cells, and F_{control} represents the fluorescence of untreated control cells.

The anti-inflammatory activity of the extracts was assessed using RAW 264.7 cells stimulated with lipopolysaccharide (LPS). Cells were seeded in 24-well plates at a density of 2×10^4 cells/mL and incubated for 24 h to allow for adhesion. The cells were then pretreated with varying concentrations of the extracts (25–200 µg/mL) for 1 h, followed by stimulation with 1 µg/mL LPS for an additional 24 h. After incubation, the culture supernatants were collected and mixed with an equal volume of Griess reagent. The mixture was incubated at room temperature for 10 min to allow for color development.

Absorbance was measured at 540 nm using a microplate reader. Nitrite levels, indicating nitric oxide (NO) production, were quantified using a sodium nitrite (NaNO_2) standard curve. Statistical significance ($p < 0.05$) was evaluated to confirm the dose-dependent inhibition of nitrite production by the extracts. The half-maximal inhibitory concentration (IC_{50}) for each extract was determined [17,22]. All experiments were conducted in triplicate.

2.7. Cosmetic Cream Formulation

The formulated cream is an oil-in-water (O/W) emulsion, where the oil phase is dispersed within the aqueous phase. The preparation of this cream involves two distinct steps. First, the aqueous phase, which constitutes 70.6% of the formula, is prepared by dissolving xanthan gum, a co-emulsifier, in distilled water. Simultaneously, the oil phase is created by combining 17% refined sweet almond oil with 8.5% stearic glycerides, which act as an emulsifier. Both phases are prepared in a water bath at approximately 70 °C with continuous stirring. After the preparation of the two phases, they are mixed using a helical disperser at 1300 rpm for 10 min until the mixture cools down to 40 °C. Finally, the active ingredient and preservative are added, and the mixture is stirred for an additional 10 min [17,22].

2.8. Stability Testing Evaluation

A stability assessment was conducted by storing the formulation at 4 °C, 25 °C, and 40 °C for 90 days. Centrifugation was conducted at 3000 rpm for 30 min at room temperature, followed by macroscopic analysis to assess the appearance and uniformity of the sample. pH was measured with a pH meter at a controlled temperature of 25 ± 2 °C. Color analysis was performed using a portable colorimeter (PCE-XXM 30, PCE Instruments, Meschede, Germany), and viscosity was measured with a rotary viscometer (VISCOSIMETER PCE-RVI 2, PCE Instruments, Sultz-sous-Forêts, GR, France).

2.9. Sensory Analysis

Sensory evaluation was performed with a group of 60 participants to evaluate the cream formulation. The analysis focused on several factors, including color, fragrance, texture, spreadability, stickiness, and absorption. Participants rated each aspect on a scale from 1 to 10. Following the evaluation, an aggregate score was calculated for the cream based on the ratings of all parameters. This sensory analysis provided insights into user perceptions and an overall evaluation of the cosmetic cream [23].

3. Results and Discussion

3.1. Design of Experiment

Response Surface Methodology (RSM) was utilized to optimize the extraction process by evaluating the effects of extraction time (X_1 , min), ultrasonic power (X_2 , %), and the EtOH/H₂O percentage (X_3 , % v/v). These factors were examined for their impact on two key response variables: total phenolic content (Y_{TPC}) and DPPH free radical scavenging activity (Y_{PI}). Based on initial experimental data, the study focused on specific ranges for each factor: extraction time (5 to 15 min), ultrasound power (50 to 70%), and ethanol concentration (25 to 75%). The results for total phenolic content (Y_{TPC}) and DPPH scavenging activity (Y_{PI}) from the 19 experiments are presented in Table 1. The TPC ranged from 5.00 mg GAE/g DM to 19.75 mg GAE/g DM and the PI from 35.05% to 66.21 %.

3.2. Interpretation of Coefficients

The significance of the model coefficients was evaluated using analysis of variance (ANOVA). Table 2 indicates the considerable effect of studied coefficients on respective

response variables. The coefficients of the second-degree polynomial model for both responses (Y_{PI} , Y_{TPC}) were evaluated using Student's t -test with a 95% confidence interval ($\alpha = 0.05$). A coefficient is deemed significant when the p -value is below 0.05. Table 2 presents the coefficient values for both models. The results show that ultrasonic power (X_1), ethanol percentage (X_2), and extraction time (X_3) all significantly affect Y_{PI} ($p < 0.05$), while only X_2 and X_3 impact Y_{TPC} . Additionally, all three factors influence the DPPH test (Y_{PI}) quadratically, whereas only X_2 affects Y_{TPC} quadratically. Significant interactions were found between ultrasonic power and extraction time ($X_1 - X_3$) and ethanol percentage and extraction time ($X_2 - X_3$). Higher coefficient values indicate more significant factors, with the $X_1 - X_3$ interaction notably affecting Y_{TPC} .

Table 2. Analysis of variance (ANOVA) of second-order polynomial models for the two responses (Y_{TPC} , Y_{PI}).

Source of Variation	Sum of Squares	Degrees of Freedom	Mean Squares	Fisher's F-Test	Significance
Y_{PI}					
Regression	1.17486	9	1.30540	43.5432	***
Validity	2.15036	5	4.30071	3.1404	14.5%
$R^2 = 0.978$			$F_{Obs} (43.54) > F_{tab} (3.18)$		
Y_{TPC}					
Regression	402.59	9	44.7329	48.2308	***
Validity	7.3166	5	1.4633	5.6794	6.0%
$R^2 = 0.980$			$F_{Obs} (48.23) > F_{tab} (3.18)$		

***: very significant.

3.3. Model Validation via ANOVA Analysis

The statistical significance of the regression equation was checked by Fisher's F-test (Table 2). According to Fisher's F-test, the observed F-ratios, which are the ratios of the mean square of the regression to the mean square of the residuals, are higher than the tabulated values ($[F_{observed} = 43.5432] PI > F_{tabulated} = 3.18$; $[F_{observed} = 48.2308] TPC > F_{tabulated} = 3.18$). Furthermore, the ratio between the mean square of validity and the estimated experimental variance is lower than the tabulated value, confirming the results of Fisher's test and the validity of the proposed model. The R^2 coefficients for the responses Y_{PI} and Y_{TPC} are 0.978 and 0.980, respectively, indicating a strong correlation between the experimental values and those predicted by the models.

3.4. Analysis of Response Surface Curves

The isoresponse curves, displayed in both 2D and 3D, were utilized to visually represent the mathematical models that describe the impact of significant factors on the PI and TPC of the extract. Figure 1 illustrates the effects of interactions between key independent variables, such as EtOH/H₂O percentage (X_2) and extraction time (X_3), on the total phenolic content of the fenugreek extract. With ultrasonic power set at 60%, the total polyphenol content exceeds 19.75 mg GAE/g DW when the EtOH/H₂O percentage ranges from 25% to 50%, and the extraction time spans 10 to 15 min. Regarding the DPPH test, Figure 1 also shows how the interactions between ultrasonic power (X_1) and EtOH/H₂O percentage (X_2) influence the ability of the fenugreek extract to neutralize the DPPH radical. Specifically, when the extraction time is held at 10 min, DPPH radical inhibition can reach up to 60.54% with ultrasonic power levels between 50% and 60% and EtOH/H₂O ratios from 25% to 50%.

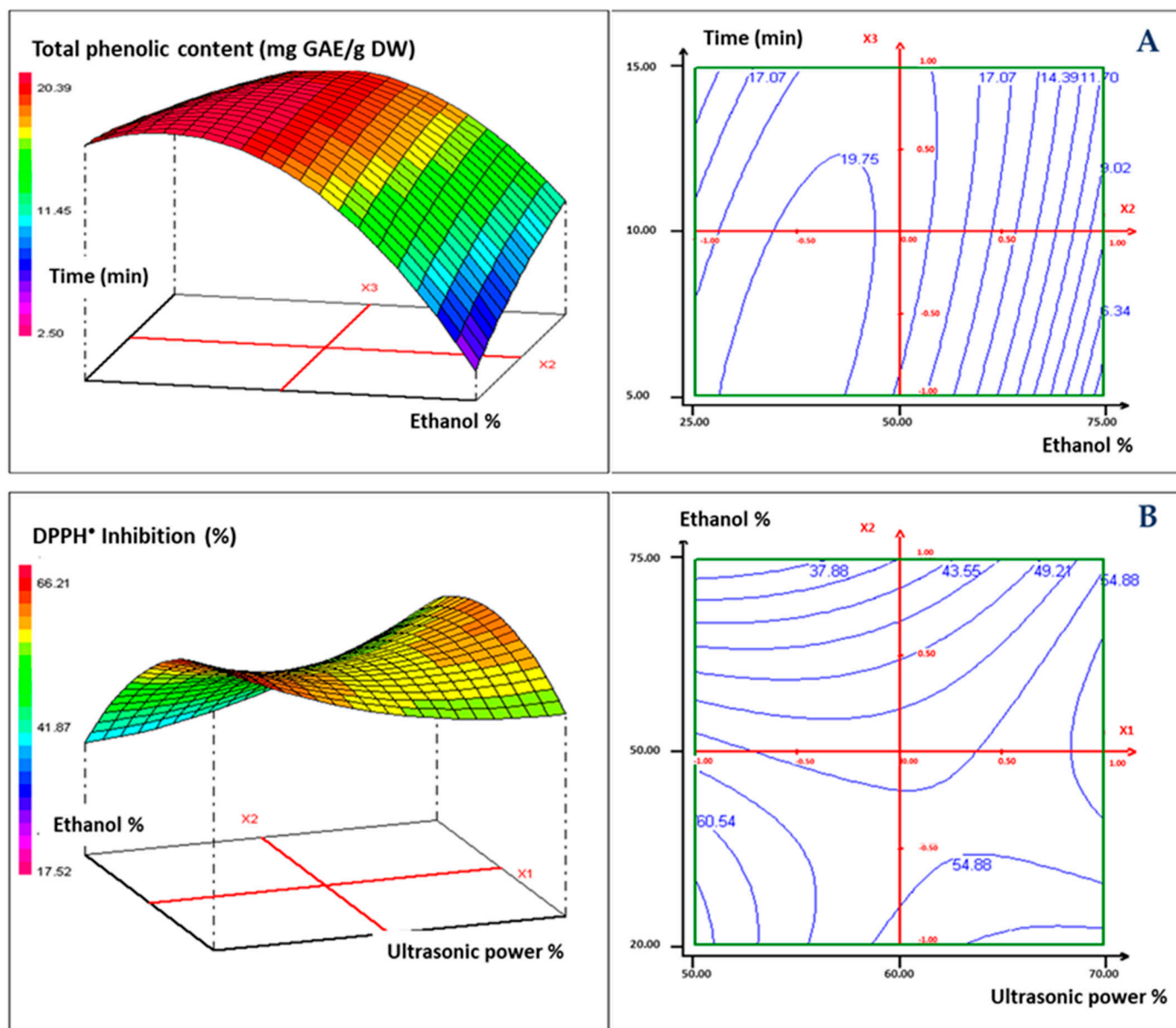


Figure 1. (A) The 2D and 3D response surface curves showing the effect of interaction between extraction time (X_1) and EtOH/ H_2O percentage (X_2) on TPC. (B) The 2D and 3D response surface curves showing the effect of the interaction between time (X_1) and EtOH/ H_2O percentage (X_2) on PI.

3.5. Optimization of Antioxidant Extraction Conditions from Fenugreek

The optimization of the ultrasound-assisted extraction (UAE) process for fenugreek seeds was achieved using response surface methodology (RSM). The optimal extraction conditions included an EtOH/ H_2O percentage of 50%, an extraction time of 10 min, and an ultrasonic power of 60% (Table 3). These parameters yielded experimental values for DPPH radical scavenging activity of 63.24% and total phenolic content (TPC) of 18.56 mg GAE/g DM. The close alignment of these experimental values with the RSM-predicted values (60.20% for DPPH and 18.87 mg GAE/g DM for TPC) underscores the accuracy of the optimization process (Table 3). These results align with those reported by Dastan et al. [24], who optimized fenugreek seed extraction using ultrasound and a solvent/water mixture (50/50), obtaining a similar total phenolic content of 16.96 mg GAE/g DW. Furthermore, Yang et al. [25], using an ultrasound-assisted extraction method with slightly different parameters (72% ethanol, an extraction time of 41 min, and a solvent-to-material ratio of 35 mL/g), reported a high radical scavenging activity of 80.33%. These observations corrob-

orate our findings, reinforcing the reliability of the applied method. The results confirm that UAE effectively enhances the extraction of phenolic compounds and antioxidant activity.

Table 3. Experimental and predicted values of the responses analyzed under optimal conditions.

Ultrasound Power (%)	Factor		Experimental Value		Predicted Value	
	EtOH/H ₂ O Percentage (%v/v)	Time (min)	PI (%)	TPC (mgGAE/gDM)	PI (%)	TPC (mgGAE/gDM)
60	50	10	63.24 ± 0.83	18.56 ± 0.03	60.20	18.78

Ultrasound-assisted extraction (UAE) was chosen for its ability to efficiently extract bioactive compounds like polyphenols from plant materials by enhancing solvent penetration and promoting cavitation [26]. This method is faster and more energy-efficient than traditional techniques. In this study, a 50% ethanol solution was used to extract both hydrophilic and lipophilic components. The ultrasonic power was optimized to ensure effective cavitation without damaging the compounds, and the extraction time was controlled to maximize yield and preserve compound integrity. These parameters balance extraction efficiency, stability, and minimal solvent usage, making UAE ideal for cosmeceutical applications [26].

The use of RSM allowed for the optimization of these parameters to achieve maximal phenolic content and antioxidant activity while minimizing variability in microbial assays. This approach is particularly critical when assessing antimicrobial effects, as the growth of microorganisms can be influenced by subtle variations in the experimental setup, such as the solvent concentration or bioavailability of active compounds. By applying RSM and ANOVA, the study ensured that the observed microbial inhibition was directly attributable to the bioactive properties of the extract under optimized conditions rather than extraneous factors.

3.6. Phenolic Content and Antioxidant Activities in Fenugreek Extract

In this study, after optimizing the experimental parameters to maximize phenolic compound content and antioxidant activity, the following conditions were established: 60% ultrasonic power, 50% hydro-ethanolic solution, and a 10 min extraction time. These optimized parameters were consistently applied in all subsequent analyses. The spectrometric evaluation of *T. foenum-graecum* (fenugreek) eco-extract revealed its substantial phenolic content and antioxidant activity (Table 4), emphasizing its potential as a rich source of bioactive compounds for diverse applications. Specifically, the total phenolic content (TPC) of the eco-extract, measured at 18.56 mg GAE/g DM, highlights the presence of bioactive molecules known for their capacity to mitigate oxidative damage, a key factor in skin aging and deterioration through well-documented mechanisms such as free radical scavenging and metal chelation [27]. The total phenolic content of fenugreek eco-extract (18.56 mg GAE/g DM) is lower compared to other natural extracts. For example, the leaves of *Arbutus unedo* exhibit a significantly higher phenolic content, reaching 76.74 mg GAE/g DW, as reported by Habachi et al. [22], through ultrasound-assisted extraction. Similarly, raspberry leaves contain 26.2 mg GAE/g DW, which is higher than that of fenugreek [27], demonstrating the varying levels of polyphenolic content across different plant extracts. Phenolic compounds are particularly valued for their hydrogen-donating properties, which make them highly effective against free radicals [28].

Table 4. Phenolic contents and antioxidant activities of fenugreek eco-extract.

Assay	Values
TPC (mg EAG/g DM)	18.56 ± 0.54
TFC (mg EC/g DM)	19.26 ± 1.63
TCT (mg EC/g DM)	10.35 ± 0.72
TAC (mg/mL)	218.75 ± 1.87
DPPH (µg/mL)	CI ₅₀ = 77.17 ± 0.75
ABTS (µg/mL)	CI ₅₀ = 93.69 ± 0.86
RP (µg/mL)	CE ₅₀ = 131.77 ± 5.88

Moreover, the total flavonoid content (TFC) was determined to be 19.26 mg CE/g DM, further emphasizing the extract's rich flavonoid profile. Flavonoids are particularly valued in cosmetic formulations for their photoprotective, anti-inflammatory, and antioxidant properties [29]. Their presence in the extract enhances its ability to protect the skin from environmental stressors, such as UV radiation, and combat visible signs of aging, including wrinkles and fine lines [30]. Additionally, the presence of condensed tannins (10.35 mg EC/g DM) strengthens the overall antioxidant capacity of the extract. Tannins have been shown to provide supplementary antioxidant effects by effectively scavenging free radicals and inhibiting oxidative damage [31].

Similarly, the antioxidant activity of the fenugreek seed eco-extract was evaluated using four complementary methods: total antioxidant capacity (TAC), DPPH radical scavenging assay, ABTS radical scavenging assay, and ferric-reducing antioxidant power (FRAP) test (Table 4).

The total antioxidant capacity (TAC) of the extract was determined, yielding a value of 218.75 mg GAE/mL. The DPPH and ABTS assays assessed the extract's ability to donate hydrogen atoms or electrons to stabilize free radicals. The results revealed IC₅₀ values of 77.17 µg/mL for DPPH and 93.69 µg/mL for ABTS, indicating strong radical scavenging activity. These values are comparable to those reported for other phenolic-rich plant extracts, such as green tea and rosemary, which are known for their antioxidant properties [32]. In addition, the FRAP assay demonstrated the extract's reducing power, with an EC₅₀ value of 131.77 µg/mL, highlighting its ability to reduce Fe³⁺ to Fe²⁺. This activity is attributed to flavonoids and condensed tannins, compounds known for their potent antioxidant properties. These compounds help chelate metal ions and prevent oxidative chain reactions, further enhancing the extract's ability to combat oxidative stress [33].

The combination of high phenolic content and antioxidant activity positions fenugreek extract as a promising candidate for cosmetic applications, particularly in anti-aging and skin-rejuvenating products. The antioxidant capacity, validated by experimental data and supported by the predictive models, highlights the extract's ability to protect the skin from oxidative damage caused by environmental stressors, UV exposure, and pollution. This also suggests that fenugreek extract may help in reducing signs of premature aging, such as wrinkles and loss of skin elasticity [34]. Additionally, the validation experiments confirm the robustness of the optimized extraction process, supporting the use of ultrasound-assisted extraction as a sustainable and effective method for producing bioactive compounds. The consistency of the results obtained from the validation experiments strengthens the potential of fenugreek extract as an eco-friendly and effective ingredient for next-generation cosmetic formulations. Incorporating fenugreek extract into skincare products offers enhanced antioxidant protection, promotes skin health, and reduces visible signs of aging, aligning with current consumer trends toward natural, sustainable, and high-performance cosmetic ingredients [34].

3.7. Phytochemicals Identification by RP-HPLC

To further understand the bioactive potential of the fenugreek extract, the specific phenolic compounds responsible for its antioxidant activity were identified and quantified using reverse-phase high-performance liquid chromatography (RP-HPLC). The analysis of the 50% hydro-ethanolic extract of *T. foenum-graecum* L. by RP-HPLC highlights the extract's rich phenolic profile and reinforces its antioxidant potential (Figure 2).

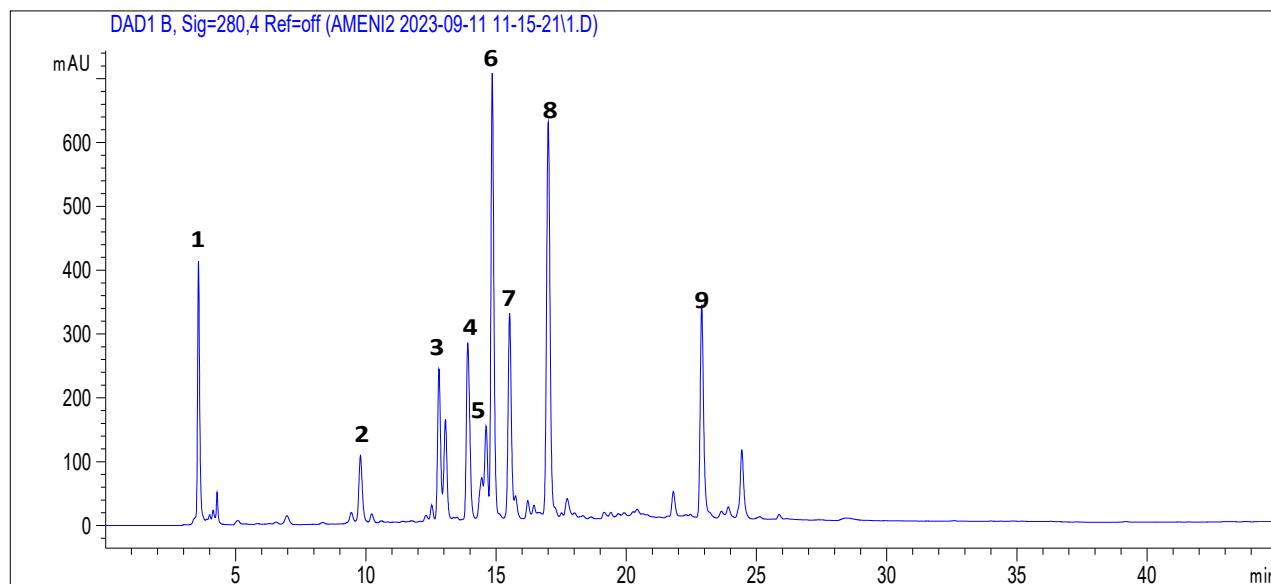


Figure 2. HPLC-DAD profile of fenugreek seed eco-extract. 1. Ascorbic acid (rt: 3.57), 2. ferulic acid (rt: 9.79), 3. catechin (rt: 12.80), 4. catechol (rt: 13.05), 5. caffeic acid (rt: 13.91), 6. epicatechin (rt: 14.85), 7. myricetin (rt: 15.51), 8. luteolin (rt: 22.89), 9. apigenin (rt: 24.43); rt: retention time.

Among the nine identified compounds, epicatechin emerged as the predominant phenolic compound (Table 5), with a concentration of 22.58 mg/g DM, followed by catechin (8.21 mg/g DM), vanillic acid (9.85 mg/g DM), myricetin (6.73 mg/g DM), and luteolin (5.6 mg/g DM). Additional compounds such as ferulic acid, apigenin, and caffeic acid further enriched the extract's bioactivity.

Table 5. HPLC analysis of optimal *T. foenum-graecum* L. extract.

Compounds	mg/g DW	Calibration Curve	R ²
Ferulic acid	0.81 ± 0.05	Y = 20.505x − 8.728	1
Vanillic acid	9.85 ± 0.08	Y = 9.02x − 1.55	0.995
Catechol	5.74 ± 0.01	Y = 3.632x + 1.8	0.998
Caffeic acid	1.74 ± 0.02	Y = 23.496x + 5.57	0.999
Myricetin	6.73 ± 0.05	Y = 6.7915x − 35.35	0.994
Epicatechin	22.58 ± 0.01	Y = 3.632x + 1.8	0.998
Catechin	8.21 ± 0.04	Y = 3.632x + 1.8	0.998
Luteolin	5.6 ± 0.02	Y = 7.4296x + 13.16	0.996
Apigenin	1.35 ± 0.05	Y = 12.418x + 59.908	0.997

Epicatechin, known for its broad therapeutic properties, including antioxidant, anti-inflammatory, neuroprotective, and anti-aging effects, plays a pivotal role in mitigating oxidative stress, preventing collagen degradation, and enhancing skin elasticity [35]. These attributes make epicatechin, alongside other phenolic compounds like catechin and luteolin, invaluable for natural skincare formulations [36]. The phenolic composition underscores

the extract's potential not only as a natural antioxidant but also as a bioactive agent for anti-aging and skin-rejuvenating applications. This aligns with growing consumer and industry trends favoring plant-derived, eco-friendly ingredients for cosmetics.

3.8. Antibacterial Activity

The antibacterial efficacy of the fenugreek seed eco-extract was demonstrated against four pathogenic bacterial strains, including *Enterococcus faecalis*, *Staphylococcus aureus*, *Escherichia coli*, and *Salmonella typhimurium*. The tests were performed in triplicate for each strain, and the extract exhibited a concentration-dependent inhibitory effect, as shown by increasing inhibition zone diameters with higher extract concentrations (Table 6). The minimum inhibitory concentration (MIC) values were consistent at 50 mg/mL for all strains except *E. coli*, which required a slightly higher bactericidal concentration (MBC) of 70 mg/mL.

Table 6. Results of the antibacterial activity of *T. foenum graecum* L. eco-extract.

Gram	ATCC	Strains	MIC (mg/mL)	MBC (mg/mL)
Gram +	29212	<i>Enterococcus faecalis</i>	50	50
Gram +	25923	<i>Staphylococcus aureus</i>	50	50
Gram –	3739	<i>Escherichia coli</i>	50	70
Gram –	14028	<i>Salmonella typhimurium</i>	50	50

These findings highlight the broad-spectrum antibacterial activity of the extract, effective against both Gram-positive (*E. faecalis*, *S. aureus*) and Gram-negative (*E. coli*, *S. typhimurium*) bacteria. This dual activity may be attributed to the presence of bioactive phenolic compounds, such as epicatechin, catechin, and vanillic acid, which have been previously reported to disrupt bacterial cell membranes and interfere with essential metabolic pathways [36]. Specifically, epicatechin is known to cause oxidative damage to bacterial cells by generating reactive oxygen species (ROS), leading to membrane integrity loss [37].

3.9. Evaluation of Anti-Inflammatory Activity

3.9.1. Evaluation of Cytotoxicity of Fenugreek Eco-Extract

The cytotoxicity of the fenugreek eco-extract was assessed using the resazurin assay on RAW 264.7 macrophage cells at varying concentrations ranging from 50 to 200 µg/mL (Figure 3). The resazurin assay is a widely used method to evaluate cell viability, as it measures the metabolic activity of cells [22]. The extract did not induce any significant decrease in cell viability, even at the highest concentration tested (200 µg/mL). In fact, at a low concentration of 50 µg/mL, cell viability increased by 32% compared to the control, suggesting a potential stimulatory effect on cell growth. These results indicate that the fenugreek eco-extract is non-toxic at the tested concentrations, and it does not exhibit cytotoxic effects on RAW 264.7 cells. This finding is consistent with previous studies on fenugreek extracts, which have shown no cytotoxicity up to concentrations of 250 µg/mL toward the J744.A1 rat macrophage cell line [38]. Consequently, the extract's safety profile supports its potential for use in cosmetic formulations, where it could promote skin health without adverse effects on cellular viability.

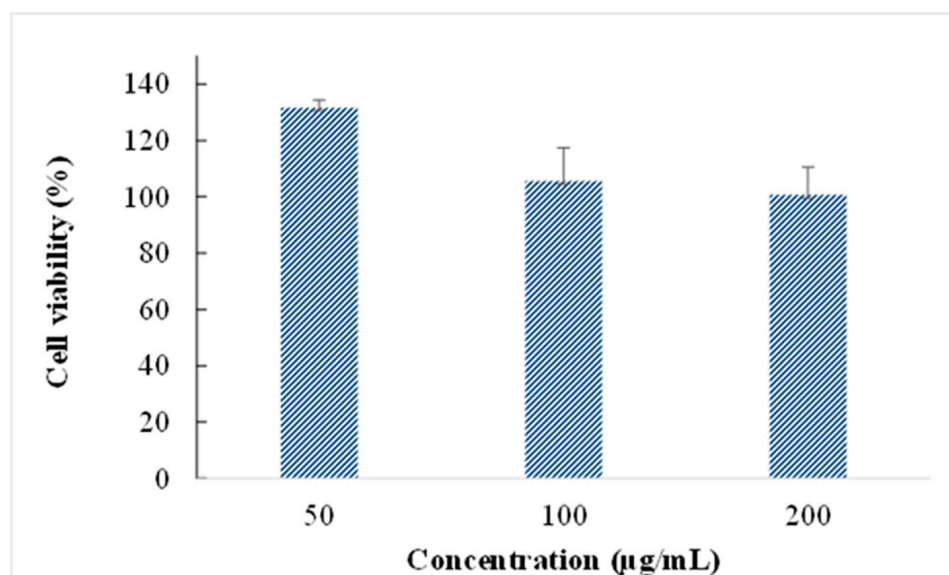


Figure 3. The viability of Raw 264.7 murine macrophage cells treated with *T. foenum graecum* L. eco-extract.

3.9.2. Measurement of Nitrite Production (NO)

To evaluate the anti-inflammatory potential of the fenugreek eco-extract, nitrite production was measured in RAW 264.7 macrophage cells stimulated with lipopolysaccharide (LPS). Nitrite, a stable product of nitric oxide (NO), serves as an indicator of inflammatory responses, particularly the activation of inducible nitric oxide synthase (iNOS) by LPS [39]. After treating the cells with increasing concentrations of the extract (ranging from 25 to 200 µg/mL) for 1 h, followed by stimulation with 1 µg/mL LPS for 24 h, the nitrite levels in the culture supernatants were quantified using the Griess reagent assay. The results showed a statistically significant ($p < 0.05$) dose-dependent reduction in nitrite production, with the extract demonstrating consistent inhibition of NO production. At concentrations of 100 µg/mL and 200 µg/mL, nitrite production was reduced by 34% and 48%, respectively, compared to the control group (Figure 4). These findings suggest that the fenugreek eco-extract has anti-inflammatory activity, likely through the inhibition of iNOS expression and NO production. This inhibition is important for modulating the inflammatory response and protecting against inflammation-related skin damage. Recent advances in fenugreek seeds have shed light on their antioxidative and anti-inflammatory activities, making them attractive candidates for applications in cosmeceuticals. Notably, the study by Eaknai et al. [34] explored the anti-aging potential of ethanolic fenugreek extracts, identifying a significant collagenase inhibitory activity ($IC_{50} = 0.57$ mg/mL), which was 2.6 times more effective than vitamin C ($IC_{50} = 1.46$ mg/mL).

To contextualize the efficacy of the fenugreek eco-extract, it is useful to compare its anti-inflammatory effects with standard drugs. Non-steroidal anti-inflammatory drugs (NSAIDs) like ibuprofen and diclofenac typically reduce NO production by 40–60%, while corticosteroids can suppress it by over 70% [40]. The fenugreek extract's 48% reduction in NO production at 200 µg/mL suggests its potential for inflammation modulation, possibly comparable to or better than some conventional treatments.

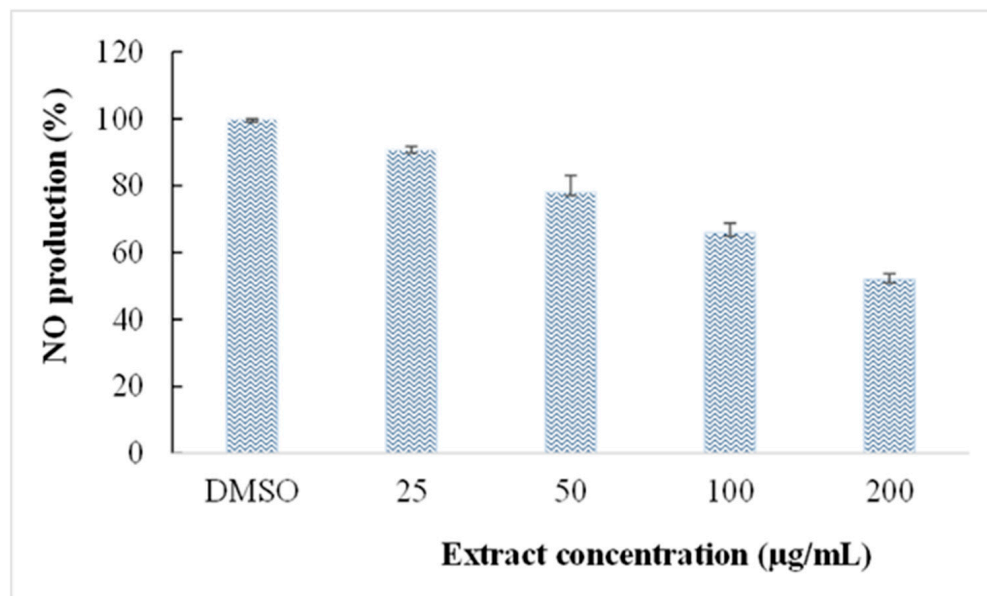


Figure 4. Anti-inflammatory activity of *T. foenum graecum* L. eco-extract.

3.10. Incorporation of Fenugreek Eco-Extract into Cosmetic Cream Formulation

Through ultrasonic-assisted extraction and design of experiments, we obtained a fenugreek extract with high levels of antioxidant compounds with potential antimicrobial and anti-inflammatory properties, making it a promising ingredient for cosmetic formulations. The extract's high content of phenolic compounds such as epicatechin, catechin, and luteolin provides powerful antioxidant effects, which help combat oxidative stress, a major contributor to skin aging [36]. These findings align with previous studies highlighting the benefits of fenugreek in promoting skin elasticity, reducing wrinkles, and enhancing skin radiance [34].

In addition to its potent antioxidant activity, the extract exhibits significant anti-inflammatory properties, as demonstrated by its ability to suppress nitric oxide (NO) production in macrophage cells. These findings suggest that the extract holds promise for managing inflammatory skin diseases. Furthermore, its lack of cytotoxic effects reinforces its safety and suitability for use in skincare formulations. Previous studies have demonstrated the anti-inflammatory properties of fenugreek [38,41]. In addition, the use of ultrasound-assisted extraction (UAE) also ensures that the bioactive compounds are efficiently recovered in an eco-friendly manner, aligning with sustainable production practices and green chemistry principles [42].

Based on these promising results, we have formulated an anti-aging and anti-wrinkle cream featuring the active eco-extract of *T. foenum-graecum* L. as the key ingredient.

3.11. Analysis and Stability of the Cosmetic Cream

The stability of the cosmetic cream containing fenugreek eco-extract was assessed over a 90-day period under different storage conditions (4 °C, 25 °C, and 40 °C) (Table 7). This comprehensive evaluation included physical property measurements such as pH, viscosity, and color, as well as centrifuge stability to ensure the cream's consistency and quality during storage. Throughout the storage period, the pH of the cream remained stable, ranging from 6.25 to 6.35, which is within the optimal range for skin care formulations. Maintaining a stable pH is critical, as it ensures the safety and effectiveness of the product on the skin [43]. These findings agree with other studies, such as that by Tan et al. [44], which reported similar pH stability in creams containing natural extracts, demonstrating that fenugreek extract does not significantly alter the pH of the formulation, maintaining

skin compatibility. The viscosity of the cream was monitored with values ranging from 7941.69 to 7956.70 cp over the 90 days (Table 7). This stability in viscosity indicates that the cream preserved its texture and spreadability, key characteristics for consumer satisfaction and effective application. Okafo et al. [45] also reported similar viscosity stability in *Psidium guajava* ethanol extract creams formulated with plant extracts, further supporting the reliability of our results in maintaining product consistency. Color analysis revealed minor changes, with a slight shift toward yellow (b^* values increasing from 4.3 to 4.8) (Table 7). Centrifuge tests were conducted at 4 °C, 25 °C, and 40 °C to evaluate the cream's homogeneity. The cream remained stable under all conditions, with no separation or phase inversion observed. These results are consistent with findings from Chaabani et al. [17], where creams containing *Cymodocea nodosa* extract demonstrated similar stability, with no significant phase separation under various temperature conditions. This suggests that the fenugreek eco-extract does not compromise the physical stability of the cream.

Table 7. Stability parameters of the cream.

	Day 0	Day 30	Day 60	Day 90
pH	6.34	6.25	6.32	6.35
Viscosity (cp)	7941.69	7956.70	7967.68	7973.62
Z-average (d.nm)	321.20 ^a ± 5.32	332.79 ^a ± 9.05	339.11 ^a ± 18.07	320.60 ^a ± 2.18
Zeta potential (mV)	−37.25 ^a ± 1.45	−36.49 ^a ± 2.16	−36.18 ^a ± 2.67	−36.70 ^a ± 3.86
L*	82.5	82.5	82.5	82.5
a*	−2.4	−2.4	−2.5	−2.6
b*	4.3	4.7	4.4	4.8
Color difference ΔE	-	0.14	0.14	0.28
Centrifuge stability 4 °C	Stable	Stable	Stable	Stable
Centrifuge stability 25 °C	Stable	Stable	Stable	Stable
Centrifuge stability 40 °C	Stable	Stable	Stable	Stable

A one-way ANOVA followed by Duncan's multiple range test was applied, revealing that values marked with same superscript letters (a) were significantly non different at a significance level of $p < 0.05$.

The zeta potential (ZP) of the cream was measured to evaluate its colloidal stability, with values ranging from −36.18 to 37.25 mV. A negative ZP indicates good stability by preventing aggregation of particles in the formulation.

In conclusion, the cosmetic cream containing fenugreek eco-extract demonstrated excellent stability over 90 days, maintaining its physical properties, color, and consistency under various storage conditions. These findings align with previous studies on the stability of plant-based cosmetic formulations and highlight the potential of fenugreek extract as a safe, effective, and sustainable ingredient for next-generation skincare products.

3.12. Evaluation of Sensory Characteristics of the Cosmetic Cream

The sensory properties of the cosmetic cream formulated with fenugreek eco-extract were evaluated by a panel of 60 participants to assess key attributes. Each parameter was rated on a 10-point scale, and an overall score was calculated based on participant feedback. The results obtained are highly encouraging, as illustrated by the radar graph (Figure 4). The cream received consistently high scores across all attributes, demonstrating strong consumer appeal. The color was rated at an average of 9.38, indicating that the cream's natural appearance was well received. The fragrance scored 9.09, reflecting the pleasant and subtle aroma of the formulation, aligning with preferences for mild fragrances in skincare products. Texture-related attributes also performed exceptionally well, with consistency scoring at 9.41, spreadability at 9.6, and stickiness at 9.8, highlighting the cream's smooth

application and non-greasy finish. As well, absorption was rated the highest, with an average score of 9.81, showing the cream's ability to penetrate the skin effectively without leaving a residue (Figure 5).

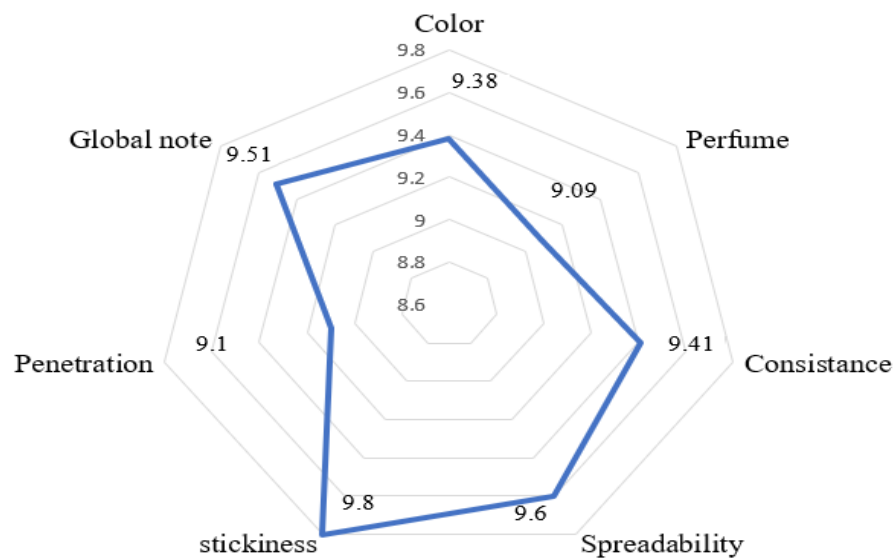


Figure 5. Sensory analysis of *T. foenum-graecum* L. cosmetic cream.

These results agree with previous studies on natural cosmetic formulations. For instance, Chaabani et al. [17] reported similar consumer satisfaction scores for creams enriched with plant-based extracts, emphasizing the importance of ease of application and lightweight textures.

Eventually, the overall score of 9.51 highlights the cream's ability to meet user expectations, demonstrating strong sensory appeal and functional benefits. Combined with its proven stability and antioxidant, anti-inflammatory, and antimicrobial properties, the fenugreek eco-extract-based cosmetic cream shows great promise as a versatile ingredient for innovative and commercially viable skincare products.

4. Conclusions

In summary, this study successfully optimized the ultrasound-assisted extraction (UAE) process for fenugreek seeds, enabling the efficient recovery of bioactive compounds with confirmed antioxidant, antimicrobial, and anti-inflammatory properties. Chromatographic analysis identified phenolic compounds known for their anti-aging benefits and protective effects against oxidative damage. The formulated cream exhibited remarkable stability across various environmental conditions, maintaining its physical integrity and sensory attributes. These results underscore the potential of fenugreek seeds as a sustainable and multifunctional ingredient for eco-friendly cosmetic formulations, offering diverse benefits for skin health. By integrating advanced green extraction techniques with plant-based actives, this study sets the stage for next-generation skincare products that prioritize efficacy and environmental sustainability, addressing the growing consumer demand for innovative and responsible solutions in the cosmetics industry.

Despite these promising findings, several limitations must be acknowledged. The study primarily relied on *in vitro* methods, which may not fully reflect the performance and stability of fenugreek extracts in practical cosmetic applications. Additionally, the focus on specific fenugreek extracts could limit the scope of findings, as results may vary with different plant sources or extraction methods. Finally, the controlled laboratory conditions and small-scale nature of the research may reduce the applicability of the results to large-scale industrial formulations. Addressing these limitations through further studies,

including in vivo testing and scaling up production, could enhance the relevance of these findings for commercial applications.

Author Contributions: Conceptualization, A.B.A. and I.B.R.; methodology, A.B.A., E.C., S.B., M.H., I.B.R. and O.M.; software, A.B.A., E.C. and M.H.; validation, A.B.A., E.C., S.B., M.H. and I.B.R.; formal analysis, A.B.A., E.C., S.B. and I.B.R.; investigation, A.B.A.; resources, M.S.T., I.B.R. and O.M.; data curation, A.B.A., E.C., S.B., M.H., M.S.T., I.B.R. and O.M.; writing—original draft preparation, A.B.A., E.C., S.B. and I.B.R.; writing—review and editing, M.S.T., I.B.R. and O.M.; visualization, A.B.A., E.C., M.H. and I.B.R.; supervision, M.S.T. and I.B.R.; project administration, I.B.R.; funding acquisition, M.S.T., I.B.R. and O.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflicts of interest.

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